



Cleaner environment. Safer workplaces.

**Mold and Indoor Air Quality Report
for**

Ann Antolini Elementary School
30 Antolini Road
New Hartford, CT 06057

-

Prepared For

New Hartford Public Schools
530 Main Street, P.O. Box 315
New Hartford, CT 06057

Date of Inspection and Testing:

March 11, 2026

EnviroMed Project

IH-26-226

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TABLE OF CONTENTS

Section	Page
I. PROJECT NARRATIVE.....	1
Overview	1
Methods	1
Summary of Results:.....	2
Conclusions and Recommendations	6
II. PHOTOGRAPHS	8
III . TABLE OF IAQ MEASUREMENTS	13
IV. LABORATORY ANALYSIS REPORT	14

I. PROJECT NARRATIVE

Overview

On March 11, 2026, EnviroMed Services conducted a microbial and indoor air quality assessment at Ann Antolini Elementary School on 30 Antolini Road, New Hartford, CT. The purpose of this assessment, as directed by the client, was to measure indoor air quality, and provide a baseline survey of the entire school for airborne fungal contamination.

Methods

The following sampling and assessment scheme was employed:

- Direct reading measurements were collected using a TSI Q-Trak™ model 8550/8551 IAQ Monitor. The baseline indoor air quality (IAQ) parameters temperature (°F), relative humidity (%), carbon monoxide (ppm), and carbon dioxide (ppm).
- Direct reading measurements for Volatile Organic Compounds (VOCs) were taken using a ppb RAE 3000 Photo Ionization Detector (PID).
- Direct reading measurements were taken for a total particulate in the air using an MIE PDR 1000 Personal Data Ram Particulate Analyzer and a Temtop LKC-1000 Series Air Quality Monitor
- Direct reading measurements for Formaldehyde were taken using a Temtop LKC-1000 Series Air Quality Monitor
- Direct reading measurements were collected using MultiRAE PGM-6228 Wireless Multi-Gas Monitor. Oxygen (O₂), Lower Explosive Limit (LEL)
- Conduct air sampling to test for the presence of airborne fungal structures, including spores, by drawing air through a Zefon Air-O-Cell cassette using a pre-calibrated vacuum pump. In this method, air enters the cassette, and particles are collected on the sampling substrate. The manufacturer-recommended flow rate of 15 liters per minute of air, for a duration of 10 minutes, was used to collect each sample. Sample locations were selected based on client direction. The samples were sent to Hayes Microbial Consulting in Midlothian, VA, for analysis by optical microscopy. Hayes Microbial Consulting is approved by the American Industrial Hygiene Association (AIHA Cert# 188863) and the state of Connecticut Department of Public Health. The results are represented in counts of fungal structures and fungal structures per cubic meter of air (spores/m³). The results are compared to the baseline sample that was collected outdoors.

Summary of Results:

Temperature: The indoor temperatures ranged from 69.7 to 75.6 °F. The ASHRAE recommended temperatures range from 68.0 to 74.0 °F. The indoor temperature readings were generally within above the recommended range.

Relative Humidity: The indoor relative humidity ranged from 30.3 to 45.7 percent. ASHRAE recommended range for indoor relative humidity is 20 to 60 percent. The relative humidity readings were within the recommended range.

Carbon Monoxide: The indoor carbon monoxide concentration did not exceed 0.0 ppm. The OSHA PEL limit for carbon monoxide is 50 ppm, reported as an 8-hour time-weighted average. The carbon monoxide readings were below the limit.

Particulates in Air (Dust): The indoor concentration of particulates in air inside the building did not exceed 15.9 ug/m³ for PM 2.5, and 26.5 ug/m³ for PM 10. The OSHA Permissible Exposure Limit (PEL) for respirable and total particulates in 5,000 ug/m³ and 15,000 ug/m³ respectively.

Volatile Organic Compounds (VOCs): The maximum concentration of Total VOCs per the Leadership in Energy and Environmental Design (LEED) Standards is 0.500 ppm. VOC readings ranged from 0.077 ppm to 0.607 ppm, which is generally within the standard. The minor levels above 0.500 ppm may be due to the presence of cleaning fluids, solvents, perfumes, or fragrance sprays.

Formaldehyde: Formaldehyde (CH₂O) levels did not exceed 0.16 ppm throughout the testing. This concentration is below the OSHA formaldehyde limit of 0.75 ppm, reported as an 8-hour time weighted average.

Carbon Dioxide: The outside carbon dioxide concentration recorded in the morning on this day was 443 ppm. The indoor concentrations during the morning ranged from 414 to 1250 ppm. The outside carbon dioxide concentration recorded in the afternoon on this day was 387 ppm. The indoor concentrations ranged from 445 to 1993 ppm.

Per the ASHRAE Standard 62-2001, Ventilation for Acceptable Indoor Air Quality, the difference between the indoor and outdoor concentrations of carbon dioxide should be less than 707 ppm. The difference between the highest indoor concentration and the outdoor concentration in the morning was 807 ppm. The difference between the highest indoor concentration and the outdoor concentration in the afternoon was 1606 ppm. The averages of the morning and afternoon carbon dioxide levels were above ASHRAE guidelines. Additionally, carbon dioxide concentrations in the morning and afternoon were well below the OSHA permissible exposure limit of 5000 ppm.

Oxygen (O₂): The indoor oxygen concentrations ranged from 21.4 to 21.7 percent. OSHA standard for oxygen concentration is 19.6 to 23.4 percent. The indoor concentration readings were all within the recommended range.

Lower Explosive Limit (LEL): The indoor lower explosive limit ranged from 0 to 0 percent. OSHA states that atmospheres with flammable vapors at or above 10 percent are considered hazardous. The lower explosive limit readings were below the hazardous limits.

Section III of this report contains the IAQ data results.

Air-O-Cell Spore Trap Sampling Results:

There are currently no regulatory standards for concentrations of mold in air under OSHA. Levels of mold within the building are routinely compared to the baseline levels outside the building to detect any amplifications or unusual toxigenic mold species.

The air sampling results indicated fungal amplification of toxigenic mold in the Nurse's Room.

Section IV of this report contains a copy of the analytical report.

