Centric Environmental Solutions, LLC

"Healthy Homes & Buildings for Quality of Life"

August 14, 2026

Collin Boozer, Project Manager
SERVPRO WFI, Inc.
121 Falls Avenue
Granite Falls, North Carolina 28630

Re: Letter Report for Limited Microbial Assessment & IAQ Sampling Services
Coddle Creek Elementary School
141 Franks Crossing Loop, Mooresville, North Carolina
Job# CES779

Dear Mr. Boozer:

This letter report summarizes the limited microbial assessment and sampling performed by Centric Environmental Solutions, LLC. ("CES") at the above referenced elementary school building ("SITE").

Mr. James T. Dallas, a Certified Indoor Environmental Consultant ("CIEC") performed the microbial assessment and sampling.

Executive Summary Limited Microbial Assessment & IAQ Sampling

On August 11th, 2026 CES mobilized to the Coddle Creek Elementary School located at 141 Franks Crossing Loop in Mooresville, North Carolina as approved by the Client (SERVPRO WFI, Inc.).

The purpose of the limited microbial assessment and sampling was to evaluate and measure reported "dampness" inside the school building and to address concerns for any visible microbial conditions that may be present in common hallway areas and classrooms.

On the day of the assessment, CES was accompanied by representatives of SERVPRO. The school was open for teachers and staff only. No children were present during CES's inspection.

Prior to any indoor air sampling, CES calibrated a Bio-pump to approximately 15-liters per minute.

The limited microbial inspection and sampling were conducted in random classrooms, access hallway areas, three (3) classroom hallways (green, yellow and blue), inside classroom D122, a cafeteria and a gymnasium ("random indoor areas of concern"), where suspected visible microbial concerns were identified by CES and where elevated relative humidity levels were measured by CES.

Suspected visible microbial conditions were generally identified on hallway and classroom ceiling diffusers and return vents.

No intrusive sampling, or intrusive investigations were performed by CES. The following pages of the report provide detail of CES's microbial assessment, measurements and sampling:

Common Hallways and Classrooms ("areas of concern")

Blue/Yellow Access Hallway

Suspected visible microbial conditions were identified on ceiling diffusers and return vents in the blue/yellow access hallway.

CES collected a non-viable indoor air sample (42439659) from the blue/yellow access hallway to determine in any elevated mold spore conditions are present.

Blue Classroom Hallway (outside Classrooms D111 and D130)

The temperature of the blue hallway area measured approximately 73.1°F and the relative humidity measured ranging from **65.1%** to **67.1%.**

The relative humidity levels measured in the blue hallway are **elevated** (ANSI/ASHRAE 62.1).

CES recommends that relative humidity levels indoors be maintained between 30% and 55%.

Suspected visible microbial conditions were identified on ceiling diffusers and return vents in the blue classroom hallway.

CES collected a non-viable indoor air sample (42439646) from the blue classroom hallway to determine in any elevated mold spore conditions are present.

Blue Classroom D122

Suspected light microbial/settled dust conditions were observed classroom desks.

Suspected visible microbial conditions were identified on ceiling diffusers and return vents in classroom D122.

The temperature of classroom D122 measured approximately 72.6°F and the relative humidity measured ranging from **72.4%** to **76.1%.**

The relative humidity levels measured in classroom D122 are **elevated** (ANSI/ASHRAE 62.1).

CES recommends that relative humidity levels indoors be maintained between 30% and 55%.

CES collected a non-viable indoor air sample (42439639) from classroom D122 to determine in any elevated mold spore conditions are present.

Blue Classroom D125

Suspected light microbial conditions were observed on a red book bag. *CES recommends that the book bag be washed, or discarded.*

Suspected visible microbial conditions were identified on ceiling diffusers and return vents in classroom D125.

Yellow/Green Access Hallway

Suspected visible microbial conditions were identified on ceiling diffusers and return vents in the yellow/green access hallway.

CES collected a non-viable indoor air sample (42439652) from the yellow/green access hallway to determine in any elevated mold spore conditions are present.

Yellow Classroom Hallway (outside Classrooms C138 and C145)

The temperature of the yellow hallway area measured approximately 71.4°F and the relative humidity measured ranging from **65.5%** to **66.6%.**

The relative humidity levels measured in the yellow hallway are **elevated** (ANSI/ASHRAE 62.1).

CES recommends that relative humidity levels indoors be maintained between 30% and 55%.

Suspected visible microbial conditions were identified on ceiling diffusers and return vents in the yellow classroom hallway.

CES collected a non-viable indoor air sample (42440762) from the yellow classroom hallway to determine in any elevated mold spore conditions are present.

Yellow Classrooms C140 and C145

Suspected visible microbial conditions were identified on ceiling diffusers and return vents in classrooms C140 and C145.

Green Classroom Hallway (outside Classrooms B120 and B133)

The temperature of the green hallway area measured approximately 72.2°F and the relative humidity measured ranging from **61.9%** to **65.2%.**

The relative humidity levels measured in the yellow hallway are **elevated** (ANSI/ASHRAE 62.1).

CES recommends that relative humidity levels indoors be maintained between 30% and 55%.

Suspected visible microbial conditions were identified on ceiling diffusers and return vents in the green classroom hallway.

CES collected a non-viable indoor air sample (42439655) from the green classroom hallway to determine in any elevated mold spore conditions are present.

Green Classroom B128

Suspected visible microbial conditions were identified on ceiling diffusers and return vents in classroom B128.

Gymnasium

Suspected visible microbial conditions were identified on wall return vents.

The temperature of the gymnasium measured approximately 70.5°F and the relative humidity measured approximately **67.2%.**

The relative humidity levels measured in the gymnasium are **elevated** (ANSI/ASHRAE 62.1).

CES recommends that relative humidity levels indoors be maintained between 30% and 55%.

CES collected a non-viable indoor air sample (42439660) from the gymnasium to determine in any elevated mold spore conditions are present.

Cafeteria/Stage

Suspected visible microbial conditions were identified on ceiling tiles in the stage area.

The temperature of the cafeteria measured approximately 70.2°F and the relative humidity measured approximately **71.4%.**

The relative humidity levels measured in the cafeteria are **elevated** (ANSI/ASHRAE 62.1).

CES recommends that relative humidity levels indoors be maintained between 30% and 55%.

CES collected a non-viable indoor air sample (42440759) from the cafeteria at the stage area to determine in any elevated mold spore conditions are present.

Janitor Closets (Cafeteria)

CES observed staining/visible microbial conditions identified on ceiling tiles inside two (2) janitor closets.

A janitor closet that contains a hot water heater was observed to contain active-leaking water valves at wall faucet fixtures into a tub wash basin below.

CES recommends that a licensed plumber fix the leaking water valves. The stained ceiling tiles should be removed and replaced by building maintenance personnel.

Outdoor Environment

CES also collected an outdoor air sample (42440760) for reference purposes. The outdoor environmental was hazy, humid with light wind conditions on the day of the sampling.

The outdoor temperature measured approximately 85.7°F and the relative humidity measured approximately 65.6%.

Sampling Results

A total of nine (9) non-viable air samples were shipped overnight to EMSL Analytical, Inc. located in Pineville, NC for laboratory analysis.

The samples were analyzed for Total Fungal Spore counts using optical microscopy using an expedited laboratory turnaround time (TAT) of 24-hours, as requested.

The results obtained from the limited mold assessment and sampling are not intended to infer if any health concerns are present, rather the results will be utilized to document existing mold conditions and if mold remediation will be required by a Mold Remediation Contractor, or if any HVAC system cleaning is required, or if any additional building investigation is required.

The results of the indoor air sampling for the Coddle Creek Elementary School are summarized in Tables 1 and 2:

Table 1: August 11th, 2026 Indoor Air Sampling Event

Spore Types	Blue/Yellow Access Hallway	Blue Hallway	Classroom D122	Yellow/Green Access Hallway	Yellow Hallway	Outdoor Reference
Ascospores	ND	ND	ND	40	ND	200
Asp/Pen-spores	***10,800***	200	***990***	***1,200***	***1,100***	ND
Basidiospores	40	40	ND	80	100	14,500
Cladosporium	ND	ND	ND	ND	ND	820
Curvularia	ND	ND	ND	ND	80	80
Fusarium	ND	ND	ND	ND	ND	200
Cercospora	ND	ND	ND	ND	ND	40
Nigrospora	ND	ND	ND	ND	40	40
Pithomyces	ND	ND	ND	ND	40	ND
Myxomycetes	ND	ND	ND	ND	40	ND
Polythrincium	ND	ND	ND	ND	ND	40
Sporidesmium	ND	ND	ND	ND	ND	40
Zygomycetes	ND	40	ND	ND	ND	ND
Total Fungi	**10,840**	**280**	**990**	**1,320**	**1,400**	**15,960**

Results are reported as counts per meters cubed air (counts/m^3).
ND – Denotes "Non-Detect"

Table 2: August 11th, 2026 Indoor Air Sampling Event				
Spore Types	**Green Hallway**	**Gymnasium**	**Cafeteria**	**Outdoor Reference**
Ascospores	ND	ND	40	200
Asp/Pen-spores	***8,210***	30	80	ND
Basidiospores	80	2,300	2,700	14,500
Cladosporium	ND	ND	ND	820
Curvularia	ND	ND	ND	80
Epicoccum	40	ND	ND	ND
Fusarium	BD	ND	ND	200
Cercospora	ND	ND	ND	40
Nigrospora	ND	ND	ND	40
Polythrincium	ND	ND	ND	40
Sporidesmium	ND	ND	ND	40
Total Fungi	**8,330**	**2,330**	**2,820**	**15,960**

Results are reported as counts per meters cubed air (counts/m^3).
ND – Denotes "Non-Detect"

Elevated counts of ***Aspergillus/Penicillium spores*** were detected inside Blue Classroom D122; the Green Classroom Hallway and Yellow Classroom Hallway and both Access Hallways as compared to the outdoor levels of *Aspergillus/Penicillium spores* (non-detect).

The Total Fungal Spore counts detected inside Gymnasium, Cafeteria, Blue Classroom D122; the Classroom Hallways and Access Hallways were **lower** than the outdoor mold spore counts (15,960 counts/m^3).

No elevated spores were detected in the indoor air samples collected from the Blue Hallway, Gymnasium and Cafeteria.

The types of molds detected in the indoor air samples are commonly associated with moisture conditions (i.e., elevated relative humidity levels) measured by CES in the common hallways, access hallways and classrooms outlined in this report.

Please note that North Carolina currently does not have any promulgated standards for evaluating, or the interpretation of fungi levels in bioaerosol samples or on surfaces. Surface contamination rating is used by the laboratory as referenced in IICRC S520 Standard Reference Guide for Mold Remediation.

Although there are no standards in North Carolina for interpreting mold on building surfaces or in air samples, CES recommends that the levels of Aspergillus/Penicillium mold spores indoors be less than or equal to the outdoor levels of Aspergillus/Penicillium spores.

When levels of Aspergillus/Penicillium spores are absent from the outdoor environment, CES recommends that the levels of Aspergillus/Penicillium spores indoors be equal to, or less than 300 counts/m^3.

A complete copy of the laboratory report is provided in Attachment 1 of this report. A building floor plan is provided in Attachment 2.

Conclusions & Recommendations

Based on the observations made during the assessment and the results of the sampling, CES presents the following conclusions and recommendations:

- Elevated counts of *Aspergillus/Penicillium spores* were detected inside Blue Classroom D122; the Green Classroom Hallway and Yellow Classroom Hallway and both Access Hallways as compared to the outdoor levels of *Aspergillus/Penicillium spores* (non-detect).
- Visible suspect microbial conditions were identified on Hallway ceiling diffusers/return vents and inside each Classroom ceiling diffusers/return vents and on Gymnasium wall return vents.
- Elevated relative humidity levels were measured by CES in Classrooms, Hallways, Gymnasium and Cafeteria as outlined in this report.
- CES also observed evidence of stained ceiling tiles inside Janitor Closets and active-leaking water valves. *CES recommends that a licensed plumber fix the leaking water valves identified by CES.*
- Several stained ceiling tiles were observed inside Janitor Closets. *ANY mold/moisture impacted ceiling tiles should be removed and replaced.*
- Based on CES's field observations and the results of the mold sampling, CES recommends that mold remediation and air scrubbing be conducted inside ALL Classrooms, Classroom Hallway areas and Access Hallway areas, to include air scrubbing, dehumidification and cleaning of ALL ceiling diffusers/return vents where suspected visible microbial conditions were identified by CES.
- In addition, CES recommends that that the HVAC system including exterior coils, exterior ductwork areas, exposed exterior sheet metal components and any interior ductwork be professionally cleaned and sanitized by a NADCA-certified HVAC Contractor to remove any potential settled dust and settled mold spores.
- CES also recommends that the HVAC Contractor evaluate and correct the elevated indoor humidity levels measured by CES, from recurring especially during humid summer months.

NOTE: CES recommends that the mold remediation outlined in this report be conducted during a non-vacancy period of approximately 10-14 consecutive work days.

The estimated time-line of mold remediation completion will be verified by the Mold Remediation Contractor.

Scope of Work for Mold Remediation Guidelines

CES recommends that the following general guidelines be followed in addition to ANSI/IICRC S520 Standard for Professional Mold Remediation guidelines to include, but not be limited to:

Containment Barriers & Equipment

Hallway Bathrooms ("Non-Work Areas")

1. Install containment barriers at EACH of the door entrances where the Boys and Girls Bathrooms are located with zipper-flaps to isolate each "non-work area" and to allow for ingress and egress.

Hallways/Classrooms ("Work Areas")

1. Install containment barriers (floor to ceiling) at EACH of the Access Hallway areas and at EACH Classroom Hallway entrance with zipper-flaps to isolate each Hallway area from Classrooms.
2. Install a containment barrier at EACH CLASSROOM door entrance with zipper-flaps to isolate EACH Classroom "work areas".
3. Install at least two (2) dehumidifier units in EACH of the Classroom Hallway areas and two (2) dehumidifier units in EACH Access Hallway area to help reduce the elevated relative humidity levels measured by CES and to control relative humidity levels during remediation activities.
4. Install one (1) dehumidifier unit in EACH of the Classrooms located in the Green, Blue and Yellow Hallway areas to help reduce the elevated relative humidity levels measured by CES and to help reduce the elevated relative humidity levels measured by CES and to control relative humidity levels during remediation activities.
5. Install one (1) HEPA-AFD unit inside the EACH Classroom located in the Green, Blue and Yellow Hallway areas "work areas" with venting through a classroom window to maintain negative-air conditions to remove any elevated mold spores detected by CES any potential elevated mold spores mitigated during mold cleaning activities.
6. Install two (2) HEPA-AFD units with open venting at EACH Classroom Hallway area and two (2) HEPA-AFD units with open venting in EACH Access Hallway area to remove the elevated mold spores detected by CES.

NOTE: CES recommends HEPA filter changes every 3-4 days of continuous use to allow sufficient removal of elevated mold spores.

Cleaning Procedures in ALL Classrooms, Access Hallways and Classroom Hallways "Work Areas" and in "Non-Work Areas"

1. HEPA vacuum ALL ceiling diffusers, return vents and flooring in ALL Classrooms located in the Green, Blue and Yellow Hallways to remove any visible mold/settled mold spores.
2. HEPA vacuum ALL ceiling diffusers, return vents and flooring located in ALL Access Hallway areas and in the Green, Blue and Yellow Hallways to remove any visible mold/settled mold spores.
3. As a precautionary measure, HEPA vacuum ALL Blue classroom contents (desks, chairs, etc.) and furnishings, as applicable.
4. Damp-wipe ALL ceiling diffusers, return vents and flooring in ALL Classrooms located in the Green, Blue and Yellow Hallways using an EPA mold disinfectant.
5. Damp-wipe ALL ceiling diffusers, return vents and flooring in ALL Access Hallway areas and in the Green, Blue and Yellow Hallway areas using an EPA mold disinfectant.
6. As a precautionary measure, clean/damp-wipe ALL classroom contents (desks, chairs, etc.) and furnishings, as applicable using an EPA mold disinfectant.
7. Damp-wipe ALL Classroom flooring areas located in the Access Hallway areas and in the Green, Blue and Yellow Hallways using an EPA mold disinfectant.
8. As a precautionary measure, damp-wipe horizontal countertops and flooring areas located inside the Boys Bathroom and Girls Bathroom using an EPA mold disinfectant to remove any potential settled mold spores.
9. Re-HEPA vacuum ALL cleaned areas inside ALL Classrooms, Access Hallways and Classroom Hallway "work areas" to remove any potential settled mold spores.
10. Allow HEPA-AFD units and dehumidifier units to run continuously for approximately 96-hours upon completion of ALL mold remediation activities inside EACH of the "work areas".
11. Keep ALL containment barriers intact and equipment operational inside EACH of the "work areas" until such time mold remediation has been completed by the Mold Remediation Contractor and CES has been notified of completion to determine IF post-mold remediation verification sampling will be required by the Client.

