



Cleaner environment. Safer workplaces.

**Mold and Indoor Air Quality Report
for**

New Hartford Elementary School
40 Wickett Street
New Hartford, CT 06057

-

Prepared For

New Hartford Public Schools
530 Main Street, P.O. Box 315
New Hartford, CT 06057

Date of Inspection and Testing:

March 12, 2026

EnviroMed Project

IH-26-226

EnviroMed Services, Inc.
470 Murdock Ave., Meriden, CT 06450
telephone (203) 238-4846 • facsimile (203) 238-4243

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I. PROJECT NARRATIVE

Overview

On March 12, 2026, EnviroMed Services conducted a microbial and indoor air quality assessment at New Hartford Elementary School on 40 Wickett Street, New Hartford, CT. The purpose of this assessment, as directed by the client, was to measure indoor air quality, and provide a baseline survey of the entire school for airborne fungal contamination.

Methods

The following sampling and assessment scheme was employed:

- Direct reading measurements were collected using a TSI Q-Trak™ model 8550/8551 IAQ Monitor. The baseline indoor air quality (IAQ) parameters temperature (°F), relative humidity (%), carbon monoxide (ppm), and carbon dioxide (ppm).
- Direct reading measurements for Volatile Organic Compounds (VOCs) were taken using a ppb RAE 3000 Photo Ionization Detector (PID).
- Direct reading measurements were taken for a total particulate in the air using an MIE PDR 1000 Personal Data Ram Particulate Analyzer and a Temtop LKC-1000 Series Air Quality Monitor
- Direct reading measurements for Formaldehyde were taken using a Temtop LKC-1000 Series Air Quality Monitor
- Direct reading measurements were collected using MultiRAE PGM-6228 Wireless Multi-Gas Monitor. Oxygen (O₂), Lower Explosive Limit (LEL)
- Conduct air sampling to test for the presence of airborne fungal structures, including spores, by drawing air through a Zefon Air-O-Cell cassette using a pre-calibrated vacuum pump. In this method, air enters the cassette, and particles are collected on the sampling substrate. The manufacturer-recommended flow rate of 15 liters per minute of air, for a duration of 10 minutes, was used to collect each sample. Sample locations were selected based on client direction. The samples were sent to Hayes Microbial Consulting in Midlothian, VA, for analysis by optical microscopy. Hayes Microbial Consulting is approved by the American Industrial Hygiene Association (AIHA Cert# 188863) and the state of Connecticut Department of Public Health. The results are represented in counts of fungal structures and fungal structures per cubic meter of air (spores/m³). The results are compared to the baseline sample that was collected outdoors.

Summary of Results:

Temperature: The indoor temperatures ranged from 65.6 to 76.7 °F. The ASHRAE recommended temperatures range from 68.0 to 74.0 °F. The indoor temperature readings were generally within above the recommended range.

Relative Humidity: The indoor relative humidity ranged from 18.4 to 48.0 percent. ASHRAE recommended range for indoor relative humidity is 20 to 60 percent. The relative humidity readings were within the recommended range.

Carbon Monoxide: The indoor carbon monoxide concentration did not exceed 0.0 ppm. The OSHA PEL limit for carbon monoxide is 50 ppm, reported as an 8-hour time-weighted average. The carbon monoxide readings were below the limit.

Particulates in Air (Dust): The indoor concentration of particulates in air inside the building did not exceed 4.7 ug/m³ for PM 2.5, and 8.3 ug/m³ for PM 10. The OSHA Permissible Exposure Limit (PEL) for respirable and total particulates in 5,000 ug/m³ and 15,000 ug/m³ respectively.

Volatile Organic Compounds (VOCs): The maximum concentration of Total VOCs per the Leadership in Energy and Environmental Design (LEED) Standards is 0.500 ppm. VOC readings ranged from 0.000 ppm to 0.258 ppm, which is generally within the standard. The minor levels above 0.500 ppm may be due to the presence of cleaning fluids, solvents, perfumes, or fragrance sprays.

Formaldehyde: Formaldehyde (CH₂O) levels did not exceed 0.07 ppm throughout the testing. This concentration is below the OSHA formaldehyde limit of 0.75 ppm, reported as an 8-hour time weighted average.

Carbon Dioxide: The outside carbon dioxide concentration recorded in the morning on this day was 426 ppm. The indoor concentrations during the morning ranged from 417 to 907 ppm. The outside carbon dioxide concentration recorded in the afternoon on this day was 464 ppm. The indoor concentrations ranged from 430 to 1081 ppm.

Per the ASHRAE Standard 62-2001, Ventilation for Acceptable Indoor Air Quality, the difference between the indoor and outdoor concentrations of carbon dioxide should be less than 707 ppm. The difference between the highest indoor concentration and the outdoor concentration in the morning was 481 ppm. The difference between the highest indoor concentration and the outdoor concentration in the afternoon was 617 ppm. The averages of the morning and afternoon carbon dioxide levels were within ASHRAE guidelines. Additionally, carbon dioxide concentrations in the morning and afternoon were well below the OSHA permissible exposure limit of 5000 ppm.

Oxygen (O₂): The indoor oxygen concentrations ranged from 21.4 to 21.6 percent. OSHA standard for oxygen concentration is 19.6 to 23.4 percent. The indoor concentration readings were all within the recommended range.

Lower Explosive Limit (LEL): The indoor lower explosive limit ranged from 0 to 0 percent. OSHA states that atmospheres with flammable vapors at or above 10 percent are considered hazardous. The lower explosive limit readings were below the hazardous limits.

Section III of this report contains the IAQ data results.

Air-O-Cell Spore Trap Sampling Results:

There are currently no regulatory standards for concentrations of mold in air under OSHA. Levels of mold within the building are routinely compared to the baseline levels outside the building to detect any amplifications or unusual toxigenic mold species.

The Air Sample results indicated fungal amplification of allergenic mold in the Library.

Section IV of this report contains a copy of the analytical report.

Ascospores: A large group consisting of more than 3000 species of fungi. Common plant pathogens and outdoor numbers become very high following rain. Most of the genera are indistinguishable by spore trap analysis and are combined on the report.

Aspergillus | Penicillium: The most common fungi isolated from the environment. Very common in soil and on decaying plant material. Can grow well indoors on a wide variety of substrates. This group contains common allergens, and many can cause hypersensitivity pneumonitis. They may cause extrinsic asthma, and many are opportunistic pathogens. Many species produce mycotoxins which may be associated with disease in humans and other animals. Toxin production is dependent on the species, the food source, competition with other organisms, and other environmental conditions.

Basidiospores: A common group of Fungi that includes mushrooms and bracket fungi. They are saprophytes and plant pathogens. In wet conditions, they can cause structural damage to buildings. These are common allergens and are also associated with hypersensitivity pneumonitis.

Cladosporium: One of the most common genera worldwide. Found in soil and plant debris and on the leaf surfaces of living plants. The outdoor numbers are lower in the winter and often relatively high in the summer, especially in high humidity. The outdoor numbers often spike in the late afternoon and evening. Indoors, it can be found growing on textiles, wood, sheetrock, moist windowsills and in HVAC supply ducts. A common allergen, producing more than 10 allergenic antigens and a common cause of hypersensitivity pneumonitis.