Alternaria: Commonly found outdoors in soil and decaying plants. Indoors, it is commonly found on windowsills and other horizontal surfaces. A common allergen and has been associated with hypersensitivity pneumonitis. Alternaria is capable of producing toxic metabolites which may be associated with disease in humans or animals. Occasionally an agent of onychomycosis ulcerated cutaneous infection and chronic sinusitis, principally in the immunocompromised patient.

Ascospores: A large group consisting of more than 3000 species of fungi. Common plant pathogens and outdoor numbers become very high following rain. Most of the genera are indistinguishable by spore trap analysis and are combined on the report.

Aspergillus | Penicillium: The most common fungi isolated from the environment. Very common in soil and on decaying plant material. Can grow well indoors on a wide variety of substrates. This group contains common allergens, and many can cause hypersensitivity pneumonitis. They may cause extrinsic asthma, and many are opportunistic pathogens. Many species produce mycotoxins which may be associated with disease in humans and other animals. Toxin production is dependent on the species, the food source, competition with other organisms, and other environmental conditions.

Basidiospores: A common group of Fungi that includes mushrooms and bracket fungi. They are saprophytes and plant pathogens. In wet conditions, they can cause structural damage to buildings. These are common allergens and are also associated with hypersensitivity pneumonitis.

Chaetomium: Ascomycete fungus, commonly isolated from soil and decaying plant materials. It is cellulolytic and grows well indoors on damp sheetrock and other paper substrates. It is often found growing with *Stachybotrys*. It is reported to be allergenic and may produce toxins.

Cladosporium: One of the most common genera worldwide. Found in soil and plant debris and on the leaf surfaces of living plants. The outdoor numbers are lower in the winter and often relatively high in the summer, especially in high humidity. The outdoor numbers often spike in the late afternoon and evening. Indoors, it can be found growing on textiles, wood, sheetrock, moist windowsills and in HVAC supply ducts. A common allergen, producing more than 10 allergenic antigens and a common cause of hypersensitivity pneumonitis.

Culvularia: They exist in soil and plant debris, and are plant pathogens. They are allergenic and a common cause of allergic fungal sinusitis. An occasional cause of human infection, including keratitis, sinusitis, onychomycosis, mycetoma, pneumonia, endocarditis and disseminated infection, primarily in the immunocompromised.

Epicoccum: It is found in soil and plant litter and is a plant pathogen. It can grow indoors on a variety of substrates, including paper and textiles and is commonly found on wet drywall. It is a common allergen. No cases of infection have been reported in humans.

Myxomycetes: Found on decaying plant material and as a plant pathogen. Some allergenic properties reported, but generally pose no health concerns to humans.

Pithomyces: Common fungus isolated from soil, decaying plant material. Rarely found indoors. Allergenic properties are poorly studied. No cases of infection in humans.

Sample ID: A1**Sample Location: Outside Baseline**

The prominent fungal genus present was *Basidiospores*. The other fungal genera present were *Ascospores*, *Cladosporium*, and *Epicoccum*.

Sample ID: A2**Sample Location: Room 5**

The prominent fungal genus present was *Cladosporium*. The other fungal genera present were *Alternaria*, *Ascospores*, *Basidiospores*, *Culvularia*, and *Myxomycetes*. There was no amplification detected for any of these fungal genera as compared to the outdoor baseline sample.

Sample ID: A3**Sample Location: Room 6**

The prominent fungal genus present was *Ascospores*. The other fungal genus present was *Cladosporium*. There was no amplification detected for any of these fungal genera as compared to the outdoor baseline sample.

Sample ID: A4**Sample Location: Room 28**

The prominent fungal genus present was *Alternaria*. The other fungal genera present were *Ascospores*. There was no amplification detected for any of these fungal genera as compared to the outdoor baseline sample.

Sample ID: A5**Sample Location: Main Library**

The only fungal genus present was *Cladosporium*. There was no amplification detected for this fungal genus as compared to the outdoor baseline sample.

Sample ID: A6**Sample Location: Room 26**

The prominent fungal genus present was *Ascospores*. The other fungal genera present were *Basidiospores*, *Cladosporium*, *Myxomycetes*, and *Pithomyces*. There was no amplification detected for any of these fungal genera as compared to the outdoor baseline sample.

Sample ID: A7**Sample Location: Room 15**

The prominent fungal genus present was *Cladosporium*. The other fungal genera present were *Ascospores* and *Basidiospores*. There was no amplification detected for any of these fungal genera as compared to the outdoor baseline sample.

Sample ID: A8**Sample Location: Nurse's Room**

The prominent fungal genus present was *Chaetomium*. *Chaetomium* was detected with a spore count slightly higher than the outdoor baseline sample, indicating an amplification of this fungi. *Chaetomium* is considered a toxigenic mold.

The other fungal genera present were *Ascospores*, *Aspergillus/Penicillium*, *Basidiospores*, *Cladosporium*, and *Myxomycetes*. There was no amplification detected for any of these fungal genera as compared to the outdoor baseline sample.

Sample ID: A9**Sample Location: Main Office**

The prominent fungal genus present was *Aspergillus/Penicillium*. The other fungal genus present was *Ascospores*, *Cladosporium*, and *Myxomycetes*. There was no amplification detected for any of these fungal genera as compared to the outdoor baseline sample.

The prominent fungal genus present was *Cladosporium*. The other fungal genera present were *Ascospores* and *Basidiospores*. There was no amplification detected for any of these fungal genera as compared to the outdoor baseline sample.

Conclusions and Recommendations

Based on the results obtained and observations made during the assessment, EnviroMed Services, Inc. has come to the following conclusions and recommendations.

Conclusions:

Indoor Air Quality:

Based on the results obtained and observations made during the assessment, EnviroMed Services, Inc. has come to the following conclusions and recommendations.

- The indoor air temperatures throughout the school were generally within the recommended ASHRAE standard range at the time of the assessment.
- All the indoor relative humidity readings throughout the school were within the recommended ASHRAE standard range at the time of the assessment.
- The indoor carbon monoxide concentrations throughout the school were all well below the OSHA recommended limit at the time of the assessment.
- All the indoor carbon dioxide concentrations throughout the school were generally within the ASHRAE recommended limit with a few exceedances, and below the OSHA permissible exposure limit at the time of the assessment.
- The concentration of particulates in air inside the building were well below the OSHA PEL at the time of the assessment.
- The concentration of formaldehyde inside the building was well below the OSHA PEL at the time of the assessment.
- The oxygen concentrations inside the building were within the OSHA standard at the time of the assessment
- The indoor lower explosive limit was below the hazardous limits at the time of the assessment.