Post-Mold Remediation Recommendations

- CES recommends that that the HVAC system including exterior coils, exterior ductwork foil insulation areas, exposed exterior sheet metal components and any interior ductwork be professionally cleaned and sanitized by a NADCA-certified HVAC Contractor.
- CES also recommends that the HVAC Contractor evaluate and correct the elevated indoor humidity levels measured by CES, from recurring especially during humid summer months.
- CES recommends that a post-mold remediation verification (PMRV) visual assessment be conducted to verify the efficacy of the mold remediation and to provide PMRV air sampling to determine air quality clearance for re-occupancy of the Classrooms/Hallways "work areas".
- If requested, CES will provide a change-order price estimate to return to the Site to conduct the PMRV assessment and sampling.

If you have any questions regarding this letter report, please call me to discuss at 732-682-6639. Thank you for the opportunity to assist you in this regard.

For Centric Environmental Solutions, LLC.



James T. Dallas, CIEC

President



ATTACHMENT 1

Laboratory Report

OrderID: 412606013

Microbiology Chain of Custody Form

EMSL Order Number / Lab Use Only: 412606013

EMSL Analytical, Inc.
10801 Southern Loop Blvd.
Pineville, NC 28134

PHONE: (704) 525-2205
EMAIL: CharlotteLab@emsl.com

EMSL ANALYTICAL INC.
TESTING LABS • PRODUCTS • TRAINING

If Bill-To is the same as Report-To leave this section blank. Third-party billing requires written authorization.

Customer Information		Billing Information	
Customer ID:		Billing ID:	
Company Name:	Centric Environmental Solutions, LLC.	Company Name:	Same
Contact Name:	Jim Dallas	Billing Contact:	Same
Street Address:	1325 Scotts Creek Road	Street Address:	Same
City, State, Zip:	Statesville, NC 28625	City, State, Zip:	Same
Country:	USA	Country:	USA
Phone:	732-682-6639	Phone:	Same
Email(s) for Report:	Jim@call centrices.com	Email(s) for Invoice:	Same

Project Information

Project Name/No.: CES779

Purchase Order: P779

EMSL LIMS Project ID: (If applicable, EMSL will provide)

State Samples Collected: NC

Zip Code Samples Collected: 28115

State of Connecticut (CT) must select project location: ☐ Commercial (Taxable) ☐ Residential (Non-taxable)

Sampled By Name: JAMES T. DALLAS, CIEC

Sampled By Signature: James T. Dallas

No. of Samples in Shipment: (9)

Sterile, Sodium Thiosulfate Preserved Bottle Used: ☐ Biocide Used in Source (specify)

Public Water Supply Samples: ☐ Note: All results may automatically be reported to DOH if required by State.

Turn-Around-Time (TAT) Please call ahead for large projects and/or compressed times 8 Hours or Less. *32 Hour TAT available for select tests only; samples must be submitted by 11:30am.

☐ 3 Hour ☐ 6 Hour ☒ 24 Hour ☐ 32* Hour ☐ 48 Hour ☐ 72 Hour ☐ 96 Hour ☐ 1 Week ☐ 2 Week

MICROBIOLOGY TEST CODES

M001 Air-O-Cell	M174 MoldSnap	M012 *Pseudomonas aeruginosa* (P/A***)
M030 MICRO 5	M032 Allergenco-D	M024 *Pseudomonas aeruginosa* (MFT*)
M041 Fungal Direct Examination		M015 Heterotrophic Plate Count
M159 Pollen ID & Enumeration		M017 Total Coliform & *E. Coli* (Colilert P/A***)
M280 Dust Characterization Level-1		M018 Total Coliform & *E. Coli* (MFT*)
M281 Dust Characterization Level-2		M114 Total Coliform & *E. Coli Enumeration* (Colilert MPN**)
M005 Viable Fungi-Air Samples (Genus ID & Count)		M019 Fecal Coliform (MFT*)
M006 Viable Fungi-Air Samples (Includes *Penicillium, Aspergillus, Cladosporium, Stachybotrys* Species ID & Count)		M020 *Fecal Streptococcus* (MFT*)
M007 Culturable Fungi-Surface Samples (Genus ID & Count)		M029 *Enterococci* (MFT*)
M008 Culturable Fungi-Surface Samples (Includes *Penicillium, Aspergillus, Cladosporium, Stachybotrys* Species ID & Count)		M129 *Enterococci* (Enterolert P/A***)
M009 Bacteria Culture Gram Stain & Count		M160 Real Time qPCR-ERMI 36 Panel
M010 Bacteria Count & ID – 3 Most Prominent		M025 Sewage Screen – Water (MFT*)
M011 Bacteria Count & ID – 5 Most Prominent		

M115 Sewage Screen – Water (P/A***)	
M116 Sewage Screen – Water (MPN**)	
M117 Sewage Screen – Swab (P/A***)	
M013 Sewage Screen – Swab (MFT*)	
M730 *Methicillin-resistant Staph. aureus* (MRSA)	
M031 Rapid-growing non-TB *Mycobacteria* Detection & Enumeration	
M014 Endotoxin Analysis	
M044 Group Allergen (Cat, Dog, Cockroach, Dust Mite)	
M095 Bacteroides	
Other – See Analytical Price Guide for Test Code	
Legionella Analysis Please use EMSL *Legionella* COC	

*MFT = Membrane Filtration Technique
**MPN = Most Probable Number
***P/A = Presence/Absence

Sample #	Sample Location/Description	Sample Type (Matrix)	Potable / Non-Potable (Only for Water)	Test Code	Volume/Area	Date / Time Collected	Temperature (Lab Use Only)
Example: Sample 1	Kitchen	Water	Potable	M017	1,000 ml	1/1/2021 3:30pm	
4243 9660	GYM	AIR		M001	75 L.	8-11-26 10:17	
4244 0759	CAFETERIA					1024	
4243 9652	ACCESS HALL (G)					1031	
4243 9655	GREEN HALLWAY					1039	
4244 0762	YELLOW HALLWAY					1047	
4243 9659	ACCESS HALL (B)					1055	

Special Instructions and/or Regulatory Requirements (Sample Specifications, Processing Methods, Limits of Detection, etc.)

Method of Shipment: FEDEX

Sample Condition Upon Receipt:

Relinquished by: J. Dallas	Date/Time: 8-11-26 1400	Received by: RC 8/12/26 1200	Date/Time: FX 8756 3082 7684
Relinquished by:	Date/Time:	Received by:	Date/Time

CoC-Rcd Incorrect - COC 34 Micro R13 03/02/2021

☐ AGREE TO ELECTRONIC SIGNATURE (By checking, I consent to signing this Chain of Custody document by electronic signature.)

EMSL Analytical, Inc.'s Laboratory Terms and Conditions are incorporated into this Chain of Custody by reference in their entirety. Submission of samples to EMSL Analytical, Inc. constitutes acceptance and acknowledgment of all terms and conditions by Customer.

OrderID: 412606013

EMSL ANALYTICAL, INC.
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Microbiology Chain of Custody Form

EMSL Order Number / Lab Use Only
412606043

EMSL Analytical, Inc.
10801 Southern Loop Blvd.
Pineville, NC 28134

PHONE: (704) 525-2205
EMAIL: CharlotteLab@emsl.com

Additional Pages of the Chain of Custody are only necessary if needed for additional sample information

Special Instructions and/or Regulatory Requirements (Sample Specifications, Processing Methods, Limits of Detection, etc.)

24-HR. TAT

Sample #	Sample Location/Description	Sample Type (Matrix)	Potable / Non-Potable (Only for Water)	Test Code	Volume/Area	Date / Time Collected	Temperature (Lab Use Only)
4243 9646	BLUE HALLWAY	AIR		M001	75 L.	8-11-26 1103	
4243 9639	CLASSROOM D122	↓		↓	↓	↓ 1114	
4244 0760	OUTDOOR REFERENCE	↓		↓	↓	↓ 1123	

Method of Shipment: FEDEX		Sample Condition Upon Receipt:	
Relinquished by: J. Dillon	Date/Time: 8-11-26 1400	Received by:	Date/Time
Relinquished by:	Date/Time:	Received by:	Date/Time

Controlled Document - COC-34 Micro R13 3.02.2021

☐ AGREE TO ELECTRONIC SIGNATURE (By checking, I consent to signing this Chain of Custody document by electronic signature.)

EMSL Analytical, Inc.'s Laboratory Terms and Conditions are incorporated into this Chain of Custody by reference in their entirety. Submission of samples to EMSL Analytical, Inc. constitutes acceptance and acknowledgment of all terms and conditions by Customer.



EXPANDED FUNGAL REPORT ™

Prepared Exclusively For

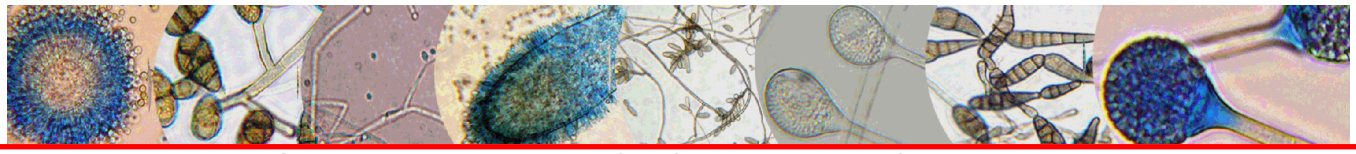
Centric Environmental Solutions, LLC

1325 Scotts Creek Road
Statesville, NC 28625
Phone:732-682-6639

Report Date:	**8/13/2026**
Project:	**CES779**
P.O:	**P779**
EMSL Order:	**412606013**

AIHA LAP, LLC.

AIHA LAP, LLCEMLAP #192283



This report has been prepared by EMSL Analytical, Inc. at the request of and for the exclusive use of the client named in this report. Completely read the important terms, conditions, and limitations that apply to this report.

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EMSL Analytical, Inc.
10801 Southern Loop Blvd Pineville, NC 28134
Phone: (704) 525-2205 Fax: (704) 525-2382 Web: http://www.EMSL.com Email:charlottelab@emsl.com

Attn: Jim Dallas
Centric Environmental Solutions, LLC
1325 Scotts Creek Road
Statesville, NC 28625

EMSL Order:	412606013
Customer ID:	CENT75
Collected:	8/11/2026
Received:	8/12/2026
Analyzed:	8/13/2026

Proj: CES779

1. Description of Analysis

Analytical Laboratory

EMSL Analytical, Inc. (EMSL) is a nationwide, full service, analytical testing laboratory network providing Asbestos, Mold, Indoor Air Quality, Microbiological, Environmental, Chemical, Forensic, Materials, Industrial Hygiene and Mechanical Testing services since 1981. Ranked as the premier independently owned environmental testing laboratory in the nation, EMSL puts analytical quality as its top priority. This quality is recognized by many well-respected federal, state and private accrediting agencies, and assured by our high quality personnel, including many Ph.D. microbiologists and mycologists.

EMSL is an independent laboratory that performed the analysis of these samples. EMSL did not conduct the sampling or site investigation for this report. The samples referenced herein were analyzed under strict quality control procedures using state-of-the-art microbiological methods. The analytical methods used and the data presented are scientifically and legally defensible.

The laboratory data is provided in compliance with ISO-IEC 17025 guidelines for the particular test(s) requested, including any associated limitations for the methods employed. These data are intended for use by professionals having knowledge of the testing methods necessary to interpret them accurately.

This report has been prepared by EMSL Analytical, Inc. at the request of and for the exclusive use of the client named in this report. Completely read the important terms, conditions, and limitations that apply to this report.
© 2006,EMSL Analytical, Inc., All rights reserved. No part of this report may be reproduced or otherwise distributed or used without the express written consent of EMSL.

EMSL Analytical, Inc.
10801 Southern Loop Blvd Pineville, NC 28134
Phone: (704) 525-2205 Fax: (704) 525-2382 Web: http://www.EMSL.com Email:charlottelab@emsl.com

Attn: Jim Dallas
Centric Environmental Solutions, LLC
1325 Scotts Creek Road
Statesville, NC 28625

EMSL Order: 412606013
Customer ID: CENT75
Collected: 8/11/2026
Received: 8/12/2026
Analyzed: 8/13/2026

Proj: CES779

Air Samples - Spore traps:

Spore traps are commercially available sampling devices that capture airborne particles on an adhesive slide. Air is pulled through the device using a vacuum pump. Spores, as well as other airborne particles, are impacted on the collection adhesive. Using spore trap collection methods has inherent limitations. These collection methods are biased towards larger spore sizes.

The analysis for total spore counts is a direct microscopic examination and does not include culturing or growing the fungi. Therefore, the results include both viable and non-viable spores. Some fungal groups produce similar spore types that cannot be distinguished by direct microscopic examination alone (i.e., *Aspergillus/Penicillium*, and others). Other spore types may lack distinguishing features that aid in their identification. These types are grouped into larger categories such as Ascospores or Basidiospores.

Fungal spores are identified and grouped by morphological characteristics including color, shape, septation, ornamentation, and fruiting structures (if present) which are compared to published mycological identification keys and texts. EMSL reports provide spore counts per cubic meter of air to three significant figures. Please note that each spore category is reported to three significant figures. Due to rounding and the application of three significant figures the sum of the individual spore numbers may not equal the total spore count on the report. EMSL does not maintain responsibility for final volume concentrations (counts/m3) since this volume is provided by the field collector and can not be verified by EMSL.

EMSL analyzes spore traps using phase contrast microscopy. There is a wide choice of collection devices (Air-O-Cell, Micro-5, Burkhard, etc.) on the market. Differences in analytical method may exist between spore trap devices.

Spore trap results are reported in spores per cubic meter of air. Due to the other airborne particles collected with the spores, EMSL reports a background particle density. Background density is an indication of overall particulate matter present on the sample (i.e. dust in the air). High background concentrations may obscure spores such as the *Penicillium*/*Aspergillus* group. The rating system is from 1-5 with 1 = 1 - 25% of the background obscured by material, 2 = 26 - 50%, 3 = 51 - 75%, 4 = 76% - 99%, 5 = 100% or overloaded. A background rating of 4 or higher should be regarded as a minimum count since the actual concentrations may be higher than those reported. EMSL will not be held responsible for overloading of samples. Sample volumes are left to the discretion of the company or persons conducting the fieldwork.

Skin fragment density is the percentage of skin cells making up the total background material, 1 = 1 - 25%, 2 = 26 - 50%, 3 = 51 - 75%, 4 = 76-100%. Skin fragment density is considered an indication of the general cleanliness in the area sampled. It has been estimated that up to 90% of household dust consists of dead skin cells.

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Attn: Jim Dallas
Centric Environmental Solutions, LLC
1325 Scotts Creek Road
Statesville, NC 28625

Proj: CES779

EMSL Order:	412606013
Customer ID:	CENT75
Collected:	8/11/2026
Received:	8/12/2026
Analyzed:	8/13/2026

2. Analytical Results

See attached data reports and charts.

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Spore Trap ASSESSMENTReport™ Air-O-Cell(™) Analysis of Fungal Spores & Particulates (Methods MICRO-SOP-201, ASTM D7391)

	Particle Identification	Raw Count	(Count/m³)	% of Total	Interpretation Guideline
412606013-0001	Alternaria (Ulocladium)	-	-	-	
	Ascospores	-	-	-	
Client Sample ID	Aspergillus/Penicillium++	2*	30*	1.3	Slightly Elevated
42439660	Basidiospores	56	2300	98.7	Acceptable
	Bipolaris++	-	-	-	
	Chaetomium++	-	-	-	
Location	Cladosporium	-	-	-	
Gym	Curvularia	-	-	-	
	Epicoccum	-	-	-	
Sample Volume (L)	Fusarium++	-	-	-	
	Ganoderma	-	-	-	
75	Myxomycetes++	-	-	-	
	Pithomyces++	-	-	-	
Sample Type	Rust	-	-	-	
	Scopulariopsis/Microascus	-	-	-	
Inside	Stachybotrys/Memnoniella	-	-	-	
Comments	Zygomycetes	-	-	-	
	Cercospora++	-	-	-	
	Nigrospora	-	-	-	
	Polythrincium	-	-	-	
	Sporidesmium++	-	-	-	
	Total Fungi	**58**	**2330**	**100**	Acceptable
	Hyphal Fragment	1*	10*	-	Slightly Elevated
	Insect Fragment	-	-	-	
	Pollen	-	-	-	

Analytical Sensitivity 600x: **41** counts/cubic meter
Analytical Sensitivity 300x *: **13*** counts/cubic meter

Skin Fragments: **3** 1 to 4 (low to high)
Fibrous Particulate: **1** 1 to 4 (low to high)
Background: **2** 1 to 4 (low to high); 5 (overloaded)

Acceptable Concentration at or below background
Slightly Elevated Concentration above background
ELEVATED Concentration 10X or more above background

Not commonly found growing indoors, spores likely come from outside.
Spores reported to be able to cause allergies in individuals.
Potential for mycotoxin production exists with these fungi.
These fungi are considered water damage indicators.