Culvularia: They exist in soil and plant debris, and are plant pathogens. They are allergenic and a common cause of allergic fungal sinusitis. An occasional cause of human infection, including keratitis, sinusitis, onychomycosis, mycetoma, pneumonia, endocarditis and disseminated infection, primarily in the immunocompromised.

Myxomycetes: Found on decaying plant material and as a plant pathogen. Some allergenic properties reported, but generally pose no health concerns to humans.

Sample ID: A1**Sample Location: Outside Baseline**

The prominent fungal genus present was *Ascospores*. The other fungal genera present were *Aspergillus/Penicillium*, *Basidiospores*, and *Cladosporium*.

Sample ID: A2**Sample Location: Main Office**

The prominent fungal genus present was *Ascospores*. The other fungal genera present were *Aspergillus/Penicillium*, *Cladosporium*, and *Myxomycetes*. There was no amplification detected for any of these fungal genera as compared to the outdoor baseline sample.

Sample ID: A3**Sample Location: Room 9**

The prominent fungal genus present was *Ascospores*. The other fungal genera present were *Aspergillus/Penicillium* and *Culvularia*. There was no amplification detected for any of these fungal genera as compared to the outdoor baseline sample.

Sample ID: A4**Sample Location: Library**

The prominent fungal genus present was *Aspergillus/Penicillium*. *Aspergillus/Penicillium* was detected with a spore count significantly higher than the outdoor baseline sample, indicating an amplification of this fungi. *Aspergillus/Penicillium* is considered an allergenic mold.

The other fungal genera present were *Ascospores* and *Cladosporium*. There was no amplification detected for any of these fungal genera as compared to the outdoor baseline sample.

Sample ID: A5**Sample Location: Room 2**

The prominent fungal genus present was *Ascospores*. The other fungal genera present were *Aspergillus/Penicillium* and *Basidiospores*. There was no amplification detected for any of these fungal genera as compared to the outdoor baseline sample.

Sample ID: A6**Sample Location: Room 5**

The prominent fungal genus present was *Ascospores*. The other fungal genera present were *Aspergillus/Penicillium* and *Cladosporium*. There was no amplification detected for any of these fungal genera as compared to the outdoor baseline sample.

Sample ID: A7**Sample Location: Room 25**

The prominent fungal genus present was *Ascospores*. The other fungal genera present were *Aspergillus/Penicillium* and *Basidiospores*. There was no amplification detected for any of these fungal genera as compared to the outdoor baseline sample.

Sample ID: A8**Sample Location: Room 27 Nurse's Office**

The prominent fungal genus present was *Ascospores*. The other fungal genus present was *Aspergillus/Penicillium*. There was no amplification detected for any of these fungal genera as compared to the outdoor baseline sample.

Sample ID: A9**Sample Location: Staff Workroom**

The prominent fungal genus present was *Ascospores*. The other fungal genera present were *Aspergillus/Penicillium* and *Cladosporium*. There was no amplification detected for any of these fungal genera as compared to the outdoor baseline sample.

Sample ID: A10**Sample Location: Room 4**

The prominent fungal genus present was *Ascospores*. The other fungal genera present were *Aspergillus/Penicillium*, *Basidiospores*, and *Myxomycetes*. There was no amplification detected for any of these fungal genera as compared to the outdoor baseline sample.

Conclusions and Recommendations

Based on the results obtained and observations made during the assessment, EnviroMed Services, Inc. has come to the following conclusions and recommendations.

Conclusions:

Indoor Air Quality:

Based on the results obtained and observations made during the assessment, EnviroMed Services, Inc. has come to the following conclusions and recommendations.

- The indoor air temperatures throughout the school were generally within the recommended ASHRAE standard range at the time of the assessment.
- All the indoor relative humidity readings throughout the school were within the recommended ASHRAE standard range at the time of the assessment.
- The indoor carbon monoxide concentrations throughout the school were all well below the OSHA recommended limit at the time of the assessment.
- All the indoor carbon dioxide concentrations throughout the school were generally within the ASHRAE recommended limit and below the OSHA permissible exposure limit at the time of the assessment.
- The concentration of particulates in air inside the building were well below the OSHA PEL at the time of the assessment.
- The concentration of formaldehyde inside the building was well below the OSHA PEL at the time of the assessment.
- The oxygen concentrations inside the building were within the OSHA standard at the time of the assessment
- The indoor lower explosive limit was below the hazardous limits at the time of the assessment.

Mold:

Air Samples:

- The air sample results indicated fungal amplification of allergenic mold in the Library.

Recommendations:

EnviroMed Services, Inc. recommends that a professional remediation contractor clean all cleanable nonporous surfaces with a disinfecting agent such as Fiberlock ShockWave or equivalent for all areas of concern within the following locations: Library; and to remove and replace any porous water or moisture impacted materials that cannot be appropriately cleaned or dried. In addition, EnviroMed Services, Inc. recommends the use of a HEPA filtered air filtration system to remove all airborne spores as amplification that were detected throughout all affected areas. The Professional Mold Remediation contractor hired will provide the specific means and methods to remediate for the best course of action as approved by the Client. Some General Recommendations for reference are as follows:

We recommend the use of a HEPA (High-Efficiency Particulate Air) vacuum. Cleaning will also need to be performed on any non-movable items located in the unit, movable items will need to be removed prior to the start of remediation. Carpeting should also be HEPA vacuum cleaned with a disinfecting agent or removed and replaced.

Prior to cleaning, Negative Air Pressure Containment should be set up to prevent the contamination of any airborne mold spores during the remediation process. Containment is to be exhausted outside the containment area during remediation. The HEPA Sandwich method, as referenced in mold remediation under the IICRC/ANSI S520 Standard, involves a three-step cleaning process designed to remove mold contamination from surfaces effectively. This method consists of Initial HEPA vacuuming, Damp wiping & Final HEPA vacuuming or additional damp wiping. The HEPA Sandwich method is recommended with the continued use of an industrial HEPA Air Unit for at least 24-48 hours post-abatement work to further clean the air.

Following the remediation, EnviroMed recommends a follow-up clearance microbial assessment be performed by a professional industrial hygienist to verify the effectiveness of the cleaning following remediation.