Mold:
Air Samples:

- The air sample results indicated fungal amplification of toxigenic mold in the Nurse's Room.

Recommendations:

EnviroMed Services, Inc. recommends that a professional remediation contractor clean all cleanable nonporous surfaces with a disinfecting agent such as Fiberlock ShockWave or equivalent for all areas of concern within the following locations: Nurse's Room; and to remove and replace any porous water or moisture impacted materials that cannot be appropriately cleaned or dried. In addition, EnviroMed Services, Inc. recommends the use of a HEPA filtered air filtration system to remove all airborne spores as amplification that were detected throughout all affected areas. The Professional Mold Remediation contractor hired will provide the specific means and methods to remediate for the best course of action as approved by the Client. Some General Recommendations for reference are as follows:

We recommend the use of a HEPA (High-Efficiency Particulate Air) vacuum. Cleaning will also need to be performed on any non-movable items located in the unit, movable items will need to be removed prior to the start of remediation. Carpeting should also be HEPA vacuum cleaned with a disinfecting agent or removed and replaced.

Prior to cleaning, Negative Air Pressure Containment should be set up to prevent the contamination of any airborne mold spores during the remediation process. Containment is to be exhausted outside the containment area during remediation. The HEPA Sandwich method, as referenced in mold remediation under the IICRC/ANSI S520 Standard, involves a three-step cleaning process designed to remove mold contamination from surfaces effectively. This method consists of Initial HEPA vacuuming, Damp wiping & Final HEPA vacuuming or additional damp wiping. The HEPA Sandwich method is recommended with the continued use of an industrial HEPA Air Unit for at least 24-48 hours post-abatement work to further clean the air.

Following the remediation, EnviroMed recommends a follow-up clearance microbial assessment be performed by a professional industrial hygienist to verify the effectiveness of the cleaning following remediation.