++ Includes other spores with similar morphology; see EMSL's fungal glossary for each specific category.
-

Initial report from: 08/13/2026 09:01:31

Lee Plumley, Laboratory Manager
or Other Approved Signatory

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EMSL Order: 412606013
Customer ID: CENT75
Collected: 8/11/2026
Received: 8/12/2026
Analyzed: 8/13/2026

Proj: CES779

Spore Trap ASSESSMENTReport™ Air-O-Cell(™) Analysis of Fungal Spores & Particulates (Methods MICRO-SOP-201, ASTM D7391)

	Particle Identification	Raw Count	(Count/m³)	% of Total	Interpretation Guideline
412606013-0002	Alternaria (Ulocladium)	-	-	-	
	Ascospores	1	40	1.4	Acceptable
Client Sample ID	Aspergillus/Penicillium++	2	80	2.8	Slightly Elevated
42440759	Basidiospores	67	2700	95.7	Acceptable
	Bipolaris++	-	-	-	
	Chaetomium++	-	-	-	
Location	Cladosporium	-	-	-	
Cafeteria	Curvularia	-	-	-	
	Epicoccum	-	-	-	
Sample Volume (L)	Fusarium++	-	-	-	
	Ganoderma	-	-	-	
75	Myxomycetes++	-	-	-	
	Pithomyces++	-	-	-	
Sample Type	Rust	-	-	-	
	Scopulariopsis/Microascus	-	-	-	
Inside	Stachybotrys/Memnoniella	-	-	-	
Comments	Zygomycetes	-	-	-	
	Cercospora++	-	-	-	
	Nigrospora	-	-	-	
	Polythrincium	-	-	-	
	Sporidesmium++	-	-	-	
	Total Fungi	**70**	**2820**	**100**	Acceptable
	Hyphal Fragment	-	-	-	
	Insect Fragment	-	-	-	
	Pollen	-	-	-	

Analytical Sensitivity 600x: **41** counts/cubic meter
Analytical Sensitivity 300x *: **13*** counts/cubic meter

Skin Fragments: **1** 1 to 4 (low to high)
Fibrous Particulate: **1** 1 to 4 (low to high)
Background: **1** 1 to 4 (low to high); 5 (overloaded)

Acceptable Concentration at or below background
Slightly Elevated Concentration above background
ELEVATED Concentration 10X or more above background

Not commonly found growing indoors, spores likely come from outside.
Spores reported to be able to cause allergies in individuals.
Potential for mycotoxin production exists with these fungi.
These fungi are considered water damage indicators.

++ Includes other spores with similar morphology; see EMSL's fungal glossary for each specific category.
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Lee Plumley, Laboratory Manager
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EMSL Order: 412606013
Customer ID: CENT75
Collected: 8/11/2026
Received: 8/12/2026
Analyzed: 8/13/2026

Proj: CES779

Spore Trap ASSESSMENTReport™ Air-O-Cell(™) Analysis of Fungal Spores & Particulates (Methods MICRO-SOP-201, ASTM D7391)

	Particle Identification	Raw Count	(Count/m³)	% of Total	Interpretation Guideline
412606013-0003	Alternaria (Ulocladium)	-	-	-	
	Ascospores	1	40	3	Acceptable
Client Sample ID	Aspergillus/Penicillium++	29	1200	90.9	ELEVATED
42439652	Basidiospores	2	80	6.1	Acceptable
	Bipolaris++	-	-	-	
	Chaetomium++	-	-	-	
Location	Cladosporium	-	-	-	
Access Hall (G)	Curvularia	-	-	-	
	Epicoccum	-	-	-	
Sample Volume (L)	Fusarium++	-	-	-	
	Ganoderma	-	-	-	
75	Myxomycetes++	-	-	-	
	Pithomyces++	-	-	-	
Sample Type	Rust	-	-	-	
	Scopulariopsis/Microascus	-	-	-	
Inside	Stachybotrys/Memnoniella	-	-	-	
Comments	Zygomycetes	-	-	-	
	Cercospora++	-	-	-	
	Nigrospora	-	-	-	
	Polythrincium	-	-	-	
	Sporidesmium++	-	-	-	
	Total Fungi	**32**	**1320**	**100**	Acceptable
	Hyphal Fragment	-	-	-	
	Insect Fragment	-	-	-	
	Pollen	-	-	-	

Analytical Sensitivity 600x: **41** counts/cubic meter
Analytical Sensitivity 300x *: **13*** counts/cubic meter

Skin Fragments: **2** 1 to 4 (low to high)
Fibrous Particulate: **1** 1 to 4 (low to high)
Background: **1** 1 to 4 (low to high); 5 (overloaded)

Acceptable Concentration at or below background
Slightly Elevated Concentration above background
ELEVATED Concentration 10X or more above background

Not commonly found growing indoors, spores likely come from outside.
Spores reported to be able to cause allergies in individuals.
Potential for mycotoxin production exists with these fungi.
These fungi are considered water damage indicators.

++ Includes other spores with similar morphology; see EMSL's fungal glossary for each specific category.
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EMSL Order: 412606013
Customer ID: CENT75
Collected: 8/11/2026
Received: 8/12/2026
Analyzed: 8/13/2026

Proj: CES779

Spore Trap ASSESSMENTReport™ Air-O-Cell(™) Analysis of Fungal Spores & Particulates (Methods MICRO-SOP-201, ASTM D7391)

	Particle Identification	Raw Count	(Count/m³)	% of Total	Interpretation Guideline
412606013-0004	Alternaria (Ulocladium)	-	-	-	
	Ascospores	-	-	-	
Client Sample ID	Aspergillus/Penicillium++	200	8210	98.6	ELEVATED
42439655	Basidiospores	2	80	1	Acceptable
	Bipolaris++	-	-	-	
	Chaetomium++	-	-	-	
Location	Cladosporium	-	-	-	
Green Hallway	Curvularia	-	-	-	
	Epicoccum	1	40	0.5	Slightly Elevated
Sample Volume (L)	Fusarium++	-	-	-	
	Ganoderma	-	-	-	
75	Myxomycetes++	-	-	-	
	Pithomyces++	-	-	-	
Sample Type	Rust	-	-	-	
	Scopulariopsis/Microascus	-	-	-	
Inside	Stachybotrys/Memnoniella	-	-	-	
Comments	Zygomycetes	-	-	-	
	Cercospora++	-	-	-	
	Nigrospora	-	-	-	
	Polythrincium	-	-	-	
	Sporidesmium++	-	-	-	
	Total Fungi	**203**	**8330**	**100**	Acceptable
	Hyphal Fragment	-	-	-	
	Insect Fragment	-	-	-	
	Pollen	-	-	-	

Analytical Sensitivity 600x: **41** counts/cubic meter
Analytical Sensitivity 300x *: **13*** counts/cubic meter

Skin Fragments: **2** 1 to 4 (low to high)
Fibrous Particulate: **1** 1 to 4 (low to high)
Background: **1** 1 to 4 (low to high); 5 (overloaded)

Acceptable Concentration at or below background
Slightly Elevated Concentration above background
ELEVATED Concentration 10X or more above background

Not commonly found growing indoors, spores likely come from outside.
Spores reported to be able to cause allergies in individuals.
Potential for mycotoxin production exists with these fungi.
These fungi are considered water damage indicators.

++ Includes other spores with similar morphology; see EMSL's fungal glossary for each specific category.
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Customer ID: CENT75
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Proj: CES779

Spore Trap ASSESSMENTReport™ Air-O-Cell(™) Analysis of Fungal Spores & Particulates (Methods MICRO-SOP-201, ASTM D7391)

	Particle Identification	Raw Count	(Count/m³)	% of Total	Interpretation Guideline
412606013-0005	Alternaria (Ulocladium)	-	-	-	
	Ascospores	-	-	-	
Client Sample ID	Aspergillus/Penicillium++	28	1100	78.6	ELEVATED
42440762	Basidiospores	3	100	7.1	Acceptable
	Bipolaris++	-	-	-	
	Chaetomium++	-	-	-	
Location	Cladosporium	-	-	-	
Yellow Hallway	Curvularia	2	80	5.7	Acceptable
	Epicoccum	-	-	-	
Sample Volume (L)	Fusarium++	-	-	-	
	Ganoderma	-	-	-	
75	Myxomycetes++	1	40	2.9	Slightly Elevated
	Pithomyces++	1	40	2.9	Slightly Elevated
Sample Type	Rust	-	-	-	
	Scopulariopsis/Microascus	-	-	-	
Inside	Stachybotrys/Memnoniella	-	-	-	
Comments	Zygomycetes	-	-	-	
	Cercospora++	-	-	-	
	Nigrospora	1	40	2.9	Acceptable
	Polythrincium	-	-	-	
	Sporidesmium++	-	-	-	
	Total Fungi	**36**	**1400**	**100**	Acceptable
	Hyphal Fragment	-	-	-	
	Insect Fragment	1	40	-	Slightly Elevated
	Pollen	-	-	-	

Analytical Sensitivity 600x: **41** counts/cubic meter
Analytical Sensitivity 300x *: **13*** counts/cubic meter

Skin Fragments: **2** 1 to 4 (low to high)
Fibrous Particulate: **1** 1 to 4 (low to high)
Background: **2** 1 to 4 (low to high); 5 (overloaded)

Acceptable Concentration at or below background
Slightly Elevated Concentration above background
ELEVATED Concentration 10X or more above background

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Spore Trap ASSESSMENTReport™ Air-O-Cell(™) Analysis of Fungal Spores & Particulates (Methods MICRO-SOP-201, ASTM D7391)

	Particle Identification	Raw Count	(Count/m³)	% of Total	Interpretation Guideline
412606013-0006	Alternaria (Ulocladium)	-	-	-	
	Ascospores	-	-	-	
Client Sample ID	Aspergillus/Penicillium++	264	10800	99.6	ELEVATED
42439659	Basidiospores	1	40	0.4	Acceptable
	Bipolaris++	-	-	-	
	Chaetomium++	-	-	-	
Location	Cladosporium	-	-	-	
Access Hall (B)	Curvularia	-	-	-	
	Epicoccum	-	-	-	
Sample Volume (L)	Fusarium++	-	-	-	
	Ganoderma	-	-	-	
75	Myxomycetes++	-	-	-	
	Pithomyces++	-	-	-	
Sample Type	Rust	-	-	-	
	Scopulariopsis/Microascus	-	-	-	
Inside	Stachybotrys/Memnoniella	-	-	-	
Comments	Zygomycetes	-	-	-	
	Cercospora++	-	-	-	
	Nigrospora	-	-	-	
	Polythrincium	-	-	-	
	Sporidesmium++	-	-	-	
	Total Fungi	**265**	**10840**	**100**	Acceptable
	Hyphal Fragment	-	-	-	
	Insect Fragment	-	-	-	
	Pollen	-	-	-	

Analytical Sensitivity 600x: **41** counts/cubic meter
Analytical Sensitivity 300x *: **13*** counts/cubic meter

Skin Fragments: **2** 1 to 4 (low to high)
Fibrous Particulate: **1** 1 to 4 (low to high)
Background: **1** 1 to 4 (low to high); 5 (overloaded)

Acceptable Concentration at or below background
Slightly Elevated Concentration above background
ELEVATED Concentration 10X or more above background

Not commonly found growing indoors, spores likely come from outside.
Spores reported to be able to cause allergies in individuals.
Potential for mycotoxin production exists with these fungi.
These fungi are considered water damage indicators.

++ Includes other spores with similar morphology; see EMSL's fungal glossary for each specific category.
-

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Lee Plumley, Laboratory Manager
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Attn: Jim Dallas
Centric Environmental Solutions, LLC
1325 Scotts Creek Road
Statesville, NC 28625

EMSL Order: 412606013
Customer ID: CENT75
Collected: 8/11/2026
Received: 8/12/2026
Analyzed: 8/13/2026

Proj: CES779

Spore Trap ASSESSMENTReport™ Air-O-Cell(™) Analysis of Fungal Spores & Particulates (Methods MICRO-SOP-201, ASTM D7391)

	Particle Identification	Raw Count	(Count/m³)	% of Total	Interpretation Guideline
412606013-0007	Alternaria (Ulocladium)	-	-	-	
	Ascospores	-	-	-	
Client Sample ID	Aspergillus/Penicillium++	5	200	71.4	Slightly Elevated
42439646	Basidiospores	1	40	14.3	Acceptable
	Bipolaris++	-	-	-	
	Chaetomium++	-	-	-	
Location	Cladosporium	-	-	-	
Blue Hallway	Curvularia	-	-	-	
	Epicoccum	-	-	-	
Sample Volume (L)	Fusarium++	-	-	-	
	Ganoderma	-	-	-	
75	Myxomycetes++	-	-	-	
	Pithomyces++	-	-	-	
Sample Type	Rust	-	-	-	
	Scopulariopsis/Microascus	-	-	-	
Inside	Stachybotrys/Memnoniella	-	-	-	
Comments	Zygomycetes	1	40	14.3	Slightly Elevated
	Cercospora++	-	-	-	
	Nigrospora	-	-	-	
	Polythrincium	-	-	-	
	Sporidesmium++	-	-	-	
	Total Fungi	**7**	**280**	**100**	Acceptable
	Hyphal Fragment	-	-	-	
	Insect Fragment	-	-	-	
	Pollen	-	-	-	

Analytical Sensitivity 600x: **41** counts/cubic meter
Analytical Sensitivity 300x *: **13*** counts/cubic meter

Skin Fragments: **2** 1 to 4 (low to high)
Fibrous Particulate: **1** 1 to 4 (low to high)
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Acceptable Concentration at or below background
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ELEVATED Concentration 10X or more above background

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Spores reported to be able to cause allergies in individuals.
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Analyzed: 8/13/2026

Proj: CES779

Spore Trap ASSESSMENTReport™ Air-O-Cell(™) Analysis of Fungal Spores & Particulates (Methods MICRO-SOP-201, ASTM D7391)

	Particle Identification	Raw Count	(Count/m³)	% of Total	Interpretation Guideline
412606013-0008	Alternaria (Ulocladium)	-	-	-	
	Ascospores	-	-	-	
Client Sample ID	Aspergillus/Penicillium++	24	990	100	ELEVATED
42439639	Basidiospores	-	-	-	
	Bipolaris++	-	-	-	
	Chaetomium++	-	-	-	
Location	Cladosporium	-	-	-	
Classroom D122	Curvularia	-	-	-	
	Epicoccum	-	-	-	
Sample Volume (L)	Fusarium++	-	-	-	
	Ganoderma	-	-	-	
75	Myxomycetes++	-	-	-	
	Pithomyces++	-	-	-	
Sample Type	Rust	-	-	-	
	Scopulariopsis/Microascus	-	-	-	
Inside	Stachybotrys/Memnoniella	-	-	-	
Comments	Zygomycetes	-	-	-	
	Cercospora++	-	-	-	
	Nigrospora	-	-	-	
	Polythrincium	-	-	-	
	Sporidesmium++	-	-	-	
	Total Fungi	**24**	**990**	**100**	Acceptable
	Hyphal Fragment	-	-	-	
	Insect Fragment	-	-	-	
	Pollen	-	-	-	

Analytical Sensitivity 600x: **41** counts/cubic meter
Analytical Sensitivity 300x *: **13*** counts/cubic meter

Skin Fragments: **2** 1 to 4 (low to high)
Fibrous Particulate: **1** 1 to 4 (low to high)
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Proj: CES779

Spore Trap ASSESSMENTReport™ Air-O-Cell(™) Analysis of Fungal Spores & Particulates (Methods MICRO-SOP-201, ASTM D7391)

	Particle Identification	Raw Count	(Count/m³)	% of Total	Interpretation Guideline
412606013-0009	Alternaria (Ulocladium)	-	-	-	
	Ascospores	6	200	1.3	
Client Sample ID	Aspergillus/Penicillium++	-	-	-	
42440760	Basidiospores	354	14500	90.9	
	Bipolaris++	-	-	-	
	Chaetomium++	-	-	-	
Location	Cladosporium	20	820	5.1	
Outdoor Reference	Curvularia	2	80	0.5	
	Epicoccum	-	-	-	
Sample Volume (L)	Fusarium++	6	200	1.3	
	Ganoderma	-	-	-	
75	Myxomycetes++	-	-	-	
	Pithomyces++	-	-	-	
Sample Type	Rust	-	-	-	
	Scopulariopsis/Microascus	-	-	-	
Background	Stachybotrys/Memnoniella	-	-	-	
Comments	Zygomycetes	-	-	-	
	Cercospora++	1	40	0.3	
	Nigrospora	1	40	0.3	
	Polythrincium	1	40	0.3	
	Sporidesmium++	1	40	0.3	
	Total Fungi	**392**	**15960**	**100**	
	Hyphal Fragment	-	-	-	
	Insect Fragment	-	-	-	
	Pollen	-	-	-	