II. PHOTOGRAPHS



Figure 1: Sample A-1 – Outdoor Baseline



Figure 2: Sample A-2 – Main Office

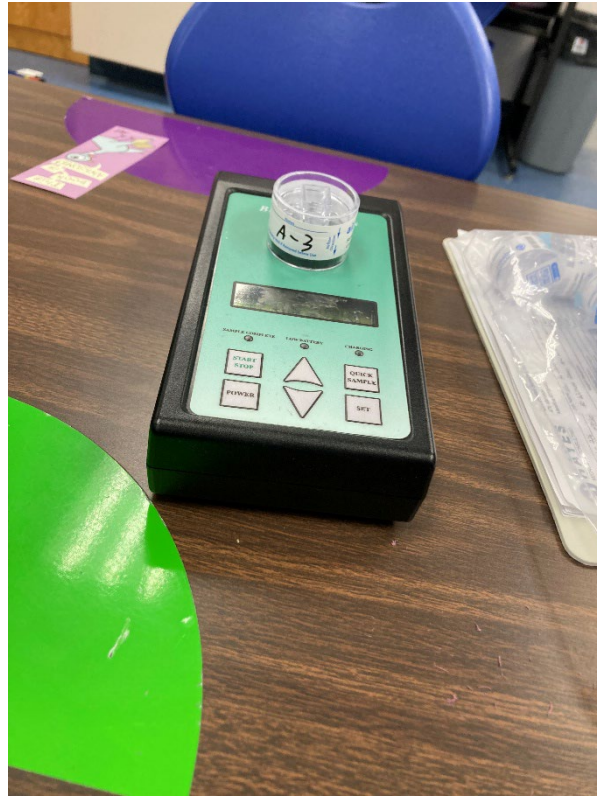


Figure 3: Sample A-3 – Room 9



Figure 4: Sample A-4 – Library

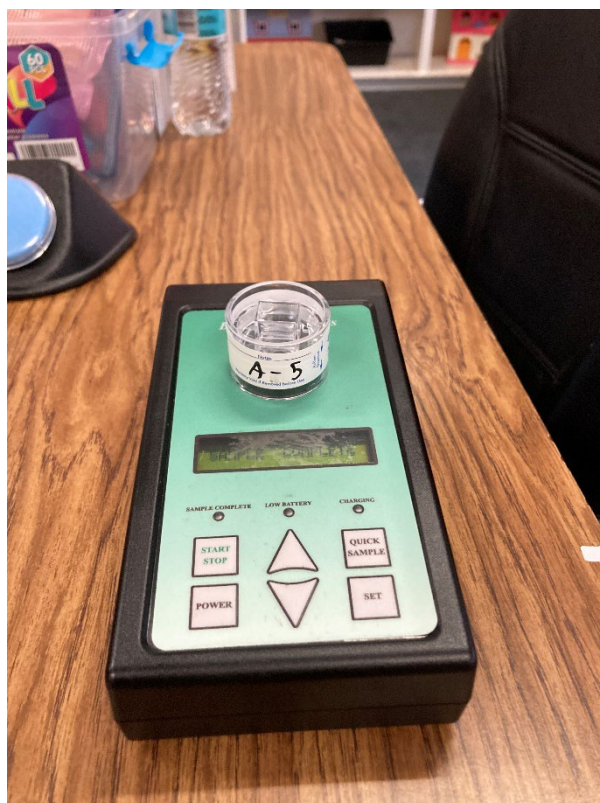


Figure 5: Sample A-5 – Room 2



Figure 6: Sample A-6 – Room 5



Figure 7: Sample A-7 – Room 25



Figure 8: Sample A-8 – Room 27 - Nurse's Office



Figure 9: Sample A-9 – Staff Workroom



Figure 10: Sample A-10 – Room 4

III . TABLE OF IAQ MEASUREMENTS

A.M. Readings

Location	Type	Temp °F	RH %	CO (ppm)	CO ₂ (ppm)	VOCs (ppm)	PM _{2.5} (ug/m ³)	PM ₁₀ (ug/m ³)	CH ₂ O (mg/m ³)	OXY	LEL %	Occupied
Outdoor Baseline	Outside	45.2	44.3	0.0	426	0.000	0.3	0.8	0.01	21.4	0	-
Main Office	Office	65.6	48.0	0.0	566	0.034	1.0	1.9	0.01	21.4	0	Yes
Office Extension	Office	69.0	42.3	0.0	526	0.036	1.1	1.5	0.01	21.4	0	No
Custodian Office	Office	72.5	37.5	0.0	417	0.000	0.8	1.4	0.01	21.4	0	No
Workroom	Office	73.1	36.0	0.0	575	0.021	0.9	1.3	0.01	21.4	0	Yes
Staff Lounge	Office	73.1	36.6	0.0	664	0.034	1.4	2.5	0.01	21.4	0	No
Room 6	Classroom/ Office	72.8	34.9	0.0	496	0.043	1.1	2.0	0.01	21.4	0	No
Room 7	Classroom	72.0	28.6	0.0	526	0.050	0.5	0.9	0.01	21.4	0	Yes
Room 8	Classroom	72.0	36.3	0.0	567	0.051	1.1	1.8	0.01	21.4	0	No
Room 9	Classroom	72.9	35.4	0.0	679	0.072	1.1	2.1	0.02	21.4	0	Yes
Room 10	Classroom	68.7	37.7	0.0	660	0.280	1.3	2.5	0.02	21.5	0	No
Room 11	Library	70.8	36.6	0.0	585	0.075	1.4	2.1	0.03	21.4	0	Yes
Library Office	Office	70.8	36.1	0.0	754	0.258	1.5	2.7	0.03	21.5	0	No
Room 1	Classroom	73.1	34.1	0.0	583	0.095	0.9	1.5	0.01	21.4	0	No
Room 2	Classroom	72.8	34.8	0.0	803	0.142	1.6	2.6	0.02	21.4	0	Yes
Room 3	Classroom	73.2	36.6	0.0	907	0.123	1.1	2.0	0.02	21.4	0	Yes
Room 4	Classroom	72.6	35.6	0.0	796	0.140	1.2	1.5	0.02	21.4	0	Yes
Room 5	Classroom	72.2	30.9	0.0	653	0.173	0.9	1.7	0.01	21.4	0	Yes

Location	Type	Temp °F	RH %	CO (ppm)	CO ₂ (ppm)	VOCs (ppm)	PM _{2.5} (ug/m ³)	PM ₁₀ (ug/m ³)	CH ₂ O (mg/m ³)	OXY	LEL %	Occupied
Room 20 Sensory Room	Classroom	72.4	25.7	0.0	450	0.146	0.8	1.3	0.01	21.5	0	No
Room 21 Sensory Room B	Classroom	74.4	27.3	0.0	515	0.151	0.3	0.5	0.01	21.5	0	No
Kitchen/Serving	Kitchen	75.0	29.1	0.0	612	0.178	0.5	0.9	0.01	21.5	0	No
Room 25 Speech	Classroom	74.5	27.1	0.0	455	0.175	1.0	1.7	0.01	21.5	0	No
Room 24 Speech Office	Office	75.9	28.5	0.0	516	0.192	0.5	0.9	0.01	21.5	0	No
Room 26 Principal	Office	76.5	28.6	0.0	535	0.195	1.3	2.3	0.01	21.5	0	No
Room 27 Nurse Office	Office	76.7	28.4	0.0	540	0.207	0.9	1.5	0.01	21.5	0	Yes
Gymateria	Gym/Cafe	72.9	30.7	0.0	524	0.206	0.9	1.5	0.01	21.5	0	No
Room 23 Mr. Ross	Office	71.9	33.3	0.0	674	0.214	1.2	1.9	0.01	21.5	0	Yes
Gym Storage/Office	Office	70.1	34.0	0.0	541	0.212	1.2	2.1	0.02	21.5	0	No