II. PHOTOGRAPHS



Figure 1: Sample A-1 – Outdoor Baseline

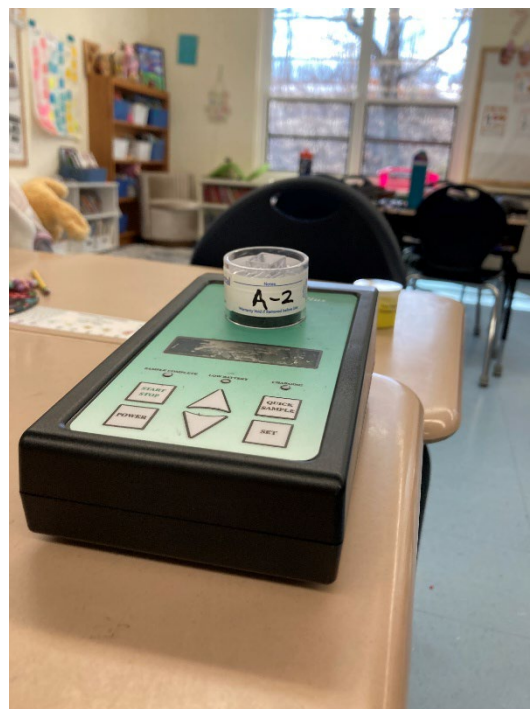


Figure 2: Sample A-2 – Room 5

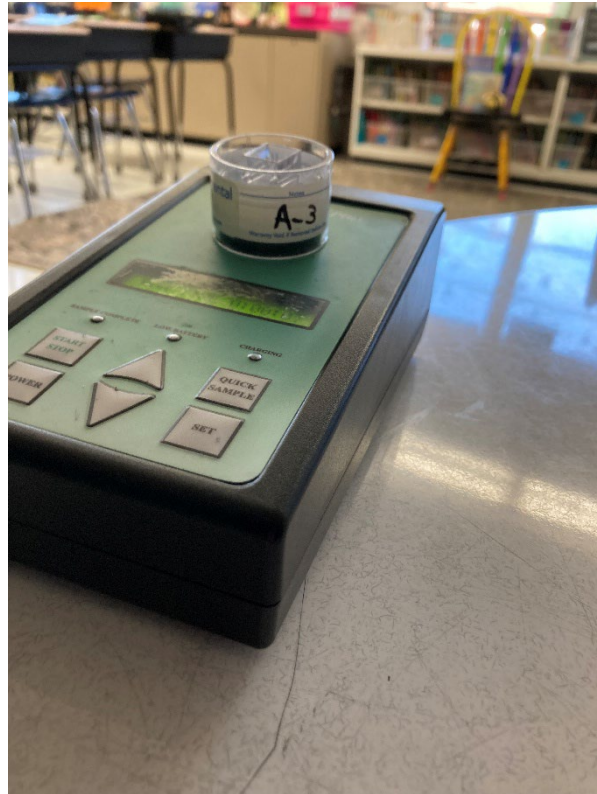


Figure 3: Sample A-3 – Room 6



Figure 4: Sample A-4 – Room 28

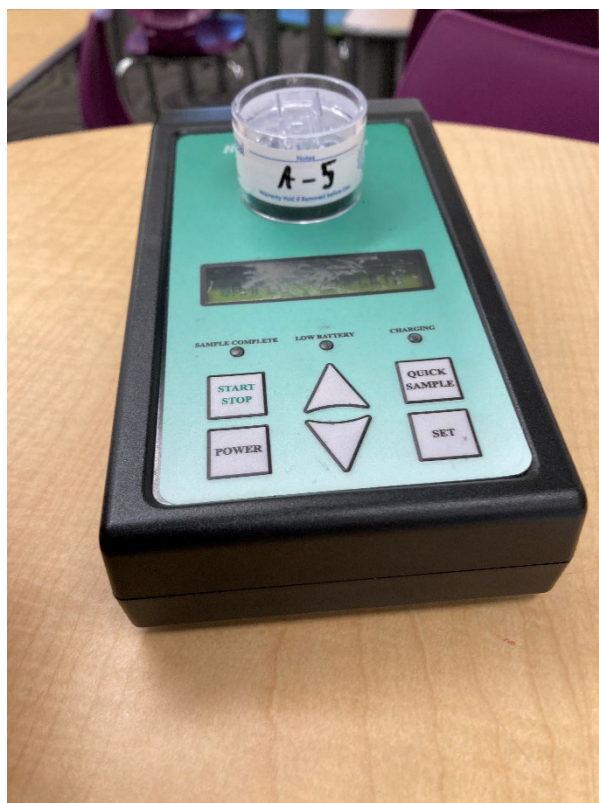


Figure 5: Sample A-5 – Main Library

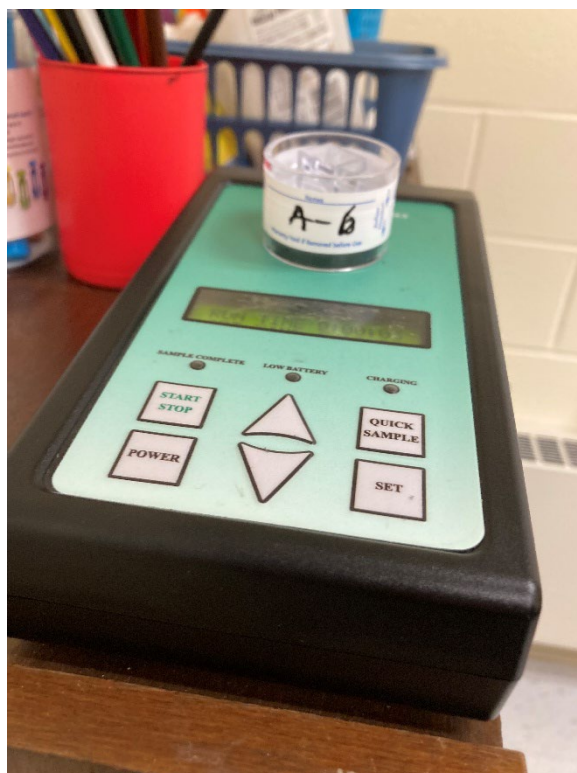


Figure 6: Sample A-6 – Room 26



Figure 7: Sample A-7 – Room 15

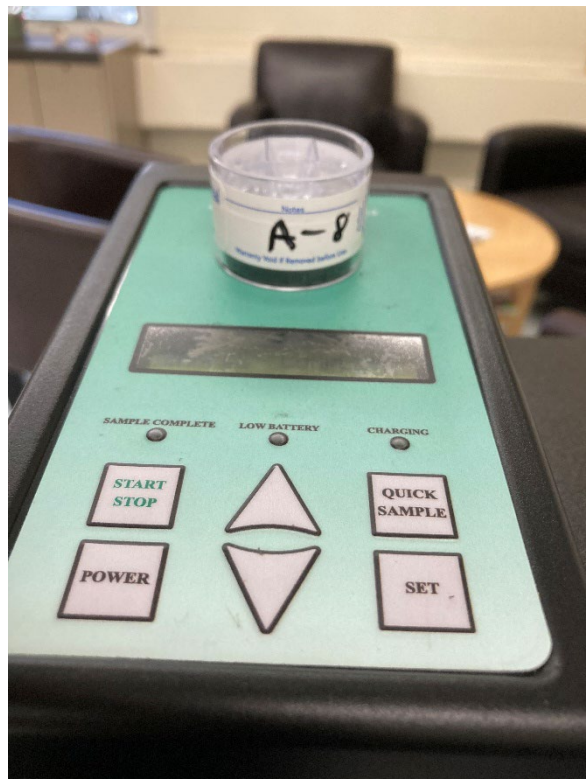


Figure 8: Sample A-8 – Nurse's Room

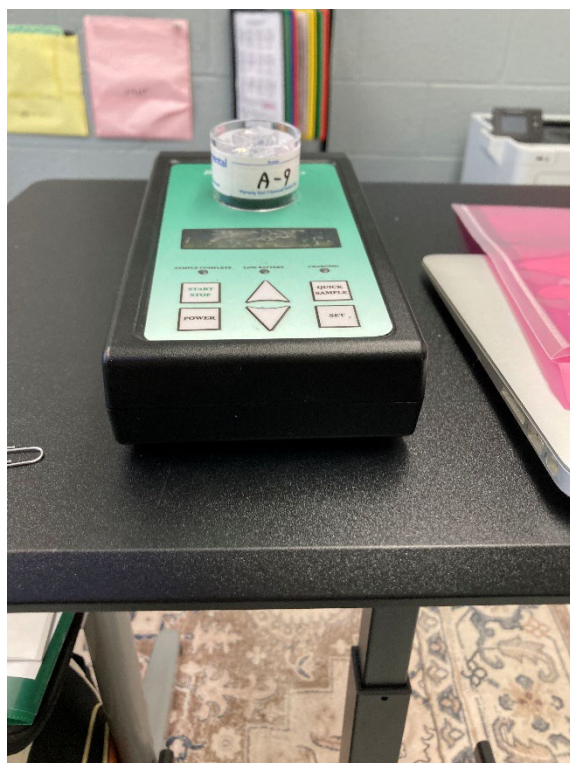


Figure 9: Sample A-9 – Main Office

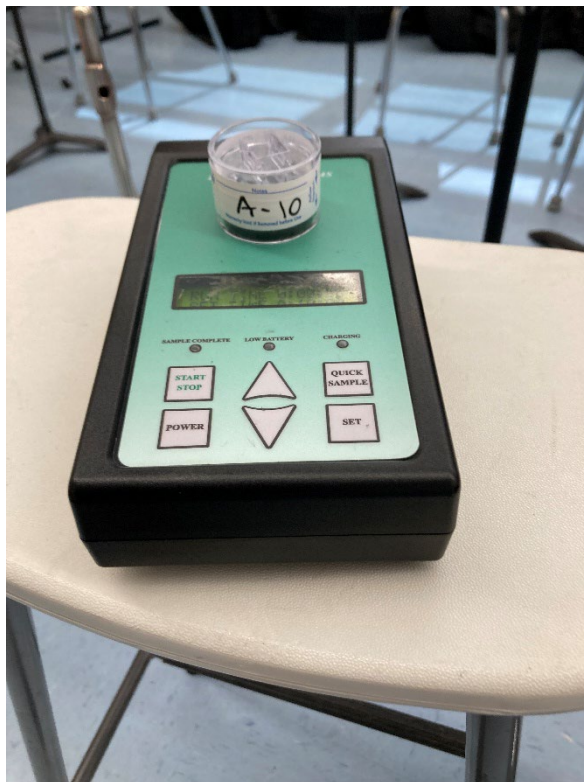


Figure 10: Sample A-10 – Band Room

III . TABLE OF IAQ MEASUREMENTS

A.M. Readings

Location	Type	Temp °F	RH %	CO (ppm)	CO ₂ (ppm)	VOCs (ppm)	PM _{2.5} (ug/m ³)	PM ₁₀ (ug/m ³)	CH ₂ O (mg/m ³)	OXY	LEL %	Occupied
Outdoor Baseline	Outside	50.9	53.3	0.0	443	0.000	10.2	17.4	0.01	21.2	0	-
Room 14	Classroom	69.7	37.2	0.0	752	0.077	3.5	6.5	0.03	21.2	0	Yes
Room 13	Classroom	71.2	34.1	0.0	869	0.115	3.8	6.5	0.02	21.2	0	Yes
Room 12	Classroom	72.1	33.9	0.0	753	0.144	4.3	7.8	0.04	21.2	0	Yes
Room 11	Classroom	72.2	31.9	0.0	641	0.160	5.2	9.0	0.04	21.2	0	Yes
Room 10	Classroom	72.7	31.5	0.0	652	0.191	3.7	5.6	0.04	21.2	0	Yes
Room 9	Classroom	73.0	30.8	0.0	598	0.316	5.2	9.4	0.16	21.2	0	Yes
Room 8	Classroom	72.8	32.7	0.0	850	0.247	4.7	8.0	0.07	21.2	0	Yes
Room 7	Classroom	73.3	34.2	0.0	1013	0.357	4.5	8.1	0.08	21.1	0	Yes
Room 6	Classroom	72.9	34.3	0.0	1113	0.340	4.0	7.4	0.06	21.2	0	Yes
Room 5	Classroom	72.7	35.7	0.0	1250	0.311	4.4	8.3	0.03	21.2	0	Yes
Room 4	Classroom	72.0	33.8	0.0	950	0.292	6.7	11.5	0.02	21.2	0	Yes
Room 3	Classroom	72.2	33.8	0.0	1050	0.321	5.0	8.7	0.03	21.2	0	Yes
East Wing Hallway	Hallway	72.3	32.3	0.0	792	0.319	5.0	8.5	0.05	21.3	0	No
Room 29	Classroom	71.3	32.3	0.0	575	0.310	4.2	7.3	0.02	21.5	0	No
IT Room 30	Office	71.4	33.4	0.0	661	0.270	4.3	7.7	0.01	21.4	0	Yes
Room 28	Classroom	70.4	33.7	0.0	701	0.607	4.6	8.1	0.10	21.4	0	Yes

Location	Type	Temp °F	RH %	CO (ppm)	CO ₂ (ppm)	VOCs (ppm)	PM _{2.5} (ug/m ³)	PM ₁₀ (ug/m ³)	CH ₂ O (mg/m ³)	OXY	LEL %	Occupied
Room 27	Classroom	70.2	32.3	0.0	565	0.308	4.1	7.2	0.02	21.5	0	No
Room 26	Classroom	71.4	32.6	0.0	620	0.379	3.9	5.8	0.08	21.4	0	Yes