Analytical Sensitivity 600x: **41** counts/cubic meter
Analytical Sensitivity 300x *: **13*** counts/cubic meter

Skin Fragments: **1** 1 to 4 (low to high)
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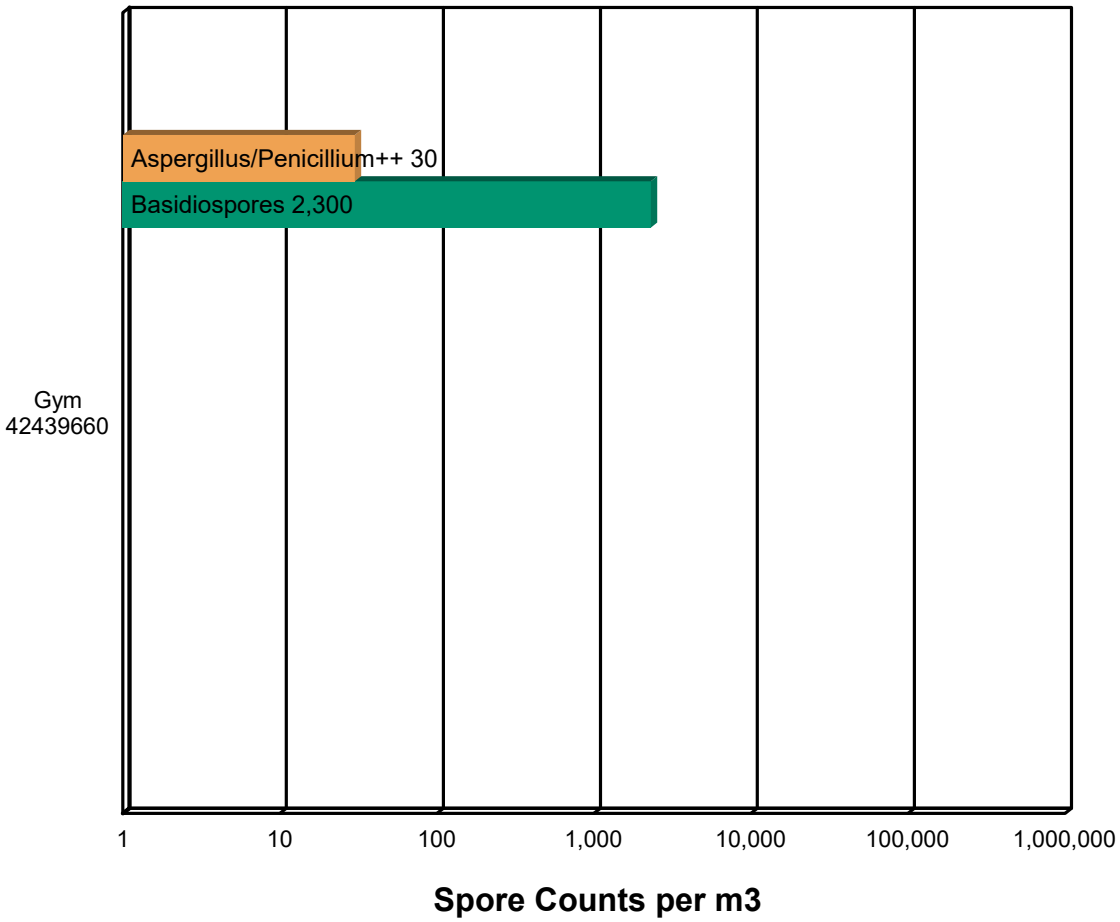
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EMSL Order: 412606013
Customer ID: CENT75
Collected: 8/11/2026
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Analyzed: 8/13/2026

Proj: CES779

Spore Trap Report: Total Counts



* The chart is displayed using a logarithmic scale. Bar size is not directly proportional to the number of spores.

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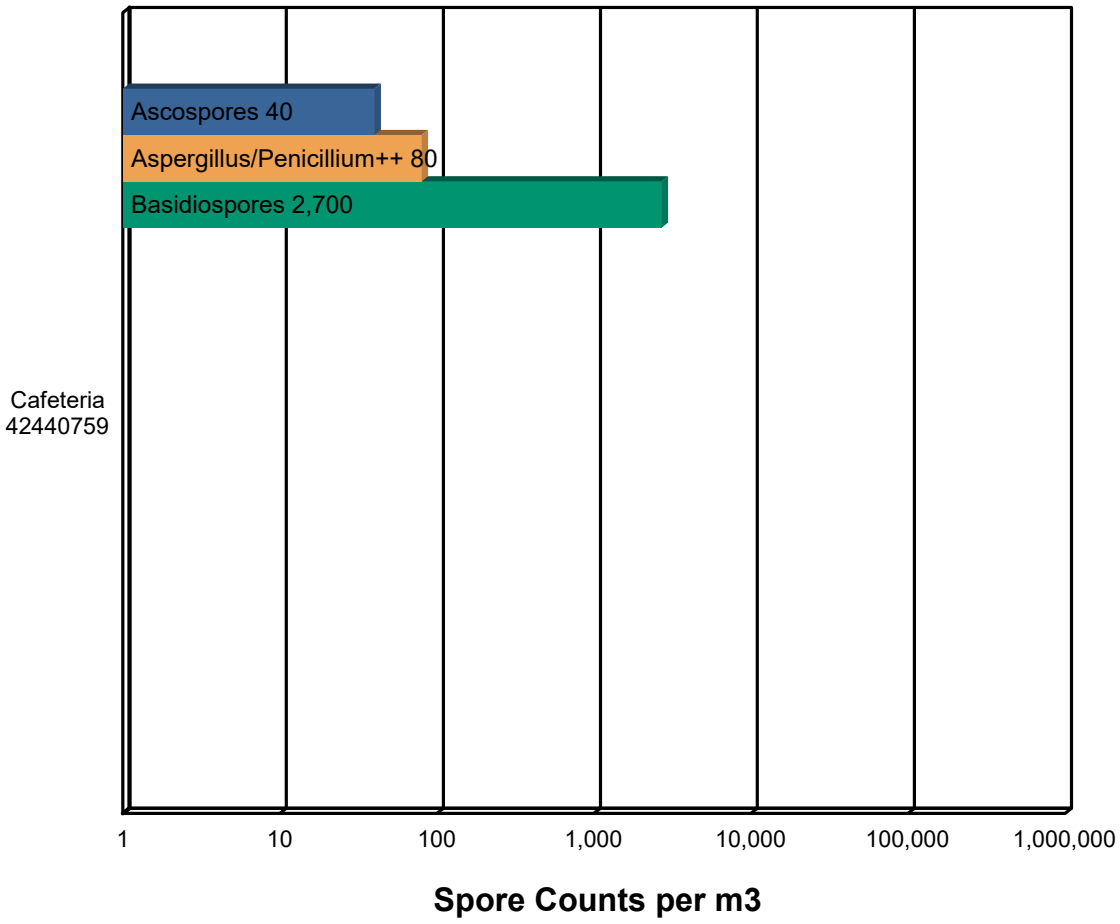
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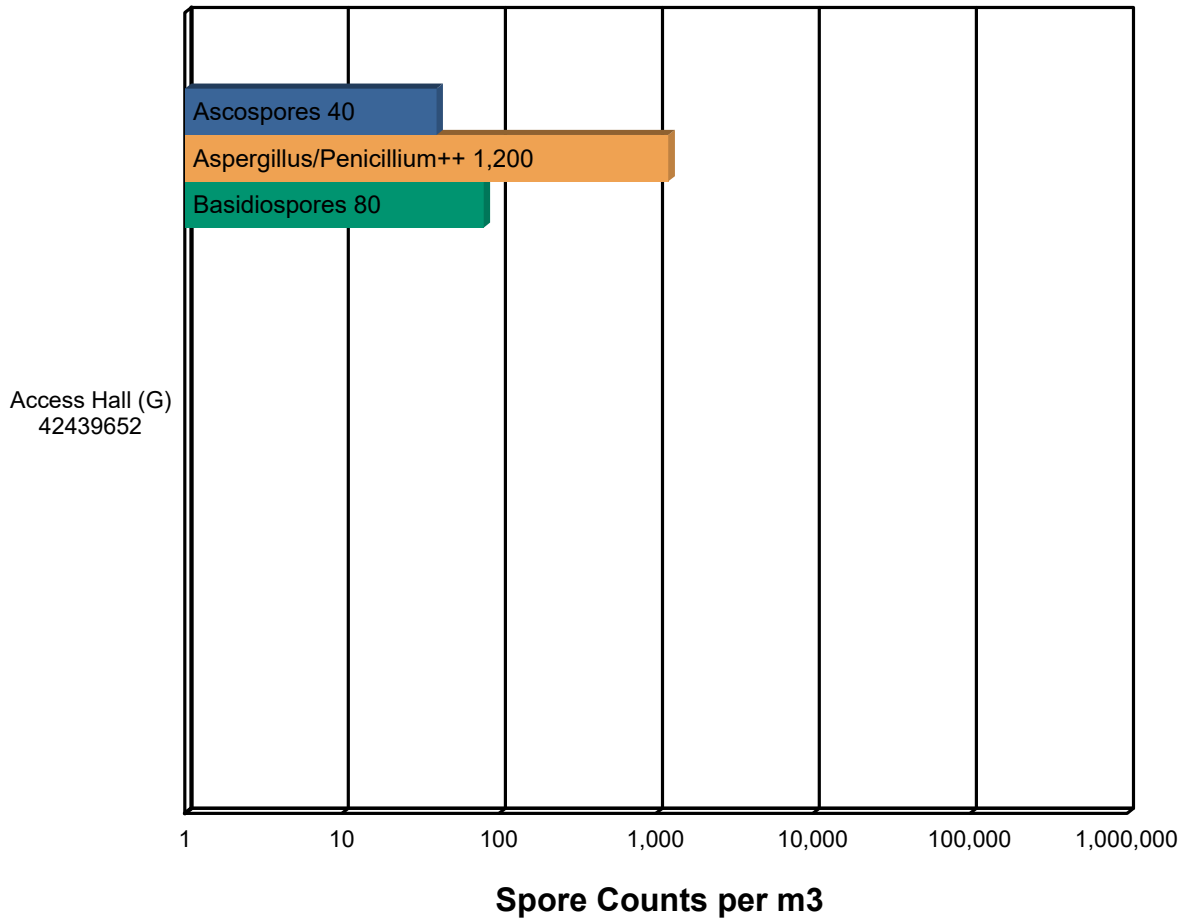
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Spore Trap Report: Total Counts



Ascospores
Aspergillus/Penicillium++
Basidiospores
Cercospora++
Cladosporium
Curvularia
Epicoccum
Fusarium++
Myxomycetes++
Nigrospora
Pithomyces++
Polythrincium
Sporidesmium++
Zygomycetes