P.M. Readings

Location	Type	Temp °F	RH %	CO (ppm)	CO ₂ (ppm)	VOCs (ppm)	PM _{2.5} (ug/m ³)	PM ₁₀ (ug/m ³)	CH ₂ O (mg/m ³)	OXY	LEL %	Occupied
Outdoor Baseline	Outside	45.9	22.0	0.0	464	0.000	0.9	1.5	0.02	21.6	0	-
Main Office	Office	68.2	45.2	0.0	706	0.053	1.6	2.9	0.02	21.5	0	Yes
Office Extension	Office	69.8	33.1	0.0	776	0.059	1.5	2.3	0.06	21.5	0	No
Custodian Office	Office	72.0	33.1	0.0	583	0.028	2.0	2.7	0.06	21.5	0	No
Workroom	Office	73.1	31.7	0.0	817	0.140	2.6	4.4	0.05	21.5	0	No
Staff Lounge	Office	73.5	32.0	0.0	941	0.164	4.7	8.3	0.07	21.5	0	No
Room 6	Classroom/ Office	73.7	29.4	0.0	769	0.113	2.0	3.8	0.07	21.5	0	Yes
Room 7	Classroom	72.6	20.4	0.0	622	0.097	1.0	1.9	0.05	21.5	0	Yes
Room 8	Classroom	73.0	32.3	0.0	820	0.069	1.3	2.4	0.02	21.5	0	Yes
Room 9	Classroom	72.8	30.7	0.0	1081	0.127	0.9	1.3	0.04	21.5	0	Yes
Room 10	Classroom	71.3	30.2	0.0	806	0.098	1.4	2.3	0.05	21.5	0	Yes
Room 11	Library	70.3	31.6	0.0	905	0.127	1.4	2.1	0.05	21.5	0	Yes
Library Office	Office	70.0	32.5	0.0	843	0.102	1.6	2.9	0.06	21.5	0	No
Room 1	Classroom	72.2	24.7	0.0	528	0.092	1.3	2.2	0.04	21.5	0	Yes
Room 2	Classroom	71.3	25.4	0.0	706	0.133	1.3	2.4	0.03	21.5	0	Yes
Room 3	Classroom	72.4	29.3	0.0	970	0.134	2.1	4.5	0.05	21.5	0	Yes
Room 4	Classroom	73.0	26.1	0.0	715	0.193	1.4	2.4	0.04	21.6	0	Yes
Room 5	Classroom	72.3	25.8	0.0	878	0.096	1.7	2.8	0.04	21.5	0	Yes

Room 20 Sensory Room	Classroom	73.0	18.4	0.0	430	0.082	2.2	4.0	0.01	21.6	0	No
Room 21 Sensory Room B	Classroom	74.4	21.1	0.0	490	0.083	1.5	2.5	0.01	21.6	0	No
Kitchen/Serving	Kitchen	75.4	26.4	0.0	701	0.118	2.0	3.3	0.01	21.6	0	No
Room 25 Speech	Classroom	75.8	20.3	0.0	608	0.104	1.9	2.7	0.02	21.6	0	Yes
Room 24 Speech Office	Office	75.8	19.4	0.0	480	0.100	0.7	1.7	0.01	21.6	0	No
Room 26 Principal	Office	76.3	21.7	0.0	563	0.125	1.4	2.3	0.01	21.6	0	No
Room 27 Nurse Office	Office	75.2	22.2	0.0	596	0.132	1.6	2.6	0.02	21.6	0	No
Gymateria	Gym/Cafe	72.0	28.4	0.0	706	0.160	1.9	3.8	0.03	21.6	0	No
Room 23 Mr. Ross	Office	71.5	29.8	0.0	744	0.163	3.9	7.3	0.05	21.6	0	No
Gym Storage/Office	Office	70.5	30.5	0.0	704	0.166	3.7	6.6	0.05	21.6	0	No

IV. LABORATORY ANALYSIS REPORT



#26011884

Analysis Report prepared for

EnviroMed Services, Inc.

470 Murdock Avenue
Meriden, CT 06450

Phone: (203) 238-4846

IH-26-226
New Hartford Elementary School
40 Wickett Street
New Hartford, CT 06057

Collected: **March 12, 2026**
Received: **March 13, 2026**
Reported: **March 13, 2026**

We would like to thank you for trusting Hayes Microbial for your analytical needs!
We received 10 samples by FedEx in good condition for this project on March 13th, 2026.

The results in this analysis pertain only to this job, collected on the stated date, and should not be used in the interpretation of any other job. Information supplied by the customer can affect the validity of results. These results apply only to the samples as received. This report may not be duplicated, except in full, without the written consent of Hayes Microbial Consulting, LLC.

All information provided to Hayes Microbial is confidential information relating to our customers and their clients. We will not disclose, copy, or distribute any information verbally or written, except to those designated by the customer(s). We take confidentiality very seriously. No changes to the distribution list will be made without the express consent of the customer.

This laboratory bears no responsibility for sample collection activities, analytical method limitations, or your use of the test results. Interpretation and use of test results are your responsibility. Any reference to health effects or interpretation of mold levels is strictly the opinion of Hayes Microbial. In no event, shall Hayes Microbial or any of its employees be liable for lost profits or any special, incidental or consequential damages arising out of the use of these test results.

Steve Hayes, BSMT (ASCP)
Laboratory Director
Hayes Microbial Consulting, LLC.



EPA Laboratory ID: VA01419



Lab ID: #188863



DPH License: #PH-0198

Sample Number*	1	A - 1		2	A - 2		3	A - 3		4	A - 4	
Sample Name*	Outdoor Baseline			Main Office			Room 9			Library		
Sample Volume*	150 L			150 L			150 L			150 L		
Reporting Limit	7 spores/m³			7 spores/m³			7 spores/m³			7 spores/m³		
Background	2			2			2			2		
Fragments	ND			ND			ND			7/m³		
Organism	Raw Count	Count / m³	% of Total	Raw Count	Count / m³	% of Total	Raw Count	Count / m³	% of Total	Raw Count	Count / m³	% of Total
Alternaria												
Ascospores	19	130	55.9%	4	27	40.0%	2	13	40.0%	4	27	3.4%
Aspergillus Penicillium	3	20	8.8%	3	20	30.0%	2	13	40.0%	112	750	95.7%
Basidiospores	7	47	20.6%									
Bipolaris Drechslera												
Chaetomium												
Cladosporium	5	33	14.7%	1	7	10.0%				1	7	<1%
Curvularia							1	7	20.0%			
Epicoccum												
Fusarium												
Memnoniella												
Myxomycetes/Periconia				2	13	20.0%						
Pithomyces												
Rusts/Smuts												
Stachybotrys												
Stemphylium												
Torula												
Ulocladium												
Total	34	230	100%	10	67	100%	5	33	100%	117	784	100%