Room 34	Office	71.2	32.1	0.0	603	0.300	3.2	4.8	0.02	21.4	0	Yes
Main Library	Library	71.0	33.3	0.0	715	0.298	4.7	8.0	0.01	21.4	0	Yes
Room 36	Office	70.9	32.6	0.0	645	0.295	3.6	6.2	0.01	21.4	0	No
Room 24	Office	70.3	33.6	0.0	648	0.325	3.7	6.0	0.02	21.4	0	No
Room 27	Classroom	69.6	32.6	0.0	566	0.296	4.5	7.7	0.02	21.5	0	No
Room 25	Classroom	70.5	32.5	0.0	568	0.313	3.9	6.6	0.02	21.5	0	No
West Hallway by Library	Hallway	70.2	33.9	0.0	634	0.322	4.6	7.8	0.02	21.4	0	No
Room 22	Classroom	70.6	33.1	0.0	624	0.329	4.7	7.6	0.02	21.5	0	Yes
Room 23	Classroom	71.0	32.5	0.0	542	0.319	4.3	7.7	0.02	21.5	0	Yes
Room 20	Classroom	72.8	38.0	0.0	1043	0.365	4.5	7.1	0.03	21.4	0	Yes
Room 21	Classroom	75.6	34.6	0.0	987	0.356	4.9	8.4	0.03	21.4	0	Yes
Room 19	Classroom	72.9	32.7	0.0	673	0.305	6.9	12.0	0.01	21.5	0	Yes
Room 18	Classroom	72.7	31.6	0.0	674	0.384	6.0	9.5	0.03	21.5	0	No
Room 17	Classroom	74.4	31.2	0.0	555	0.322	6.2	10.5	0.04	21.5	0	Yes
Room 16	Classroom	75.5	30.8	0.0	549	0.298	7.6	13.0	0.02	21.6	0	No
Room 15	Classroom	74.7	33.7	0.0	855	0.405	6.1	11.0	0.03	21.5	0	Yes
Nurse's Office	Office	74.2	31.4	0.0	642	0.352	5.9	10.5	0.04	21.5	0	Yes

Location	Type	Temp °F	RH %	CO (ppm)	CO ₂ (ppm)	VOCs (ppm)	PM _{2.5} (ug/m ³)	PM ₁₀ (ug/m ³)	CH ₂ O (mg/m ³)	OXY	LEL %	Occupied
Music Room (40)	Classroom	75.8	30.9	0.0	705	0.347	6.1	10.3	0.03	21.5	0	No
Staff Room (42)	Office	74.8	30.3	0.0	635	0.346	6.5	10.6	0.03	21.5	0	No
Main Office	Office	73.0	32.6	0.0	887	0.338	6.0	11.1	0.03	21.5	0	Yes
Principal's Office	Office	71.7	33.4	0.0	757	0.332	5.7	10.0	0.03	21.5	0	Yes
Art Room (41)	Classroom	73.4	31.6	0.0	492	0.288	7.4	12.8	0.01	21.6	0	Yes
Room 44	Office	73.0	35.1	0.0	864	0.332	6.7	11.5	0.01	21.5	0	Yes
Room 46	Classroom	71.6	32.4	0.0	717	0.408	4.9	9.6	0.02	21.5	0	No
Gymateria	Common	71.6	35.4	0.0	852	0.409	5.7	10.0	0.03	21.5	0	Yes
Kitchen	Kitchen	71.7	34.8	0.0	772	0.372	6.3	10.7	0.04	21.6	0	Yes
Kitchen Office	Office	72.5	35.6	0.0	831	0.343	6.0	10.1	0.04	21.6	0	No
Multipurpose Room	Auditorium	72.2	31.1	0.0	414	0.255	4.4	7.8	0.01	21.6	0	No
Room 43 Band Room	Classroom	72.9	32.1	0.0	439	0.265	6.4	11.1	0.02	21.6	0	No
North Hallway	Hallway	72.5	33.0	0.0	730	0.310	5.8	10.1	0.01	21.6	0	No