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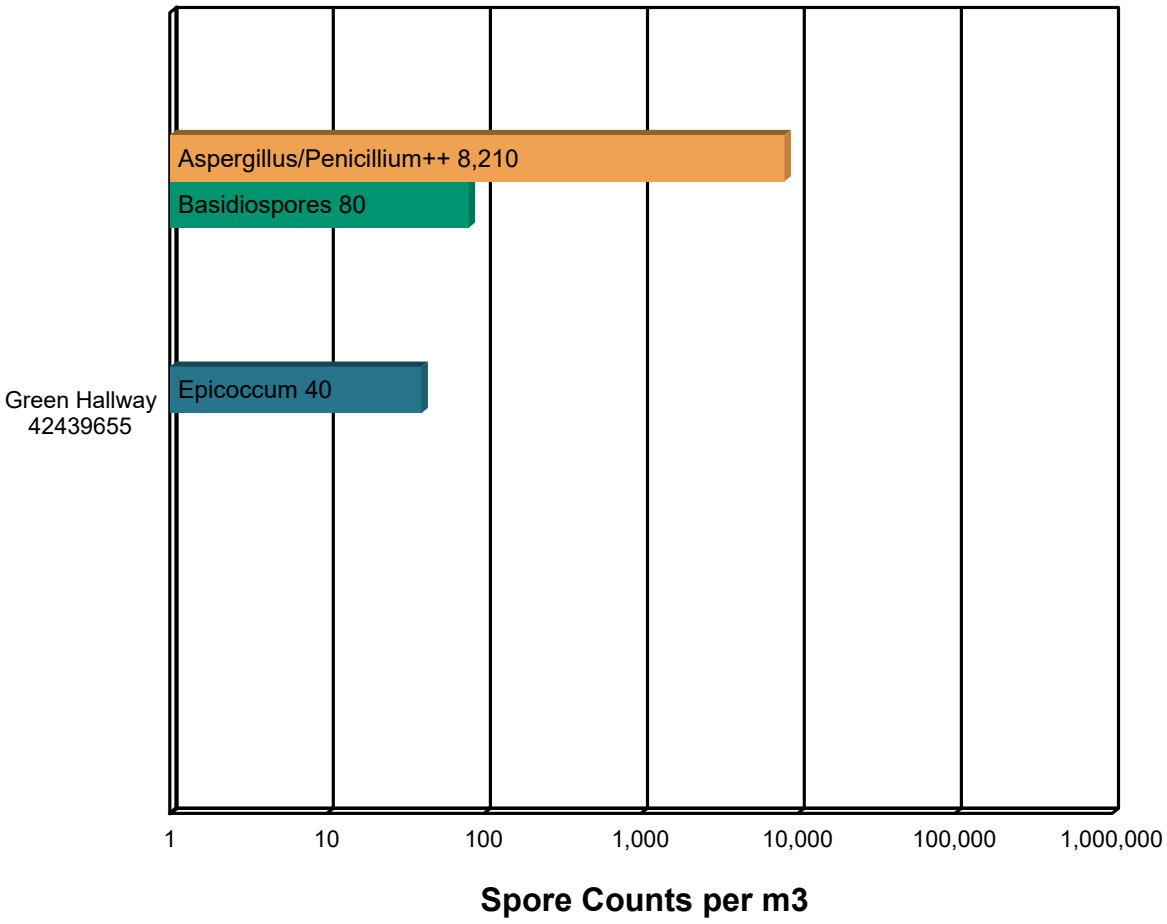
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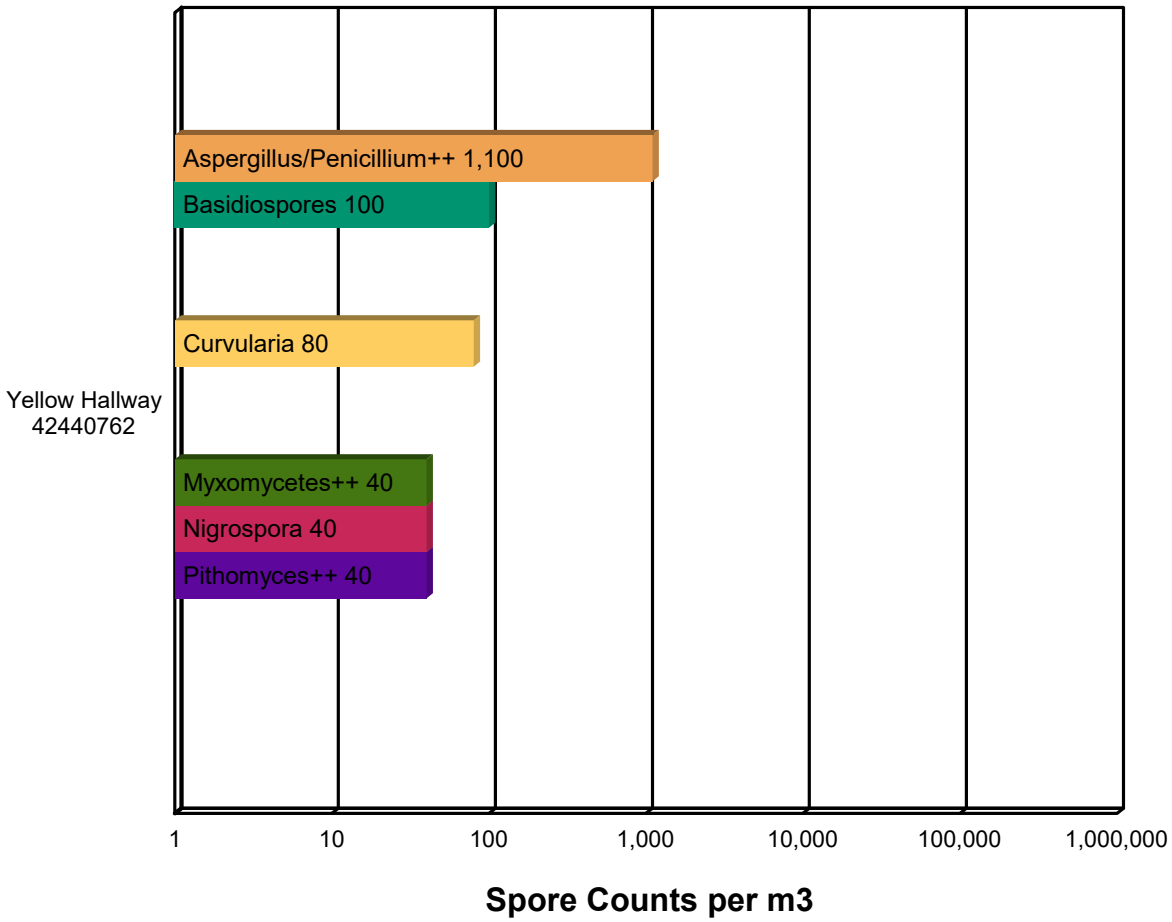
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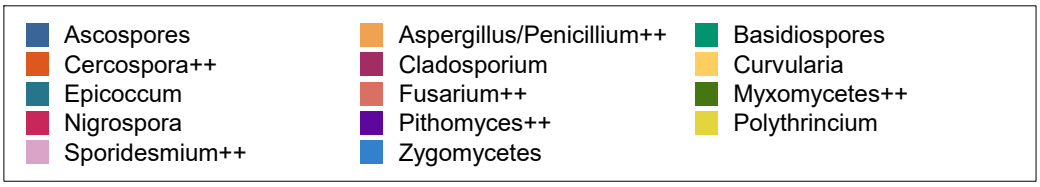
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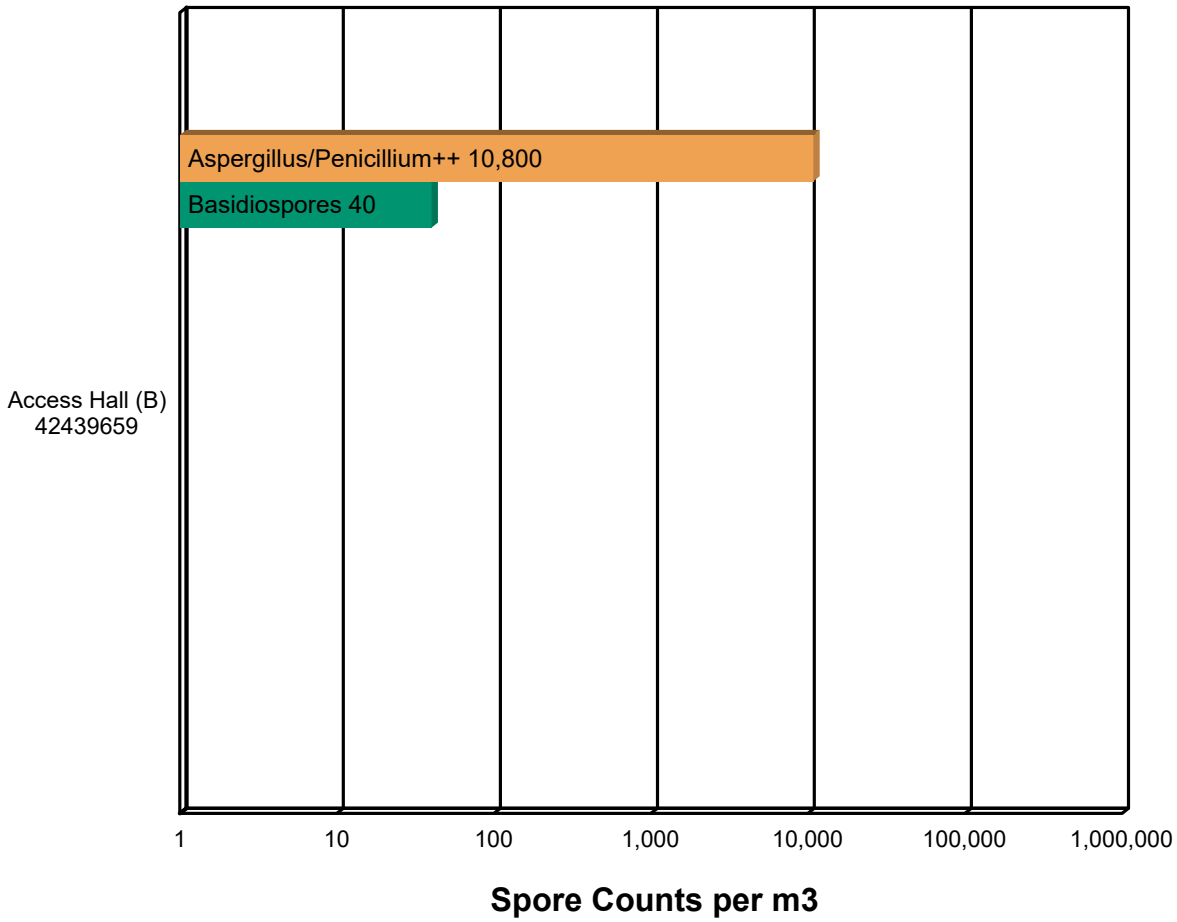
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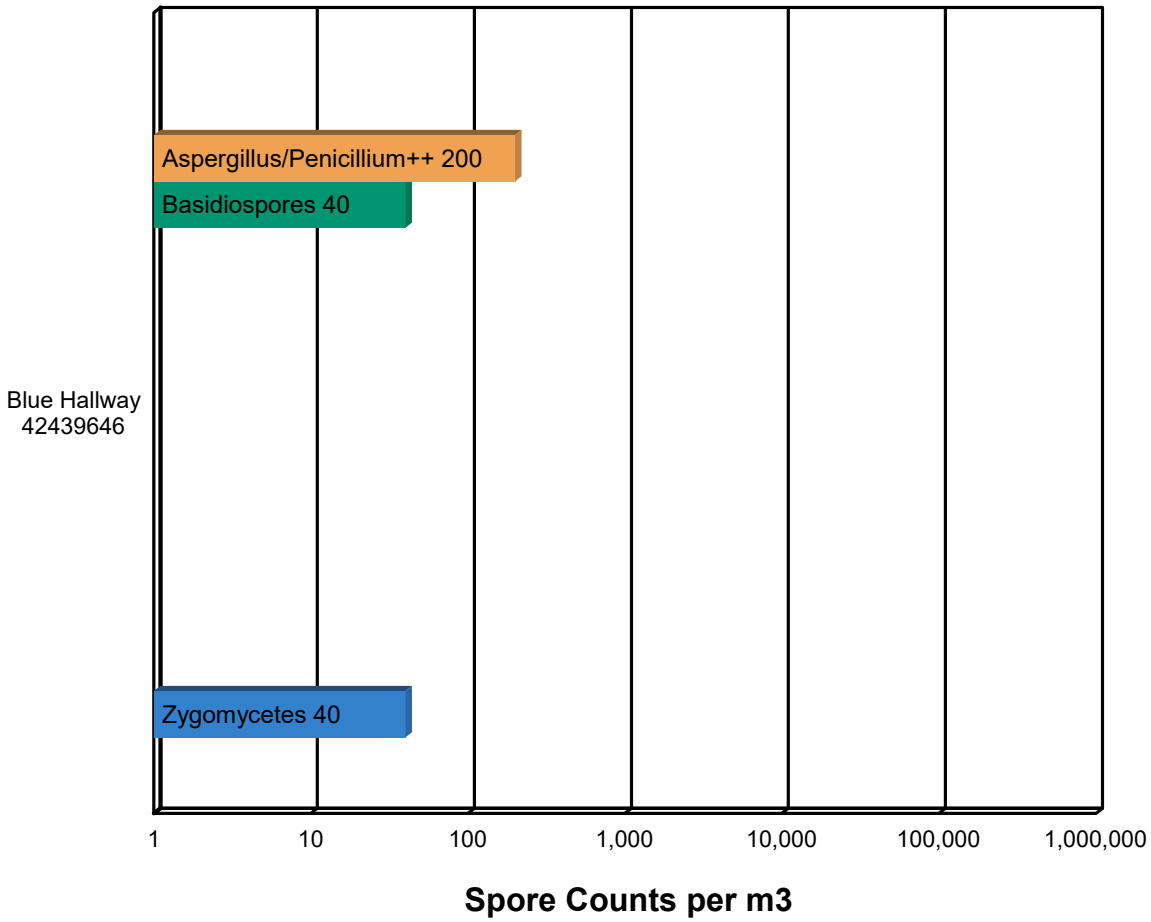
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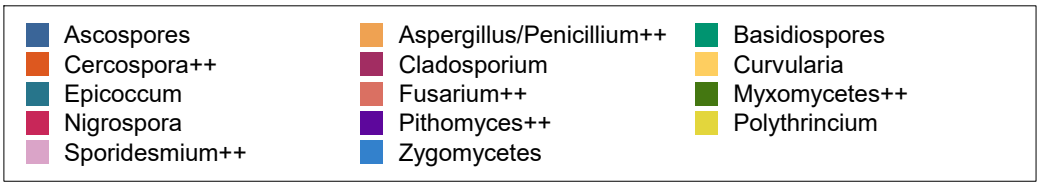
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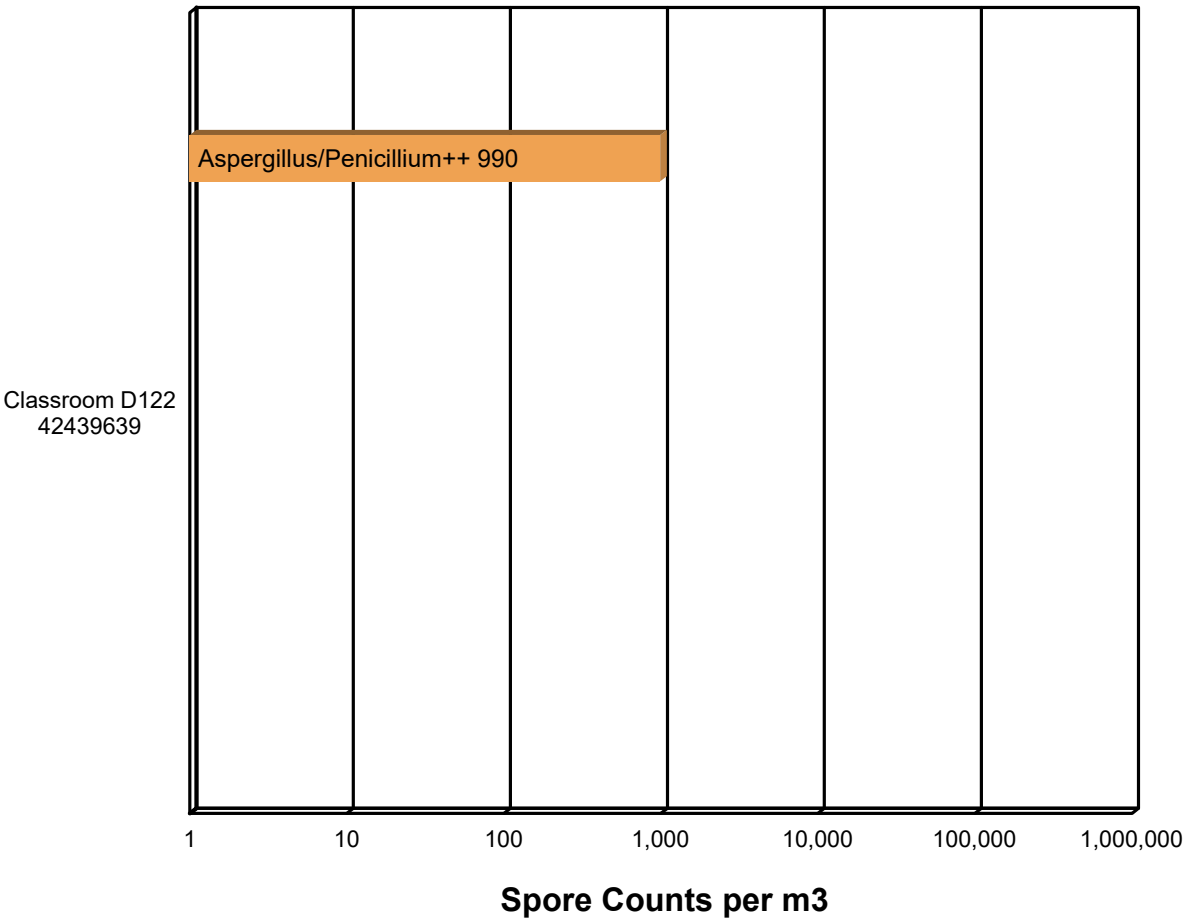
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Phone: (704) 525-2205 Fax: (704) 525-2382 Web: http://www.EMSL.com Email:charlottelab@emsl.com

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Centric Environmental Solutions, LLC
1325 Scotts Creek Road
Statesville, NC 28625

Proj: CES779

EMSL Order:	412606013
Customer ID:	CENT75
Collected:	8/11/2026
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Analyzed:	8/13/2026

Spore Trap Report: Total Counts



* The chart is displayed using a logarithmic scale. Bar size is not directly proportional to the number of spores.

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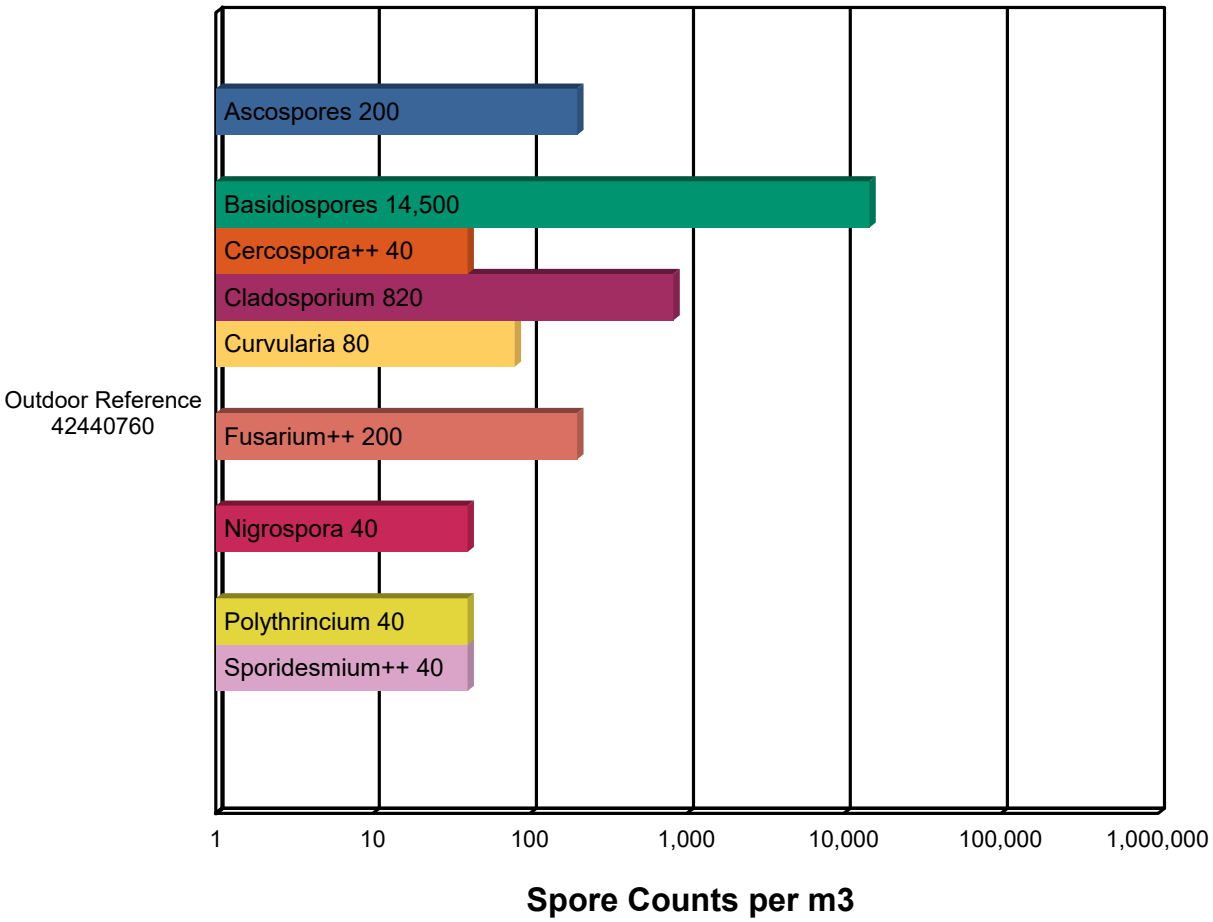
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Spore Trap Report: Total Counts