Water Damage Indicator	Common Allergen	Slightly Higher than Baseline	Significantly Higher than Baseline	Ratio Abnormality
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* indicates data provided by the customer



Collected: **Mar 12, 2026**

Received: **Mar 13, 2026**

Reported: **Mar 13, 2026**

Project Analyst:
 Steve Hayes, BSMT *Stephen N. Hayes*

Date:
03 - 13 - 2026

Reviewed By:
 Ramesh Poluri, PhD *P. Ramesh*

Date:
03 - 13 - 2026

Sample Number*	5	A - 5		6	A - 6		7	A - 7		8	A - 8	
Sample Name*	Room 2			Room 5			Room 25			Room 27 Nurse Office		
Sample Volume*	150 L			150 L			150 L			150 L		
Reporting Limit	7 spores/m³			7 spores/m³			7 spores/m³			7 spores/m³		
Background	2			2			2			2		
Fragments	ND			ND			ND			ND		
Organism	Raw Count	Count / m³	% of Total	Raw Count	Count / m³	% of Total	Raw Count	Count / m³	% of Total	Raw Count	Count / m³	% of Total
Alternaria												
Ascospores	5	33	50.0%	4	27	40.0%	3	20	60.0%	2	13	66.7%
Aspergillus Penicillium	3	20	30.0%	4	27	40.0%	1	7	20.0%	1	7	33.3%
Basidiospores	2	13	20.0%				1	7	20.0%			
Bipolaris Drechslera												
Chaetomium												
Cladosporium				2	13	20.0%						
Curvularia												
Epicoccum												
Fusarium												
Memnoniella												
Myxomycetes/Periconia												
Pithomyces												
Rusts/Smuts												
Stachybotrys												
Stemphylium												
Torula												
Ulocladium												
Total	10	66	100%	10	67	100%	5	34	100%	3	20	100%

Water Damage Indicator	Common Allergen	Slightly Higher than Baseline	Significantly Higher than Baseline	Ratio Abnormality
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* indicates data provided by the customer



Collected: **Mar 12, 2026**

Received: **Mar 13, 2026**

Reported: **Mar 13, 2026**

Project Analyst:
 Steve Hayes, BSMT *Stephen N. Hayes*

Date:
03 - 13 - 2026

Reviewed By:
 Ramesh Poluri, PhD *P. Ramesh*

Date:
03 - 13 - 2026

Sample Number*	9	A - 9		10	A - 10			
Sample Name*	Staff Workroom			Room 4				
Sample Volume*	150 L			150 L				
Reporting Limit	7 spores/m³			7 spores/m³				
Background	2			2				
Fragments	ND			ND				
Organism	Raw Count	Count / m³	% of Total	Raw Count	Count / m³	% of Total		
Alternaria								
Ascospores	3	20	50.0%	6	40	46.2%		
Aspergillus Penicillium	2	13	33.3%	5	33	38.5%		
Basidiospores				1	7	7.7%		
Bipolaris Drechslera								
Chaetomium								
Cladosporium	1	7	16.7%					
Curvularia								
Epicoccum								
Fusarium								
Memnoniella								
Myxomycetes Periconia				1	7	7.7%		
Pithomyces								
Rusts Smuts								
Stachybotrys								
Stemphylium								
Torula								
Ulocladium								
Total	6	40	100%	13	87	100%		

Water Damage Indicator	Common Allergen	Slightly Higher than Baseline	Significantly Higher than Baseline	Ratio Abnormality
------------------------	-----------------	-------------------------------	------------------------------------	-------------------

* indicates data provided by the customer

Spore Trap Information

Reporting Limit	The Reporting Limit is the lowest number of spores that can be detected based on the total volume of the sample collected and the percentage of the slide that is counted. At Hayes Microbial, 100% of the slide is read so the LOD is based solely on the total volume. Raw spore counts that exceed 500 spores will be estimated.
Blanks	Results have not been corrected for field or laboratory blanks.
Background	The Background is the amount of debris that is present in the sample. This debris consists of skin cells, dirt, dust, pollen, drywall dust and other organic and non-organic matter. As the background density increases, the likelihood of spores, especially small spores such as those of <i>Aspergillus</i> and <i>Penicillium</i> may be obscured. The background is rated on a scale of 1 to 5 and each level is determined as follows: NBD: No background detected due to possible pump or cassette malfunction. Recollect sample. (Field Blanks will display NBD) 1 : <5% of field occluded. No spores will be uncountable. 2 : 5-25% of field occluded. 3 : 25-75% of field occluded. 4 : 75-90% of field occluded. 5 : >90% of field occluded. Suggested recollection of sample.
Fragments	Fragments are small pieces of fungal mycelium or spores. They are not identifiable as to type and when present in very large numbers, may indicate the presence of mold amplification.
Control Comparisons	There are no national standards for the numbers of fungal spores that may be present in the indoor environment. As a general rule and guideline that is widely accepted in the indoor air quality field, the numbers and types of spores that are present in the indoor environment should not exceed those that are present outdoors at any given time. There will always be some mold spores present in "normal" indoor environments. The purpose of sampling and counting spores is to help determine whether an abnormal condition exists within the indoor environment and if it does, to help pinpoint the area of contamination. Spore counts should not be used as the sole determining factor of mold contamination. There are many factors that can cause anomalies in the comparison of indoor and outdoor samples due to the dynamic nature of both of those environments.
Water Damage Indicator	Blue: These molds are commonly seen in conditions of prolonged water intrusion and usually indicate a problem.
Common Allergen	Green: Although all molds are potential allergens, these are the most common allergens that may be found indoors.
Slightly Higher than Baseline	Orange: The spore count is slightly higher than the outside count and may or may not indicate a source of contamination.
Significantly Higher than Baseline	Red: The spore count is significantly higher than the baseline count and probably indicates a source of contamination.
Ratio Abnormality	Violet: The types of spores found indoors should be similar to the ones that were identified in the baseline sample. Significant increases (more than 25%) in the ratio of a particular spore type may indicate the presence of abnormal levels of mold, even if the total number of spores of that type is lower in the indoor environment than it was outdoors.
Color Coding	Fungi that are present in indoor samples at levels lower than 200 per cubic meter are not color coded on the report, unless they are one of the water damage indicators.
Significant Figures	Raw counts and column totals may reflect more than 2 significant figures, but results should only be considered significant to 2 figures.