P.M. Readings

Location	Type	Temp °F	RH %	CO (ppm)	CO ₂ (ppm)	VOCs (ppm)	PM _{2.5} (ug/m ³)	PM ₁₀ (ug/m ³)	CH ₂ O (mg/m ³)	OXY	LEL %	Occupied
Outdoor Baseline	Outside	69.8	38.7	0.0	387	0.000	9.2	15.3	0.01	21.6	0	-
Room 14	Classroom	73.3	38.8	0.0	1270	0.080	3.9	7.5	0.05	21.4	0	Yes
Room 13	Classroom	74.9	38.0	0.0	1490	0.200	4.9	8.2	0.09	21.4	0	Yes
Room 12	Classroom	74.9	37.0	0.0	1017	0.091	5.5	9.5	0.12	21.5	0	Yes
Room 11	Classroom	74.3	33.0	0.0	1033	0.150	4.0	6.7	0.06	21.5	0	No
Room 10	Classroom	74.2	34.1	0.0	1033	0.148	3.4	5.4	0.06	21.4	0	Yes
Room 9	Classroom	74.7	34.0	0.0	1092	0.172	4.5	7.6	0.06	21.5	0	Yes
Room 8	Classroom	73.9	38.5	0.0	1536	0.160	4.3	6.9	0.07	21.4	0	Yes
Room 7	Classroom	74.8	39.4	0.0	1550	0.302	3.7	6.0	0.09	21.4	0	Yes
Room 6	Classroom	74.0	38.4	0.0	963	0.154	5.5	9.5	0.10	21.5	0	Yes
Room 5	Classroom	74.5	40.8	0.0	1993	0.233	3.9	7.6	0.08	21.3	0	Yes
Room 4	Classroom	74.3	39.7	0.0	1705	0.171	5.3	8.6	0.07	21.4	0	Yes
Room 3	Classroom	74.2	37.9	0.0	1023	0.179	7.2	12.9	0.04	21.5	0	Yes
East Wing Hallway	Hallway	74.1	34.0	0.0	1130	0.175	3.8	6.9	0.06	21.5	0	No
Room 29	Classroom	73.0	33.3	0.0	674	0.165	5.3	9.4	0.01	21.6	0	No
IT Room 30	Office	72.4	34.4	0.0	683	0.165	3.6	6.6	0.01	21.6	0	Yes
Room 28	Classroom	71.6	38.3	0.0	893	0.404	6.4	11.0	0.10	21.5	0	Yes

Location	Type	Temp °F	RH %	CO (ppm)	CO ₂ (ppm)	VOCs (ppm)	PM _{2.5} (ug/m ³)	PM ₁₀ (ug/m ³)	CH ₂ O (mg/m ³)	OXY	LEL %	Occupied
Room 27	Classroom	71.2	35.2	0.0	626	0.212	5.3	9.7	0.07	21.6	0	No
Room 26 OTPT	Classroom	71.0	35.4	0.0	674	0.208	5.4	9.6	0.03	21.6	0	No
Room 34	Office	71.2	38.3	0.0	681	0.311	4.1	8.1	0.06	21.6	0	Yes
Main Library	Library	71.9	39.1	0.0	734	0.331	4.5	8.3	0.11	21.6	0	Yes
Room 36	Inaccessible											
Room 24 study	Classroom	71.0	36.7	0.0	706	0.199	3.8	6.6	0.03	21.6	0	No
Room 27	Classroom	71.4	35.2	0.0	667	0.199	3.7	6.3	0.09	21.6	0	No
Room 25	Classroom	71.9	35.2	0.0	628	0.199	3.7	6.8	0.05	21.6	0	No
West Hallway by Library	Hallway	71.9	40.1	0.0	915	0.231	4.7	7.9	0.04	21.6	0	No
Room 22	Classroom	71.2	35.8	0.0	678	0.214	6.0	10.2	0.05	21.6	0	No
Room 23	Classroom	71.8	36.2	0.0	653	0.203	5.1	8.1	0.03	21.6	0	No
Room 20	Classroom	74.0	43.4	0.0	1094	0.199	4.9	8.4	0.04	21.5	0	Yes
Room 21	Classroom	74.8	39.8	0.0	930	0.203	7.2	12.4	0.03	21.5	0	Yes
Room 19	Classroom	74.7	43.0	0.0	913	0.155	9.2	15.9	0.04	21.6	0	Yes
Room 18	Classroom	73.6	35.2	0.0	774	0.203	7.1	11.7	0.02	21.6	0	No
Room 17	Classroom	74.0	39.1	0.0	608	0.188	7.5	11.8	0.02	21.6	0	Yes
Room 16	Classroom	74.2	34.5	0.0	525	0.198	7.6	12.6	0.02	21.6	0	No