Ascospores
Aspergillus/Penicillium++
Basidiospores
Cercospora++
Cladosporium
Curvularia
Epicoccum
Fusarium++
Myxomycetes++
Nigrospora
Pithomyces++
Polythrincium
Sporidesmium++
Zygomycetes

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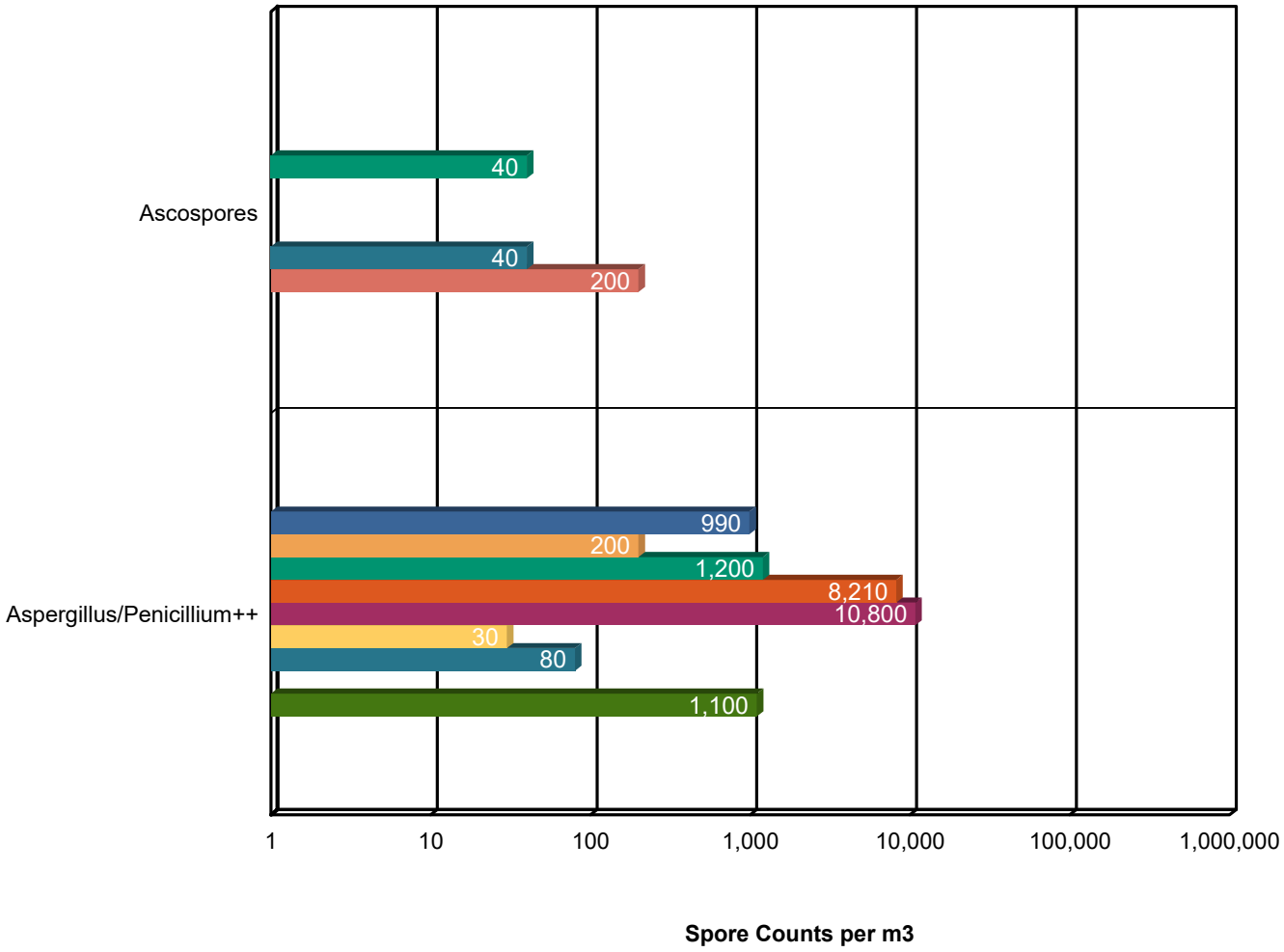
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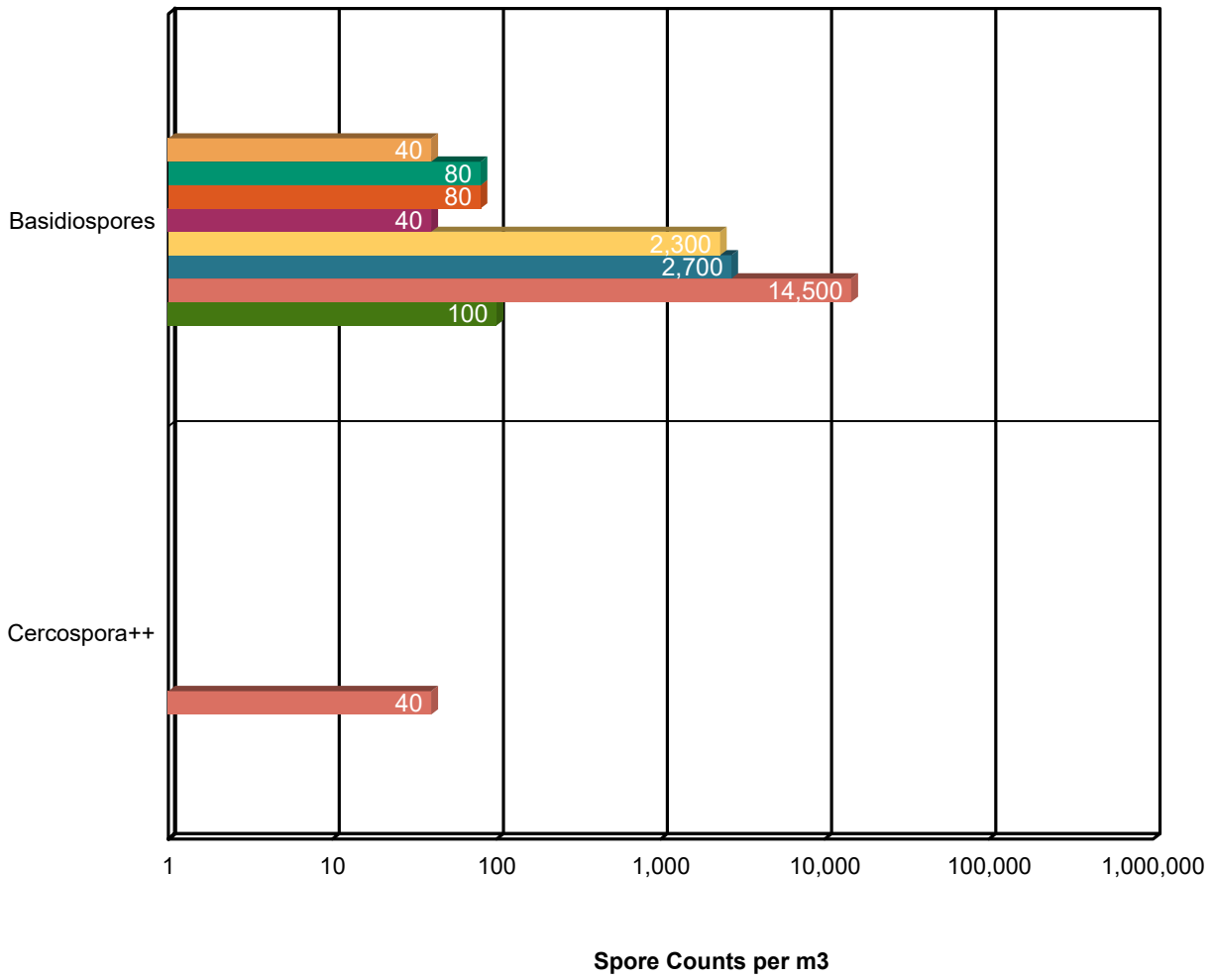
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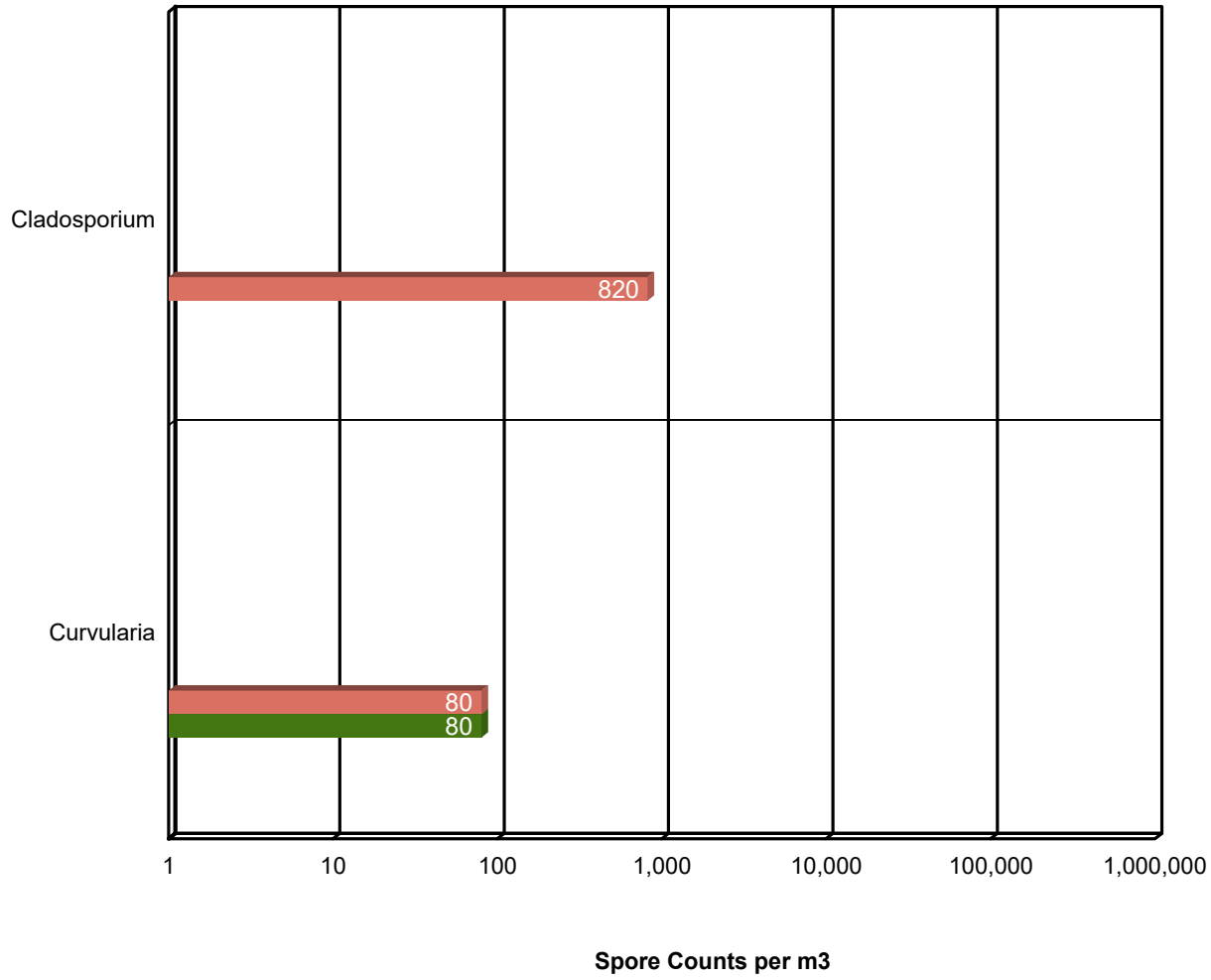
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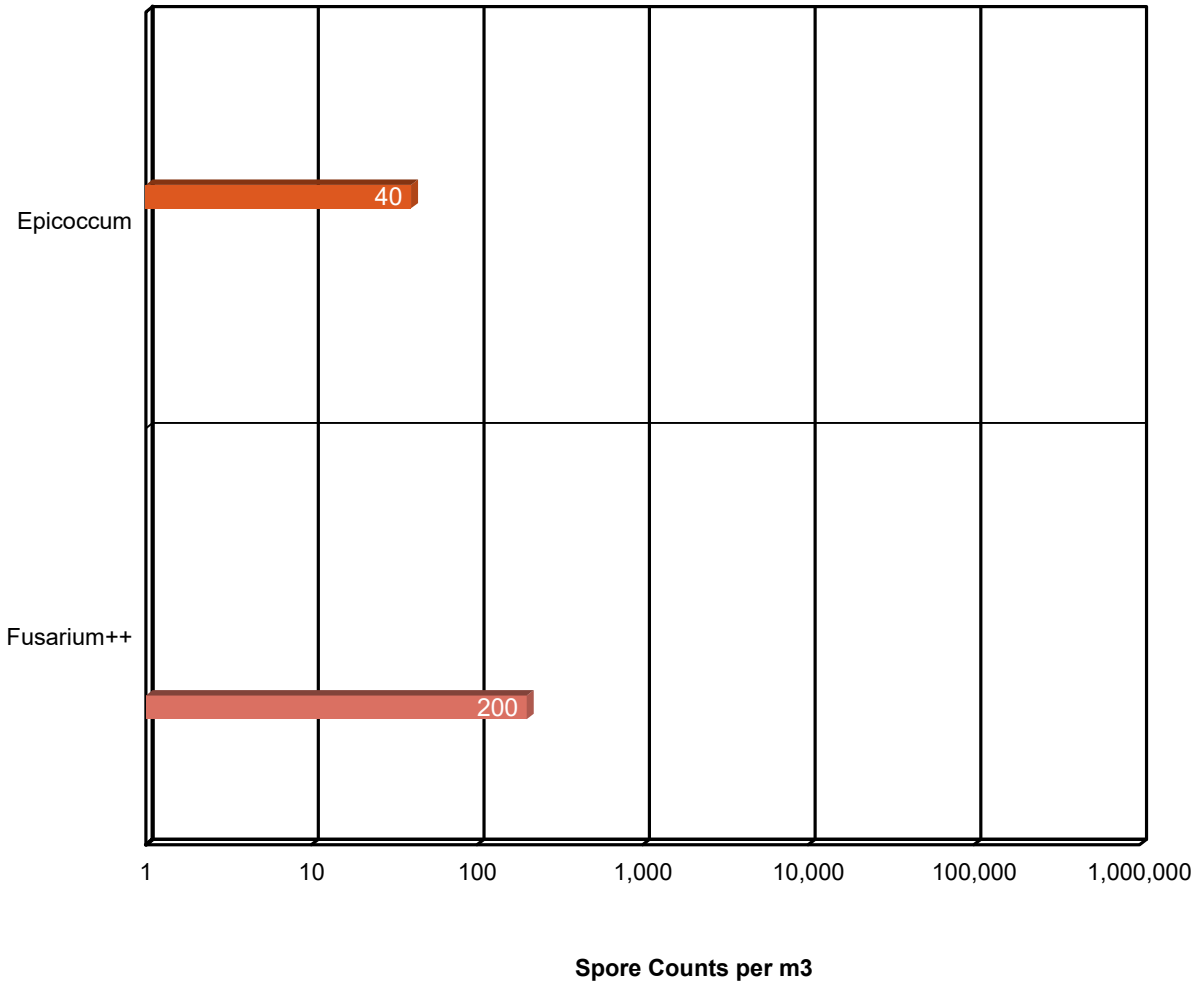
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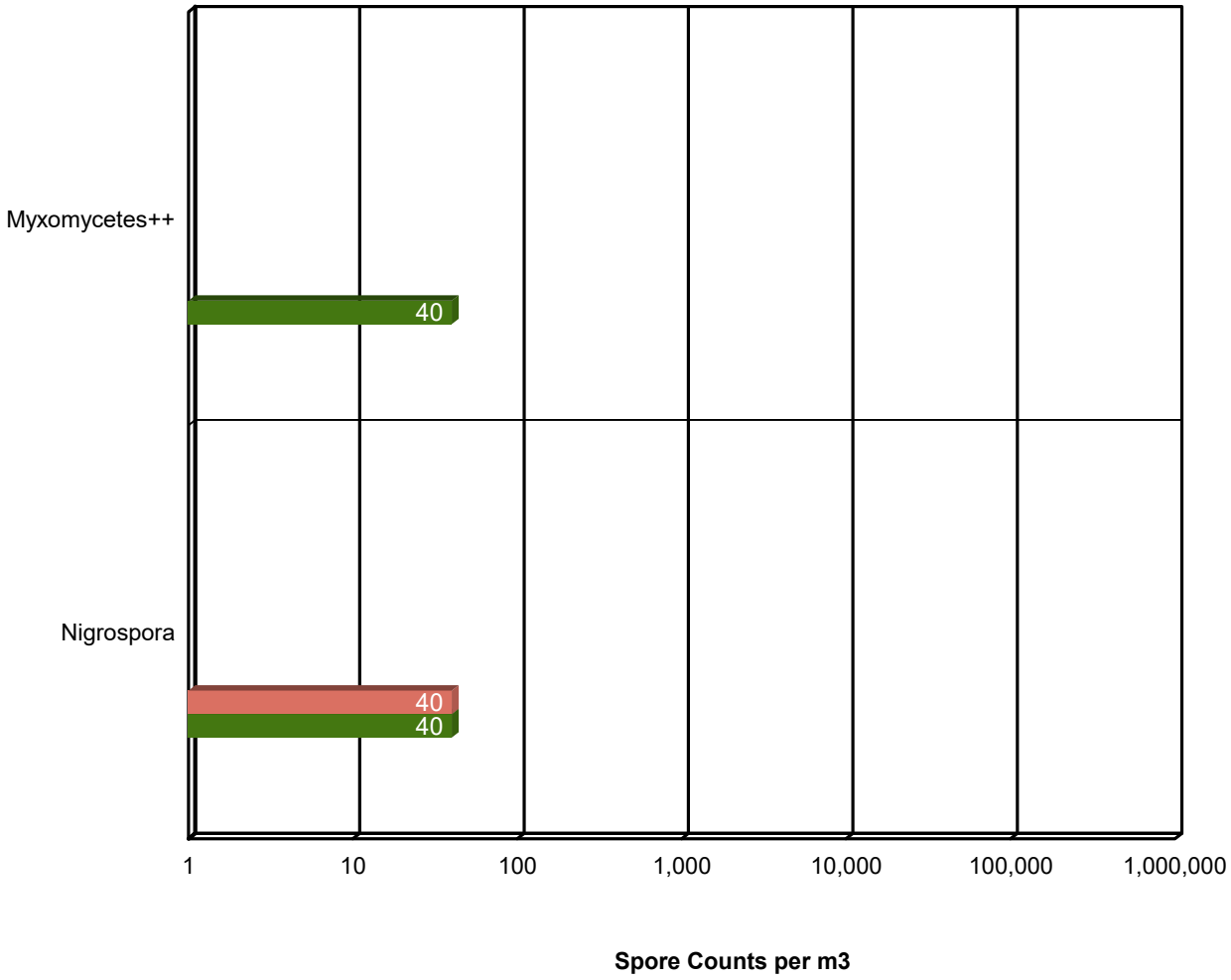
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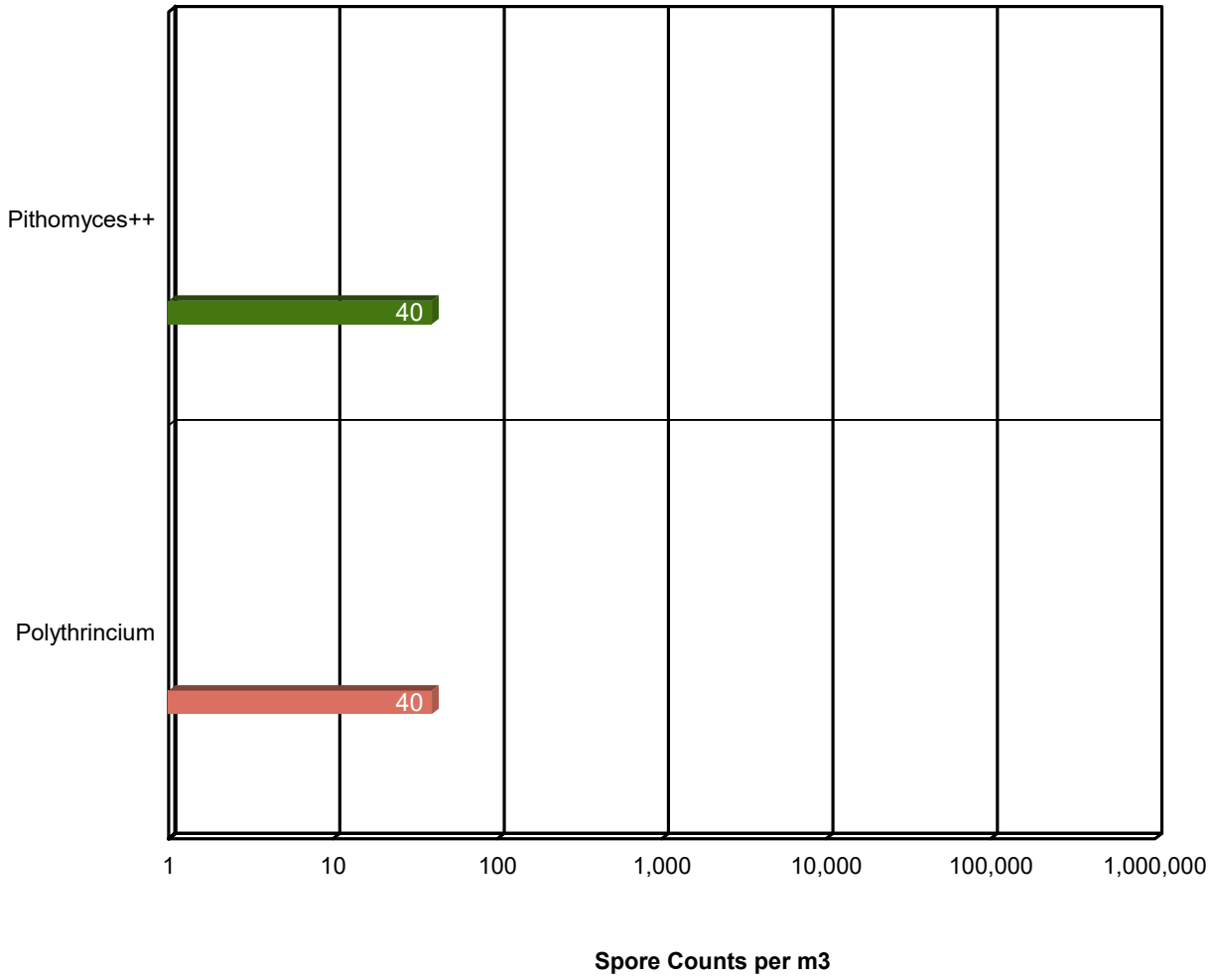
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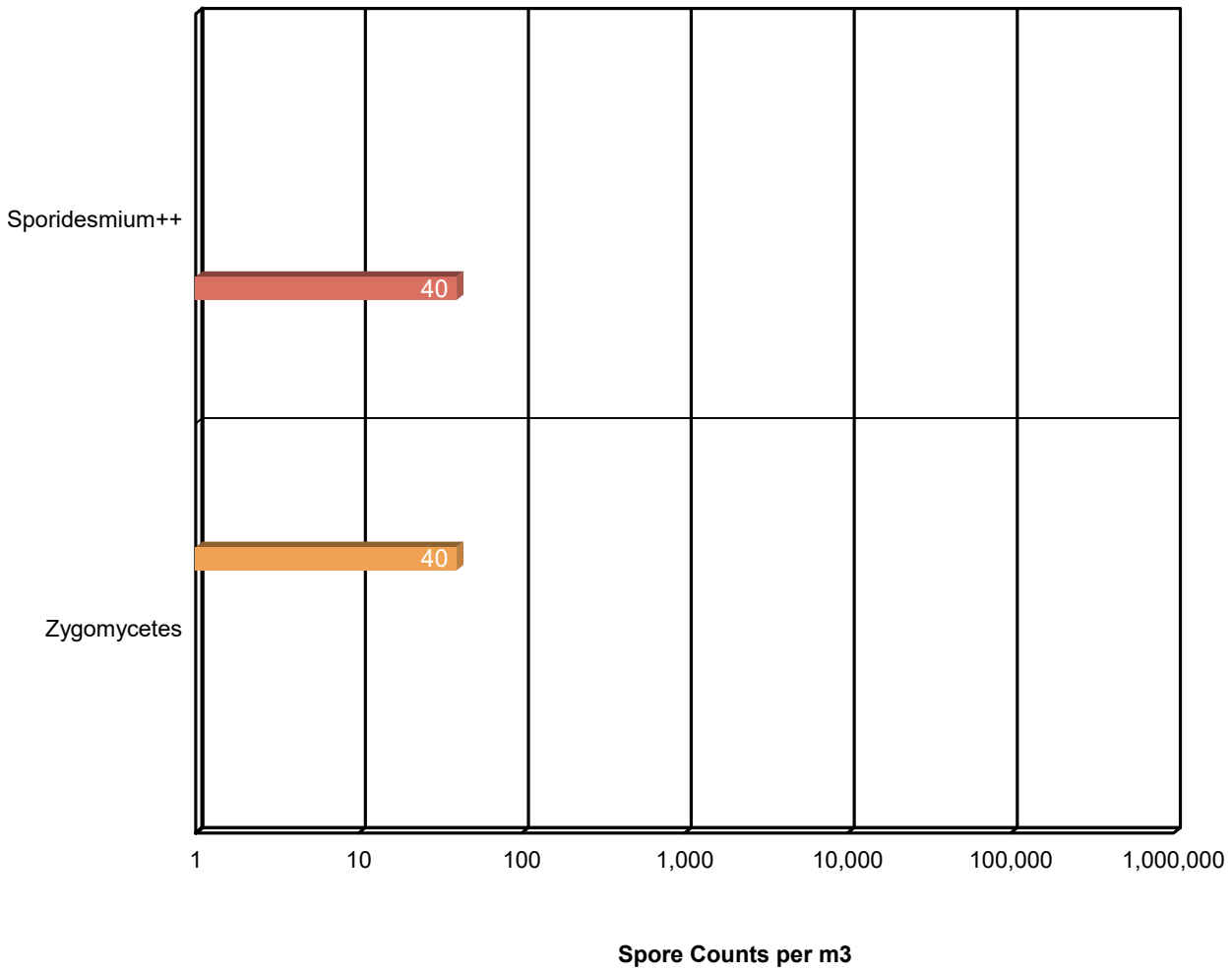
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3. Understanding the Results