Analyte Descriptions

Ascospores	Habitat:	A large group consisting of more than 3000 species of fungi. Common plant pathogens and outdoor numbers become very high following rain. Most of the genera are indistinguishable by spore trap analysis and are combined on the report.
	Health Effects:	Health affects are poorly studied, but many are likely to be allergenic.
Aspergillus Penicillium	Habitat:	The most common fungi isolated from the environment. Very common in soil and on decaying plant material. Are able to grow well indoors on a wide variety of substrates.
	Health Effects:	This group contains common allergens and many can cause hypersensitivity pneumonitis. They may cause extrinsic asthma, and many are opportunistic pathogens. Many species produce mycotoxins which may be associated with disease in humans and other animals. Toxin production is dependent on the species, the food source, competition with other organisms, and other environmental conditions.
Basidiospores	Habitat:	A common group of Fungi that includes the mushrooms and bracket fungi. They are saprophytes and plant pathogens. In wet conditions they can cause structural damage to buildings.
	Health Effects:	Common allergens and are also associated with hypersensitivity pneumonitis.
Cladosporium	Habitat:	One of the most common genera worldwide. Found in soil and plant debris and on the leaf surfaces of living plants. The outdoor numbers are lower in the winter and often relatively high in the summer, especially in high humidity. The outdoor numbers often spike in the late afternoon and evening. Indoors, it can be found growing on textiles, wood, sheetrock, moist window sills and in HVAC supply ducts.
	Health Effects:	A common allergen, producing more than 10 allergenic antigens and a common cause of hypersensitivity pneumonitis.
Curvularia	Habitat:	They exist in soil and plant debris, and are plant pathogens.
	Health Effects:	They are allergenic and a common cause of allergic fungal sinusitis. An occasional cause of human infection, including keratitis, sinusitis, onychomycosis, mycetoma, pneumonia, endocarditis and disseminated infection, primarily in the immunocompromised.
Myxomycetes/Periconia	Description:	Found on decaying plant material and as a plant pathogen.
	Health Effects:	Some allergenic properties reported, but generally pose no health concerns to humans.



Cleaner environment. Safer workplaces.

Mold Inspection & Moisture Report

For

New Hartford Elementary School
40 Wickett St, Pine Meadow, CT 06061

LIBRARY

Prepared For

New Hartford Public Schools 530 Main
Street
New Hartford, CT

Date of Inspection:

July 10, 2026

EnviroMed Project

IH-26-2462

EnviroMed Services, Inc.
470 Murdock Ave., Meriden, CT 06450
telephone (203) 238-4846 • facsimile (203) 238-4243

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I. PROJECT NARRATIVE

Overview

On July 10, 2026, EnviroMed Services, Inc. conducted a Mold Inspection & moisture inspection at New Hartford Elementary School, New Hartford, CT. The purpose of this assessment, as directed by the client, was to survey for airborne and surface fungal contamination within the Library.

Methods

The following sampling and assessment scheme was employed:

- Conduct air sampling to test for the presence of airborne fungal structures, including spores, by drawing air through a Zefon Air-O-Cell cassette using a pre-calibrated vacuum pump. In this method, air enters the cassette and particles are collected on the sampling substrate. The manufacturer-recommended flow rate of 15 liters per minute of air, for a duration of 10 minutes, was used to collect each sample. Sample locations were selected based on client direction. The samples were sent to Hayes Microbial Consulting in Midlothian, VA, for analysis by optical microscopy. Hayes Microbial Consulting is approved by the American Industrial Hygiene Association (AIHA Cert# 188863) and the State of Connecticut Department of Public Health. The results are represented in counts of fungal structures and fungal structures per cubic meter of air (spores/m³). The results are compared to the Clearance sample that was collected outdoors.
- Conduct surface sampling to check for the presence of fungal structures, including spores, by using Bio-Tape adhesive glass slides. The sample locations were selected based on visual observations of potential mold and accessibility. The purpose of this sampling was to lift any spore colonies present from the surface so they can be identified, and relative growth density can be quantified. The sample was sent to Hayes Microbial Consulting in Midlothian, VA, for analysis by optical microscopy. The results of this sampling are reported by spore estimate by genus type along with Mycelial Estimate for determining probable growth.
- Monitoring for moisture was performed using a General Tools MM8 Pinless Moisture Meter. The scan function of the meter was used, which measures readings in REL (relative content). REL – Readings on the relative content are used for wallboard and wood. The REL value is a theoretical moisture level indicating potential severity of the moisture content with the material ranging from 0 to 100%. The dry area is below 25%, moderately wet area is between 26% and 75% and excessively wet is greater than 76%.

Summary of Results

Analyte Descriptions

Acronium: Found in soil and decaying plant debris. This species is a type I and Type III allergen. It may cause mycetoma, infections of the nails, corneal ulcers, meningitis, and endocarditis.

Ascospores: A Large group consisting of more than 3000 species of fungi. Common plant pathogens and outdoor numbers become very high following rain. Most of the genera are indistinguishable by spore trap analysis and are combined on the report. Health effects are poorly studied, but many are likely to be allergenic.

Aspergillus/Penicillium: The most common fungi isolated from the environment. Very common in soil and on decaying plant material. Are able to grow well indoors on a wide variety of substrates. This group contains common allergens, and many can cause hypersensitive pneumonitis.

Basidiospores: A common group of fungi that includes the mushroom and bracket fungi. They are saprophytes and plant pathogens. In wet conditions, they can cause structural damage to buildings. Effects are that they are common allergens and are also associated with hypersensitive pneumonitis.

Cladosporium: A common genus worldwide. Found in soil and plant debris and on the leaf surfaces of living plants. Indoors, it can be found growing on textiles, wood, sheetrock, moist windowsills and in HVAC supply ducts. It is a common allergen, producing more than 10 allergenic antigens and a common cause of hypersensitivity pneumonitis.

Curvularia: Found in soil and plant debris and are plant pathogens. This species is allergenic and a common cause of allergic fungal sinusitis. They are an occasional cause for infections in human that result in keratitis, sinusitis, onychomycosis, mycetoma, pneumonia, endocarditis, and disseminated infection.

Myxomycetes: Found on decaying plant material and as a plant pathogen. Some allergenic properties are reported, but generally pose no health concerns to humans.

Stachybotrys: Commonly found in soil and on decaying plant material. Allergenic properties are poorly studied and no cases of infection have been reported in humans. They do however produce potent tricothecene mycotoxins. The toxins produced by this fungus can suppress the immune system affecting the lymphoid tissue and the bone marrow. The mycotoxin is also reported to be a liver and kidney carcinogen.

Air-O-Cell Spore Trap Sampling Results:

There are currently no federal or State of Connecticut regulatory standards for concentrations of mold in air. However, some commonly used guidelines have been established. Levels of mold within the building are routinely compared to the Baseline levels outside the building to detect any amplifications or unusual mold species as compared to the natural airborne conditions of that day.

Section III of this report contains a copy of the analytical report.

Sample ID: A1

Sample Location: Outdoor Baseline

The prominent fungal genera present was *Cladosporium*. The other fungal genera present were *Alternaria*, *Ascospores*, *Basidiospores*, and *Epicoccum*.

Sample ID: A2

Sample Location: Library

The only fungal genera present was *Cladosporium*. There was no amplification detected for the fungal genera as compared to the outdoor baseline sample.

Sample ID: A3

Sample Location: Library (2)

The prominent fungal genera present was *Cladosporium*. The other fungal genera present was *Myxomycetes/Periconia*.

Sample ID: A4

Sample Location: Library (3)

The only fungal genera present was *Ascospores*.

Bio-Tape Spore Trap Sampling Results:

Sample ID: T1

Sample Location: Library Windowsill

There was no fungi detected.