Location	Type	Temp °F	RH %	CO (ppm)	CO ₂ (ppm)	VOCs (ppm)	PM _{2.5} (ug/m ³)	PM ₁₀ (ug/m ³)	CH ₂ O (mg/m ³)	OXY	LEL %	Occupied
Room 15	Classroom	74.5	39.0	0.0	867	0.209	8.3	14.3	0.03	21.6	0	Yes
Nurse's Office	Nurse	72.0	37.2	0.0	672	0.202	8.5	13.7	0.03	21.6	0	No
Music Room (40)	Classroom	71.8	39.7	0.0	445	0.172	9.2	15.1	0.02	21.6	0	No
Staff Room (42)	Lounge	73.0	40.1	0.0	617	0.215	9.2	18.7	0.02	21.6	0	No
Main Office	Office	72.9	38.2	0.0	734	0.226	8.2	12.8	0.02	21.6	0	Yes
Principal's Office	Office	72.5	39.2	0.0	949	0.256	7.5	13.6	0.02	21.6	0	No
Art Room (41)	Classroom	72.3	45.4	0.0	568	0.152	15.9	26.5	0.03	21.6	0	Yes
Room 44	Office	72.6	40.2	0.0	623	0.192	11.8	20.4	0.01	21.6	0	Yes
Room 46	Classroom	72.0	40.1	0.0	607	0.270	7.8	14.6	0.02	21.6	0	No
Gymateria	Common	72.9	45.7	0.0	684	0.205	10.7	17.8	0.03	21.6	0	Yes
Kitchen	Kitchen	73.2	45.2	0.0	703	0.185	10.7	18.2	0.03	21.6	0	No
Kitchen Office	Office	73.7	44.6	0.0	567	0.190	10.5	18.2	0.03	21.6	0	No
Multipurpose Room	Auditorium	72.6	42.0	0.0	410	0.128	11.1	18.5	0.01	21.7	0	No
Room 43 Band Room	Classroom	73.0	42.0	0.0	580	0.319	9.0	15.9	0.02	21.6	0	Yes
North Hallway	Hallway	72.4	43.3	0.0	609	0.156	10.5	18.8	0.01	21.6	0	Yes

IV. LABORATORY ANALYSIS REPORT



#26011687

Analysis Report prepared for

EnviroMed Services, Inc.

470 Murdock Avenue
Meriden, CT 06450

Phone: (203) 238-4846

IH-26-226

Ann Antolini Elementary School
30 Antolini Rd.
New Hartford, CT 06057

Collected: **March 11, 2026**
Received: **March 12, 2026**
Reported: **March 12, 2026**

We would like to thank you for trusting Hayes Microbial for your analytical needs!
We received 10 samples by FedEx in good condition for this project on March 12th, 2026.

The results in this analysis pertain only to this job, collected on the stated date, and should not be used in the interpretation of any other job. Information supplied by the customer can affect the validity of results. These results apply only to the samples as received. This report may not be duplicated, except in full, without the written consent of Hayes Microbial Consulting, LLC.

All information provided to Hayes Microbial is confidential information relating to our customers and their clients. We will not disclose, copy, or distribute any information verbally or written, except to those designated by the customer(s). We take confidentiality very seriously. No changes to the distribution list will be made without the express consent of the customer.

This laboratory bears no responsibility for sample collection activities, analytical method limitations, or your use of the test results. Interpretation and use of test results are your responsibility. Any reference to health effects or interpretation of mold levels is strictly the opinion of Hayes Microbial. In no event, shall Hayes Microbial or any of its employees be liable for lost profits or any special, incidental or consequential damages arising out of the use of these test results.

A handwritten signature in black ink that reads 'Stephen N. Hayes'.

Steve Hayes, BSMT (ASCP)
Laboratory Director
Hayes Microbial Consulting, LLC.



EPA Laboratory ID: VA01419



Lab ID: #188863



DPH License: #PH-0198

Sample Number*	1	A - 1		2	A - 2		3	A - 3		4	A - 4	
Sample Name*	Outdoor Baseline			Room 5			Room 6			Room 28		
Sample Volume*	150 L			150 L			150 L			150 L		
Reporting Limit	7 spores/m³			7 spores/m³			7 spores/m³			7 spores/m³		
Background	2			2			3			3		
Fragments	13/m³			7/m³			ND			ND		
Organism	Raw Count	Count / m³	% of Total	Raw Count	Count / m³	% of Total	Raw Count	Count / m³	% of Total	Raw Count	Count / m³	% of Total
Alternaria				1	7	3.1%						
Ascospores	132	880	32.0%	6	40	18.8%	3	20	75.0%	2	13	25.0%
Aspergillus Penicillium										2	13	25.0%
Basidiospores	202	1300	48.9%	5	33	15.6%				1	7	12.5%
Bipolaris Drechslera												
Chaetomium												
Cladosporium	77	510	18.6%	16	110	50.0%	1	7	25.0%	3	20	37.5%
Curvularia				3	20	9.4%						
Epicoccum	2	13	<1%									
Fusarium												
Memnoniella												
Myxomycetes/Periconia				1	7	3.1%						
Pithomyces												
Rusts/Smuts												
Stachybotrys												
Stemphylium												
Torula												
Ulocladium												
Total	413	2703	100%	32	217	100%	4	27	100%	8	53	100%

Water Damage Indicator	Common Allergen	Slightly Higher than Baseline	Significantly Higher than Baseline	Ratio Abnormality
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* indicates data provided by the customer



Collected: **Mar 11, 2026**

Received: **Mar 12, 2026**

Reported: **Mar 12, 2026**

Project Analyst:
 Joseph Lape, *Joseph Lape*

Date:
03 - 12 - 2026

Reviewed By:
 David McDonald, PHR *David McDonald*

Date:
03 - 12 - 2026

Sample Number*	5	A - 5		6	A - 6		7	A - 7		8	A - 8	
Sample Name*	Main Library			Room 26			Room 15			Nurse's Room		
Sample Volume*	150 L			150 L			150 L			150 L		
Reporting Limit	7 spores/m³			7 spores/m³			7 spores/m³			7 spores/m³		
Background	2			2			2			3		
Fragments	ND			7/m³			ND			7/m³		
Organism	Raw Count	Count / m³	% of Total	Raw Count	Count / m³	% of Total	Raw Count	Count / m³	% of Total	Raw Count	Count / m³	% of Total
Alternaria	1	7	50.0%									
Ascospores	1	7	50.0%	12	80	57.1%	3	20	23.1%	8	53	18.2%
Aspergillus Penicillium										7	47	15.9%
Basidiospores				4	27	19.0%	4	27	30.8%	21	140	47.7%
Bipolaris Drechslera												
Chaetomium										2	13	4.5%
Cladosporium				3	20	14.3%	6	40	46.2%	5	33	11.4%
Curvularia												
Epicoccum												
Fusarium												
Memnoniella												
Myxomycetes/Periconia				1	7	4.8%				1	7	2.3%
Pithomyces				1	7	4.8%						
Rusts/Smuts												
Stachybotrys												
Stemphylium												
Torula												
Ulocladium												
Total	2	14	100%	21	141	100%	13	87	100%	44	293	100%