EMSL Analytical, Inc. is an independent laboratory, providing unbiased and scientifically valid results. These data represent only a portion of an overall IAQ investigation. Visual information and environmental conditions measured during the site assessment (humidity, moisture readings, etc.) are crucial to any final interpretation of the results. Many factors impact the final results; therefore, result interpretation should only be conducted by qualified individuals. The American Conference of Governmental Industrial Hygienists (ACGIH) has published a good reference book covering sampling and data interpretation. It is entitled, Bioaerosols: Assessment and Control, 1999.

Fungal spores are found everywhere. Whether or not symptoms develop in people exposed to fungi depends on the nature of the fungal material (e.g., allergenic, toxic, or infectious), the exposure level, and the susceptibility of exposed persons. Susceptibility varies with the genetic predisposition (e.g., allergic reactions do not always occur in all individuals), age, pre-existing medical conditions (e.g., diabetes, cancer, or chronic lung conditions), use of immunosuppressive drugs, and concurrent exposures. These reasons make it difficult to identify dose/response relationships that are required to establish “safe” or “unsafe” levels (i.e., permissible exposure limits).

It is generally accepted in the industry that indoor fungal growth is undesirable and inappropriate, necessitating removal or other appropriate remedial actions. The New York City guidelines and EPA guidelines for mold remediation in schools and commercial buildings define the conditions warranting mold remediation. Always remember that water is the key. Preventing water damage or water condensation will prevent mold growth.

This report is not intended to provide medical advice or advice concerning the relative safety of an occupied space. Always consult an occupational or environmental health physician who has experience addressing indoor air contaminants if you have any questions.

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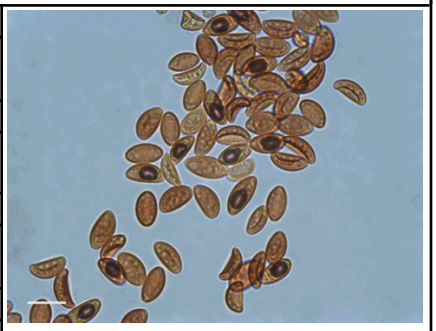
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4. Glossary of Fungi

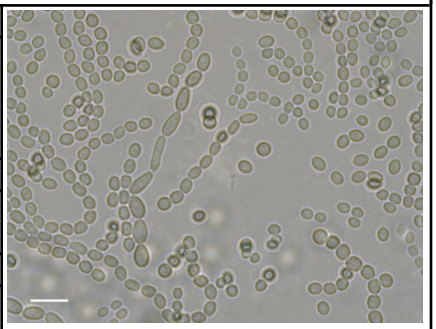
ASCOSPORES

Natural Habitat	Everywhere in nature.
Suitable Substrates in the Indoor Environment	Depends on genus and species.
Water Activity	Depends on genus and species.
Mode of Dissemination	Forcible ejection or passive release and dissemination by wind or insects.
Allergic Potential	Depends on genus and species.
Potential or Opportunistic Pathogens	Depends on genus and species.
Industrial Uses	Depends on genus and species.
Potential Toxins Produced	Depends on genus and species.
Other Comments	Ascospores are the result of sexual reproduction and produced in a saclike structure called an ascus. All ascospores belong to members of the Phylum Ascomycota, which encompasses a plethora of genera worldwide.



ASPERGILLUS/PENICILLIUM++

Natural Habitat	Plant debris ·Seed ·Cereal crop
Suitable Substrates in the Indoor Environment	Grows on a wide range of substrates indoors ·Prevalent in water damaged buildings ·Foods (blue mold on cereals, fruits, vegetables, dried foods) ·House dust ·Fabrics ·Leather ·Wallpaper ·Wallpaper glue
Allergic Potential	Type I (hay fever, asthma) ·Type III (hypersensitivity)
Potential Opportunist or Pathogen	Possible depending on the species.
Potential Toxins Produced	Possible depending on the species.
Free moisture required for mold growth	Aw=0.75-0.94
Mode of Dissemination	Wind ·Insects
Industrial Uses	Many depending on the species
Other comments	Spores of Aspergillus and Penicillium (including others such as Geosmithia, Goidanichella, Nalanthamala, Rasamsonia, Samsoniella, and Talaromyces) are small and spherical with few distinguishing characteristics. They cannot be differentiated by non-viable impaction sampling methods. Some species with very small spores may be undercounted in samples with high background debris.



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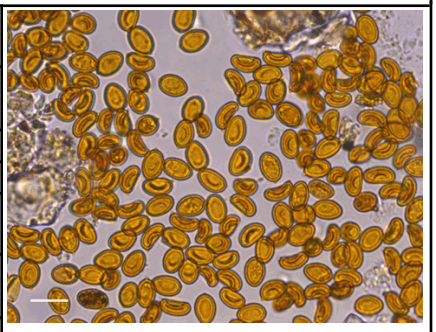
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BASIDIOSPORES

Natural Habitat	Forest floors. Lawns .Plants (saprobes or pathogens depending on genus)
Suitable Substrates in the Indoor Environment	Depends on genus. Wood products
Water Activity	Unknown.
Mode of Dissemination	Forcible ejection. Wind currents.
Allergic Potential	Type I allergies (hay fever, asthma) . Type III (hypersensitivity pneumonitis)
Potential or Opportunistic Pathogens	Depends on genus.
Industrial Uses	Edible mushrooms are used in the food industry.
Potential Toxins Produced	Amanitins. monomethyl-hydrazine. muscarine. ibotenic acid. psilocybin.
Other Comments	Basidiospores are the result of sexual reproduction and formed on a structure called the basidium. Basidiospores belong to the members of the Phylum Basidiomycota, which includes mushrooms, shelf fungi, rusts, and smuts.



CERCOSPORA++

Natural Habitat	Parasite on higher plants, commonly causes leaf spot diseases.
Suitable Substrates in the Indoor Environment	Unknown
Water Activity	Moderate –High humidity
Mode of Dissemination	Irrigation water, Insects, Rain Wind
Allergic Potential	Unknown
Potential or Opportunistic Pathogens	Unknown
Other Comments	Includes morphologically similar spores of Cercospora, Pseudocercospora, Septoriella, and Septoria.



CLADOSPORIUM

Natural Habitat	Dead plant matter. Straw. Soil. Woody plants
Suitable Substrates in the Indoor Environment	Fiberglass duct liner. Paint. Textiles. Found in high concentration in water-damaged building materials.
Water Activity	Aw 0.84-0.88
Mode of Dissemination	Air
Allergic Potential	Type I (asthma and hay fever).
Potential or Opportunistic Pathogens	Edema. keratitis. onychomycosis. pulmonary infections. Sinusitis.
Industrial Uses	Produces 10 antigens.
Potential Toxins Produced	Cladosporin and Emodin.