Conclusions and Findings

Based on the results obtained and observations made during the assessment, EnviroMed Services, Inc. has come to the following conclusions.

Conclusions

Air Samples

- There is no evidence of airborne toxigenic mold in the library.
- There is no evidence of airborne mold contamination or mold amplification in the library.

Bio-Tape Sample

- There is no evidence of surface mold contamination on the library windowsill.

Based on our review of the airborne mold samples and surface tape lift sample collected within the Library at New Hartford Elementary School on July 10, 2026, EnviroMed Services, Inc. has determined that the areas tested are fit for occupancy. Laboratory analysis identified no evidence of airborne toxigenic mold, mold amplification or surface mold contamination within the areas evaluated.

II. PHOTOGRAPHS



Figure 1. Photograph of the Outdoor Baseline sample, A1.



Figure 2. Photograph of the Library sample, A2.



Figure 3. Photograph of the Library sample, A3.



Figure 4. Photograph of the Library sample, A4.



Figure 5. Photograph of the Library Windowsill sample, T1.

III. LABORATORY ANALYSIS REPORT

Analysis Report prepared for

EnviroMed Services, Inc.

470 Murdock Avenue
Meriden, CT 06450

Phone: (203) 238-4846

IH - 26 - 2462
New Hartford Public Schools
New Hartford Elementary School

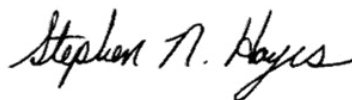
Collected: **July 10, 2026**
Received: **July 13, 2026**
Reported: **July 13, 2026**

We would like to thank you for trusting Hayes Microbial for your analytical needs!
We received 5 samples by FedEx in good condition for this project on July 13th, 2026.

The results in this analysis pertain only to this job, collected on the stated date, and should not be used in the interpretation of any other job. Information supplied by the customer can affect the validity of results. These results apply only to the samples as received. This report may not be duplicated, except in full, without the written consent of Hayes Microbial Consulting, LLC.

All information provided to Hayes Microbial is confidential information relating to our customers and their clients. We will not disclose, copy, or distribute any information verbally or written, except to those designated by the customer(s). We take confidentiality very seriously. No changes to the distribution list will be made without the express consent of the customer.

This laboratory bears no responsibility for sample collection activities, analytical method limitations, or your use of the test results. Interpretation and use of test results are your responsibility. Any reference to health effects or interpretation of mold levels is strictly the opinion of Hayes Microbial. In no event, shall Hayes Microbial or any of its employees be liable for lost profits or any special, incidental or consequential damages arising out of the use of these test results.



Steve Hayes, BSMT (ASCP)
Laboratory Director
Hayes Microbial Consulting, LLC.



EPA Laboratory ID: VA01419



Lab ID: #188863



DPH License: #PH-0198

Sample Number*	1	A1		2	A2		3	A3		4	A4	
Sample Name*	Outdoor Baseline			Library			Library			Library		
Sample Volume*	150 L			150 L			150 L			150 L		
Reporting Limit	7 spores/m³			7 spores/m³			7 spores/m³			7 spores/m³		
Background	1			2			2			1		
Fragments	ND			ND			ND			ND		
Organism	Raw Count	Count / m³	% of Total	Raw Count	Count / m³	% of Total	Raw Count	Count / m³	% of Total	Raw Count	Count / m³	% of Total
Alternaria	2	13	7.1%									
Ascospores	8	53	28.6%							1	7	100.0%
Aspergillus Penicillium												
Basidiospores	8	53	28.6%									
Bipolaris Drechslera												
Chaetomium												
Cladosporium	9	60	32.1%	1	7	100.0%	8	53	88.9%			
Curvularia												
Epicoccum	1	7	3.6%									
Fusarium												
Memnoniella												
Myxomycetes/Periconia							1	7	11.1%			
Pithomyces												
Rusts/Smuts												
Stachybotrys												
Stemphylium												
Torula												
Ulocladium												
Total	28	186	100%	1	7	100%	9	60	100%	1	7	100%

Water Damage Indicator	Common Allergen	Slightly Higher than Baseline	Significantly Higher than Baseline	Ratio Abnormality
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* indicates data provided by the customer



Collected: Jul 10, 2026

Received: Jul 13, 2026

Reported: Jul 13, 2026

Project Analyst:
 Katelyn Harris, *Katelyn Harris*

Date:
07 - 13 - 2026

Reviewed By:
 Steve Hayes, BSMT *Stephen N. Hayes*

Date:
07 - 13 - 2026

#5	Bio-Tape (1.00 cm2*)	Organism	Spore Estimate	Mycelial Estimate
T1 - Library Window Sill		No Fungi Detected		

* indicates data provided by the customer



Collected: Jul 10, 2026

Received: Jul 13, 2026

Reported: Jul 13, 2026

Project Analyst: Katelyn Harris, <i>Katelyn Harris</i>	Date: 07 - 13 - 2026	Reviewed By: Steve Hayes, BSMT <i>Stephen N. Hayes</i>	Date: 07 - 13 - 2026
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Spore Trap Information

Reporting Limit	The Reporting Limit is the lowest number of spores that can be detected based on the total volume of the sample collected and the percentage of the slide that is counted. At Hayes Microbial, 100% of the slide is read so the LOD is based solely on the total volume. Raw spore counts that exceed 500 spores will be estimated.
Blanks	Results have not been corrected for field or laboratory blanks.
Background	The Background is the amount of debris that is present in the sample. This debris consists of skin cells, dirt, dust, pollen, drywall dust and other organic and non-organic matter. As the background density increases, the likelihood of spores, especially small spores such as those of <i>Aspergillus</i> and <i>Penicillium</i> may be obscured. The background is rated on a scale of 1 to 5 and each level is determined as follows: NBD: No background detected due to possible pump or cassette malfunction. Recollect sample. (Field Blanks will display NBD) 1 : <5% of field occluded. No spores will be uncountable. 2 : 5-25% of field occluded. 3 : 25-75% of field occluded. 4 : 75-90% of field occluded. 5 : >90% of field occluded. Suggested recollection of sample.
Fragments	Fragments are small pieces of fungal mycelium or spores. They are not identifiable as to type and when present in very large numbers, may indicate the presence of mold amplification.
Control Comparisons	There are no national standards for the numbers of fungal spores that may be present in the indoor environment. As a general rule and guideline that is widely accepted in the indoor air quality field, the numbers and types of spores that are present in the indoor environment should not exceed those that are present outdoors at any given time. There will always be some mold spores present in "normal" indoor environments. The purpose of sampling and counting spores is to help determine whether an abnormal condition exists within the indoor environment and if it does, to help pinpoint the area of contamination. Spore counts should not be used as the sole determining factor of mold contamination. There are many factors that can cause anomalies in the comparison of indoor and outdoor samples due to the dynamic nature of both of those environments.
Water Damage Indicator	Blue: These molds are commonly seen in conditions of prolonged water intrusion and usually indicate a problem.
Common Allergen	Green: Although all molds are potential allergens, these are the most common allergens that may be found indoors.
Slightly Higher than Baseline	Orange: The spore count is slightly higher than the outside count and may or may not indicate a source of contamination.
Significantly Higher than Baseline	Red: The spore count is significantly higher than the baseline count and probably indicates a source of contamination.
Ratio Abnormality	Violet: The types of spores found indoors should be similar to the ones that were identified in the baseline sample. Significant increases (more than 25%) in the ratio of a particular spore type may indicate the presence of abnormal levels of mold, even if the total number of spores of that type is lower in the indoor environment than it was outdoors.
Color Coding	Fungi that are present in indoor samples at levels lower than 200 per cubic meter are not color coded on the report, unless they are one of the water damage indicators.
Significant Figures	Raw counts and column totals may reflect more than 2 significant figures, but results should only be considered significant to 2 figures.