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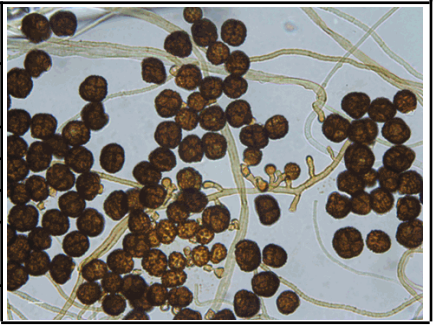
CURVULARIA

Natural Habitat	A worldwide saprophytic fungi, being isolated from dead plant material and soil.
Suitable Substrates in the Indoor Environment	Paper, wood products
Free moisture required for mold growth	Unknown
Mode of Dissemination	Wind
Allergic Potential	Hay fever, asthma, allergic fungal sinusitis
Potential or Opportunistic Pathogens	In immunocompromised patients can cause cerebral abscess, endocarditis, mycetoma, ocular keratitis, onychomycosis, and pneumonia.



EPICOCCUM

Natural Habitat	A worldwide saprophytic fungi, being isolated from dead plant material and soil.
Suitable Substrates in the Indoor Environment	Paper, textiles
Water Activity	0.86-0.90
Mode of Dissemination	Wind
Allergic Potential	Hay fever, asthma
Potential or Opportunistic Pathogens	Unknown



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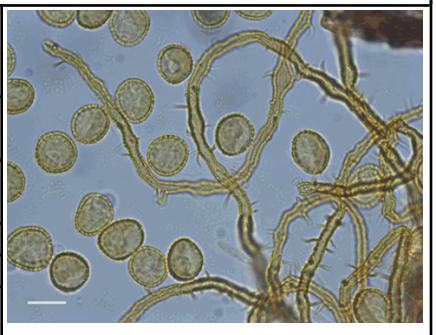
FUSARIUM++

Natural Habitat	Soil. Plant pathogen causing root rot, stem rot, and wilt of many ornamental and crop plants. Genera with like spores include Fusarium, Albonectria, Atractium, Bisifusarium, Corinectria, Cosmospora, Cosmosporella, Cyanonectria, Dialonectria, Fusicolla, Geejayessia, Ilyonectria, Luteonectria, Macroconia, Mariannaea, Microcera, Neocosmospora, Neonectria, Nothofusarium, Pseudofusicolla Rectifusarium, Rugonectria, Scolecofusarium, Setofusarium, Stylonectria, Thelonectria, and Tumenectria.
Suitable Substrates in the Indoor Environment	Often found in humidifiers. Wet, cellulose-based building materials
Water Activity	Aw=0.86-0.91
Mode of Dissemination	Insects. Water droplets, rain. Wind when spores become dry.
Allergic Potential	Type I allergies (hay fever, asthma).
Potential or Opportunistic Pathogens	Esophageal cancer is believed to happen after consumption of F. moniliforme infected corn. Keratitis. Endophthalmitis. Onychomycosis. Cutaneous infections. Mycetoma. Sinusitis. Pulmonary infections. Endocarditis. Peritonitis. Central venous catheter infections. Septic arthritis. Neurological disease in horses after consumption of F. moniliforme infected corn. Respiratory disease in pigs after consumption of F. moniliforme infected corn.
Industrial Uses	Biological Weapon.
Potential Toxins Produced	Trichothecenes. Zearalenone. Fumonisins.
Other Comments	Major plant pathogen.
Reference	Atlas of Moulds in Europe causing respiratory Allergy, Foundation for Allergy Research in Europe, Edited by Knud Wilken-Jensen and Suzanne Gravesen, ASK Publishing, Denmark, 1984.



MYXOMYCETES++

Natural Habitat	Decaying logs, Dead leaves , Dung , Lawns , Mulched flower beds, Lawns
Suitable Substrates in the Indoor Environment	Rotting lumber
Free moisture required for mold growth	Unknown
Mode of Dissemination	Insects, Water, Wind
Allergic Potential	Type I
Potential or Opportunistic Pathogens	Unknown
Industrial Uses	
Other Comments	Includes Myxomycetes, Smut, Rust, and Periconia.



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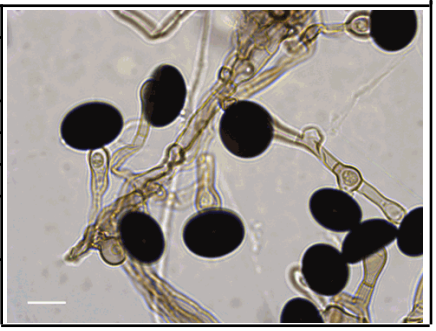
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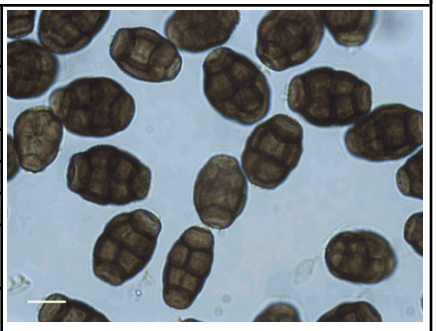
NIGROSPORA

Natural Habitat	Common on live or dead grass, seeds & soil.
Suitable Substrates in the Indoor Environment	Unknown
Water Activity	Unknown
Mode of Dissemination	Forcibly projected.
Allergic Potential	Type 1 allergies (hey fever, asthma)
Potential or Opportunistic Pathogens	Keratitis & skin lesions



PITHOMYCES++

Natural Habitat	A worldwide saprophytic fungi, being isolated from dead plant material and soil.
Suitable Substrates in the Indoor Environment	Paper
Water Activity	Requires high moisture for spore germination
Mode of Dissemination	Wind
Allergic Potential	Unknown
Potential or Opportunistic Pathogens	Mycosis in immunocompromised patients
Other Comments	Pithomyces++ includes spores of Pithomyces and Pseudopithomyces.



POLYTHRINCIUM

Natural Habitat	Many Basidiomycetes form arthrospores during their mycelial stage. Geotrichum and Oidiodendron are typical ascomycete arthrospore formers. Arthrospores are formed by microfungi, and yeast-like fungi. Arthrospores are disarticulated cells of a formerly vegetative filament that function as spores.
Suitable Substrates in the Indoor Environment	Unknown
Allergic Potential	Allergenic potential in this genus is not well understood, and is currently being studied.
Potential Opportunist or Pathogen	Unknown
Potential Toxins Produced	Unknown



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Statesville, NC 28625

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ZYGOMYCETES

Natural Habitat	Decaying plant matter, Decaying animal matter	
Suitable Substrates in the Indoor Environment	Fruits and vegetables	
Free moisture required for mold growth	Aw=0.90-0.95	
Mode of Dissemination	Water splash, Wind	
Allergic Potential	Type I (hay fever, asthma) & Type III (hypersensitivity)	
Potential or Opportunistic Pathogens	Some Zygomycetes can cause zygomycosis in immonocompromised patients. Zygomycosis can occur in the lungs, nasal sinus, brain, eye, skin, and mucous membranes.	

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5. References and Informational Links

Books

- Bioaerosols: Assessment and Control. Janet Macher, Ed., American Conference of Governmental Industrial Hygienists, Cincinnati, OH 1999.
- Exposure Guidelines for Residential Indoor Air Quality. Environmental Health Directorate, Health Protection Branch, Health Canada, Ottawa, Ontario, 1989.
- Fungal Contamination in Public Buildings: Health Effects and Investigation Methods. Health Canada, Ottawa, Ontario, 2004.
- IICRC: S500 Standard and Reference Guide for Professional Water Damage Restoration. 3rd Edition, Institute of Inspection, Cleaning, and Restoration Certification, Vancouver, WA, 2006

  IICRC: S520 Standard and Reference Guide for Professional Mold Remediation. 1st Edition, Institute of Inspection, Cleaning, and Restoration Certification, Vancouver, WA, 2004
- Field Guide for the Determination of Biological Contaminants in Environmental Samples. 2nd Edition, American Industrial Hygiene Association, 2005.

Consumer Links

Read the full text of AIHA's "The Facts About Mold" consumer brochure.
https://aiha-assets.sfo2.digitaloceanspaces.com/AIHA/resources/Facts-About-Mold-A-Consumer-Focus-Fact-Sheet.pdf

The Occupational Safety and Health Administration (OSHA)
http://www.osha.gov/SLTC/molds/index.html

CDC Mold Facts
https://www.cdc.gov/mold-health/about/index.html?CDC_AAref_Val=https://www.cdc.gov/mold/faqs.htm

CDC Stachybotrys - Questions and answers on Stachybotrys chartarum and other molds
https://www.cdc.gov/mold-health/data-research/facts-stats/?CDC_AAref_Val=https://www.cdc.gov/mold/stachy.htm

IOM, NAS: Clearing the Air: Asthma and Indoor Air Exposures
https://www.epa.gov/indoor-air-quality-iaq/should-you-have-air-ducts-your-home-cleaned

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National Library of Medicine-Mold website
http://www.nlm.nih.gov/medlineplus/molds.html

California Department of Health Services (CADOHS)
https://www.cdph.ca.gov/Programs/cls/dehl/ehl/Pages/AQS/Mold.aspx

Minnesota Department of Health
https://www.health.state.mn.us/communities/environment/air/mold/index.html

New York City Department of Health and Mental Hygiene
https://www.nyc.gov/site/doh/health/health-topics/mold.page

EPA

"Should You Have the Air Ducts in Your Home Cleaned?"
https://www.epa.gov/indoor-air-quality-iaq/should-you-have-air-ducts-your-home-cleaned

General information about molds and actions that can be taken to clean up or prevent a mold problem.
https://www.epa.gov/mold/mold-cleanup-your-home

“A Brief Guide to Mold, Moisture, and Your Home” - Includes basic information on mold, cleanup guidelines, and moisture and mold prevention
https://www.epa.gov/mold/brief-guide-mold-moisture-and-your-home

“Mold Remediation in Schools and Commercial Buildings” - Information on remediation in schools and commercial property, references for potential mold and moisture remediators.
https://www.epa.gov/mold/mold-remediation-schools-and-commercial-buildings-guide

FEMA

“Homes That Were Flooded May Harbor Mold Problems” - Information and tips for cleaning mold.
https://www.fema.gov/press-release/20210318/fact-sheet-mold-problems-and-solutions

“Dealing With Mold & Mildew in Your Flood Damaged Home.
http://www.fema.gov/pdf/rebuild/recover/fema_mold_brochure_english.pdf

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6. Important Terms, Conditions, and Limitations

A. Sample Retention

Samples analyzed by EMSL will be retained for 60 days after analysis date Storage beyond this period is available for a fee with written request prior to the initial 30 day period. Samples containing hazardous/toxic substances which require special handling will be returned to the client immediately. EMSLreserves the right to charge a sample disposal fee or return samples to the client.

B. Change Orders and Cancellation

All changes in the scope of work or turnaround time requested by the client after sample acceptance must be made in writing and confirmed in writing by EMSL. If requested changes result in a change in cost the client must accept payment responsibility. In the event work is cancelled by a client, EMSL will complete work in progress and invoice for work completed to the point of cancellation notice. EMSL is not responsible for. holding times that are exceeded due to such changes.

C. Warranty

EMSL warrants to its clients that all services provided hereunder shall be performed in accordance with established and recognized analytical testing procedures and with reasonable care in accordance with applicable federal, state and local laws. The foregoing express warranty is exclusive and is given in lieu of all other warranties, expressed or implied. EMSL disclaims any other warranties, express or implied, including a warranty of fitness for particular purpose and warranty of merchantability.

D. Limits of Liability

In no event shall EMSL be liable for indirect, special, consequential, or incidental damages, including, but not limited to, damages for loss of profit or goodwill regardless of the negligence (either sole or concurrent) of EMSL and whether EMSL has been informed of the possibility of such damages, arising out of or in connection with EMSL's services thereunder or the delivery, use, reliance upon or interpretation of test results by client or any third party. We accept no legal responsibility for the purposes for which the client uses the test results. EMSL will not be held responsible for the improper selection of sampling devices even if we supply the device to the user. The user of the sampling device has the sole responsibility to select the proper sampler and sampling conditions to insure that a valid sample is taken for analysis. Any resampling performed will be at the sole discretion of EMSL, the cost of which shall be limited to the reasonable value of the original sample delivery group (SDG) samples. In no event shall EMSL be liable to a client or any third party, whether based upon theories

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of tort, contract or any other legal or equitable theory, in excess of the amount paid to EMSL by client thereunder.

E. Indemnification

Client shall indemnify EMSL and its officers, directors and employees and hold each of them harmless for any liability, expense or cost, including reasonable attorney's fees, incurred by reason of any third party claim in connection with EMSL services, the test result data or its use by client

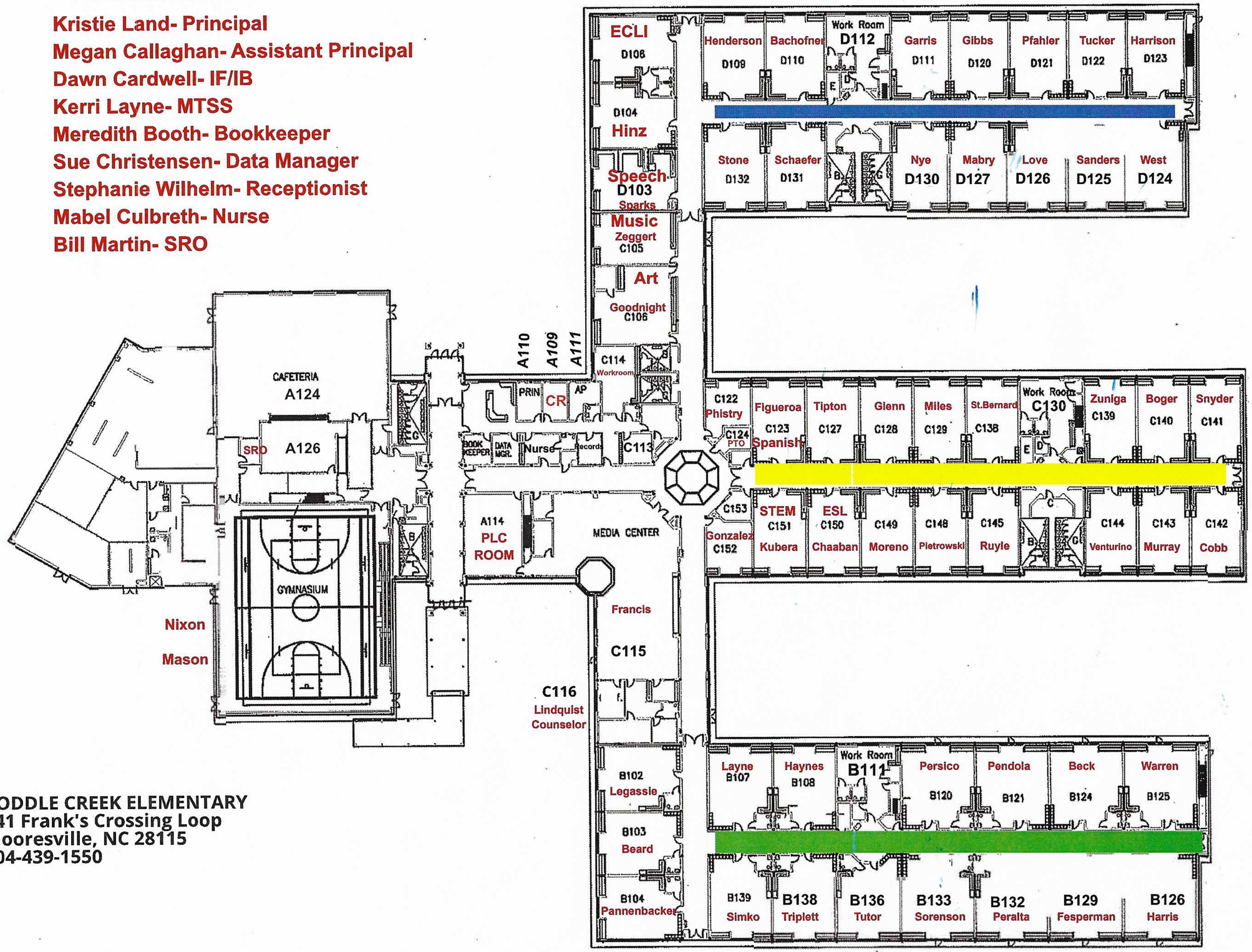
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ATTACHMENT 2

Building Floor Plan
(not to scale)

Administration

Kristie Land- Principal
Megan Callaghan- Assistant Principal
Dawn Cardwell- IF/IB
Kerri Layne- MTSS
Meredith Booth- Bookkeeper
Sue Christensen- Data Manager
Stephanie Wilhelm- Receptionist
Mabel Culbreth- Nurse
Bill Martin- SRO



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