Direct Analysis Information

Spore Estimate		Percentages
ND	None Detected	0%
Rare	Less than 10 spores	< 1%
Light	10 - 99 spores	1-10%
Moderate	100 - 999 spores	11-25%
Heavy	1000 - 9999 spores	26-50%
Very Heavy	10000 or greater spores	51-100%

Mycelial Estimate	
ND	None Detected No active growth at site.
Trace	Very small amount of Mycelium Probably no active growth at site.
Few	Some Mycelium Possible active growth at site.
Many	Large amount of Mycelium Probable active growth at site.

Analyte Descriptions

Alternaria	Habitat:	Commonly found outdoors in soil and decaying plants. Indoors, it is commonly found on window sills and other horizontal surfaces.
	Health Effects:	A common allergen and has been associated with hypersensitivity pneumonitis. Alternaria is capable of producing toxic metabolites which may be associated with disease in humans or animals. Occasionally an agent of onychomycosis, ulcerated cutaneous infection and chronic sinusitis, principally in the immunocompromised patient.
Ascospores	Habitat:	A large group consisting of more than 3000 species of fungi. Common plant pathogens and outdoor numbers become very high following rain. Most of the genera are indistinguishable by spore trap analysis and are combined on the report.
	Health Effects:	Health affects are poorly studied, but many are likely to be allergenic.
Basidiospores	Habitat:	A common group of Fungi that includes the mushrooms and bracket fungi. They are saprophytes and plant pathogens. In wet conditions they can cause structural damage to buildings.
	Health Effects:	Common allergens and are also associated with hypersensitivity pneumonitis.
Cladosporium	Habitat:	One of the most common genera worldwide. Found in soil and plant debris and on the leaf surfaces of living plants. The outdoor numbers are lower in the winter and often relatively high in the summer, especially in high humidity. The outdoor numbers often spike in the late afternoon and evening. Indoors, it can be found growing on textiles, wood, sheetrock, moist window sills and in HVAC supply ducts.
	Health Effects:	A common allergen, producing more than 10 allergenic antigens and a common cause of hypersensitivity pneumonitis.
Epicoccum	Habitat:	It is found in soil and plant litter and is a plant pathogen. It can grow indoors on a variety of substrates, including paper and textiles and is commonly found on wet drywall.
	Health Effects:	It is a common allergen. No cases of infection have been reported in humans.
Myxomycetes/Periconia	Description:	Found on decaying plant material and as a plant pathogen.
	Health Effects:	Some allergenic properties reported, but generally pose no health concerns to humans.

IV. ADDITIONAL NOTES

Airborne mold concentrations (levels) and/or surfaces may change over a period of time. The sample results are a representation of the indoor air quality and/or surfaces at the time the test was performed. Mold may grow in indoor environments without producing airborne mold spores or hyphae. EnviroMed cannot guarantee the identification of all/every mold growth area based on airborne testing. EnviroMed Services Inc. is not liable for inaccessible areas, illness, allergies, or damage to your property as a result of this report's findings. EnviroMed Services Inc. is not liable for the outcome of the remediation process/completion as a result of this report's findings.



AIHA Laboratory Accreditation Programs, LLC
acknowledges that
Hayes Microbial Consulting
3005 E. Boundary Terrace, Suite F Midlothian, VA 23112
Laboratory ID: LAP-188863

along with all premises from which key activities are performed, as listed above, has fulfilled the requirements of the AIHA Laboratory Accreditation Programs, LLC (AIHA LAP) accreditation to the ISO/IEC 17025:2017 international standard, General Requirements for the Competence of Testing and Calibration Laboratories in the following:

LABORATORY ACCREDITATION PROGRAMS

☐	INDUSTRIAL HYGIENE	Accreditation Expires:
☐	ENVIRONMENTAL LEAD	Accreditation Expires:
☒	ENVIRONMENTAL MICROBIOLOGY	Accreditation Expires: April 01, 2027
☐	FOOD	Accreditation Expires:
☐	UNIQUE SCOPES	Accreditation Expires:
☐	BE FIELD/MOBILE	Accreditation Expires:

Specific Field(s) of Testing/Method(s) within each Accreditation Program for which the above named laboratory maintains accreditation is outlined on the attached Scope of Accreditation. Continued accreditation is contingent upon successful on-going compliance with ISO/IEC 17025:2017 and AIHA LAP requirements. This certificate is not valid without the attached Scope of Accreditation. Please review the AIHA LAP website (www.aihaaccreditedlabs.org) for the most current Scope.

A handwritten signature in black ink that reads 'Cheryl O. Morton'.

Cheryl O Morton
Managing Director, AIHA Laboratory Accreditation Programs, LLC



AIHA Laboratory Accreditation Programs, LLC

SCOPE OF ACCREDITATION

Hayes Microbial Consulting

3005 E. Boundary Terrace, Suite F Midlothian, VA
23112

Laboratory ID: LAP-188863

Issue Date: 05/01/2023

The laboratory is approved for those specific field(s) of testing/methods listed in the table below. Clients are urged to verify the laboratory's current accreditation status for the particular field(s) of testing/Methods, since these can change due to proficiency status, suspension and/or withdrawal of accreditation.

Environmental Microbiology Laboratory Accreditation Program (EMLAP)

Initial Accreditation Date: 08/01/2010

EMLAP Scope Category	Field of Testing (FOT)	Component, parameter or characteristic tested	Method	Method Description (for internal methods only)
Fungal	Air - Culturable	Air	HMC-#103	In House: Viable Air Culture
Fungal	Air - Direct Examination	Spore Trap	HMC-#101	In House: Spore Trap Analysis
Fungal	Bulk - Culturable	Bulk Material	HMC-#104	In House: Bulk Culture Analysis
Fungal	Bulk - Direct Examination	Bulk Material	HMC-#102	In House: Direct Identification Analysis
Fungal	Surface - Culturable	Swab	HMC-#105	In House: Swab Culture Analysis
Fungal	Surface - Direct Examination	Bio-Tape, Swab	HMC-#102	In House: Direct Identification Analysis

A complete listing of currently accredited EMLAP laboratories is available on the AIHA LAP, LLC website at:
<http://www.aihaaccreditedlabs.org>