Water Damage Indicator	Common Allergen	Slightly Higher than Baseline	Significantly Higher than Baseline	Ratio Abnormality
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* indicates data provided by the customer



Collected: **Mar 11, 2026**

Received: **Mar 12, 2026**

Reported: **Mar 12, 2026**

Project Analyst:
 Joseph Lape, *Joseph Lape*

Date:
03 - 12 - 2026

Reviewed By:
 David McDonald, PHR *David McDonald*

Date:
03 - 12 - 2026

Sample Number*	9	A - 9		10	A - 10			
Sample Name*	Main Office			Band Room				
Sample Volume*	150 L			150 L				
Reporting Limit	7 spores/m³			7 spores/m³				
Background	3			2				
Fragments	13/m³			ND				
Organism	Raw Count	Count / m³	% of Total	Raw Count	Count / m³	% of Total		
Alternaria								
Ascospores	2	13	25.0%	3	20	33.3%		
Aspergillus Penicillium	4	27	50.0%					
Basidiospores				2	13	22.2%		
Bipolaris Drechslera								
Chaetomium								
Cladosporium	1	7	12.5%	4	27	44.4%		
Curvularia								
Epicoccum								
Fusarium								
Memnoniella								
Myxomycetes Periconia	1	7	12.5%					
Pithomyces								
Rusts Smuts								
Stachybotrys								
Stemphylium								
Torula								
Ulocladium								
Total	8	54	100%	9	60	100%		

Water Damage Indicator	Common Allergen	Slightly Higher than Baseline	Significantly Higher than Baseline	Ratio Abnormality
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* indicates data provided by the customer

Spore Trap Information

Reporting Limit	The Reporting Limit is the lowest number of spores that can be detected based on the total volume of the sample collected and the percentage of the slide that is counted. At Hayes Microbial, 100% of the slide is read so the LOD is based solely on the total volume. Raw spore counts that exceed 500 spores will be estimated.
Blanks	Results have not been corrected for field or laboratory blanks.
Background	The Background is the amount of debris that is present in the sample. This debris consists of skin cells, dirt, dust, pollen, drywall dust and other organic and non-organic matter. As the background density increases, the likelihood of spores, especially small spores such as those of <i>Aspergillus</i> and <i>Penicillium</i> may be obscured. The background is rated on a scale of 1 to 5 and each level is determined as follows: NBD: No background detected due to possible pump or cassette malfunction. Recollect sample. (Field Blanks will display NBD) 1 : <5% of field occluded. No spores will be uncountable. 2 : 5-25% of field occluded. 3 : 25-75% of field occluded. 4 : 75-90% of field occluded. 5 : >90% of field occluded. Suggested recollection of sample.
Fragments	Fragments are small pieces of fungal mycelium or spores. They are not identifiable as to type and when present in very large numbers, may indicate the presence of mold amplification.
Control Comparisons	There are no national standards for the numbers of fungal spores that may be present in the indoor environment. As a general rule and guideline that is widely accepted in the indoor air quality field, the numbers and types of spores that are present in the indoor environment should not exceed those that are present outdoors at any given time. There will always be some mold spores present in "normal" indoor environments. The purpose of sampling and counting spores is to help determine whether an abnormal condition exists within the indoor environment and if it does, to help pinpoint the area of contamination. Spore counts should not be used as the sole determining factor of mold contamination. There are many factors that can cause anomalies in the comparison of indoor and outdoor samples due to the dynamic nature of both of those environments.
Water Damage Indicator	Blue: These molds are commonly seen in conditions of prolonged water intrusion and usually indicate a problem.
Common Allergen	Green: Although all molds are potential allergens, these are the most common allergens that may be found indoors.
Slightly Higher than Baseline	Orange: The spore count is slightly higher than the outside count and may or may not indicate a source of contamination.
Significantly Higher than Baseline	Red: The spore count is significantly higher than the baseline count and probably indicates a source of contamination.
Ratio Abnormality	Violet: The types of spores found indoors should be similar to the ones that were identified in the baseline sample. Significant increases (more than 25%) in the ratio of a particular spore type may indicate the presence of abnormal levels of mold, even if the total number of spores of that type is lower in the indoor environment than it was outdoors.
Color Coding	Fungi that are present in indoor samples at levels lower than 200 per cubic meter are not color coded on the report, unless they are one of the water damage indicators.
Significant Figures	Raw counts and column totals may reflect more than 2 significant figures, but results should only be considered significant to 2 figures.

Analyte Descriptions

Alternaria	Habitat:	Commonly found outdoors in soil and decaying plants. Indoors, it is commonly found on window sills and other horizontal surfaces.
	Health Effects:	A common allergen and has been associated with hypersensitivity pneumonitis. Alternaria is capable of producing toxic metabolites which may be associated with disease in humans or animals. Occasionally an agent of onychomycosis, ulcerated cutaneous infection and chronic sinusitis, principally in the immunocompromised patient.
Ascospores	Habitat:	A large group consisting of more than 3000 species of fungi. Common plant pathogens and outdoor numbers become very high following rain. Most of the genera are indistinguishable by spore trap analysis and are combined on the report.
	Health Effects:	Health affects are poorly studied, but many are likely to be allergenic.
Aspergillus Penicillium	Habitat:	The most common fungi isolated from the environment. Very common in soil and on decaying plant material. Are able to grow well indoors on a wide variety of substrates.
	Health Effects:	This group contains common allergens and many can cause hypersensitivity pneumonitis. They may cause extrinsic asthma, and many are opportunistic pathogens. Many species produce mycotoxins which may be associated with disease in humans and other animals. Toxin production is dependent on the species, the food source, competition with other organisms, and other environmental conditions.
Basidiospores	Habitat:	A common group of Fungi that includes the mushrooms and bracket fungi. They are saprophytes and plant pathogens. In wet conditions they can cause structural damage to buildings.
	Health Effects:	Common allergens and are also associated with hypersensitivity pneumonitis.
Chaetomium	Habitat:	Ascomycete fungus, commonly isolated from soil and decaying plant materials. It is cellulolytic and grows well indoors on damp sheetrock and other paper substrates. It is often found growing with Stachybotrys.
	Health Effects:	It is reported to be allergenic and may produce toxins.
Cladosporium	Habitat:	One of the most common genera worldwide. Found in soil and plant debris and on the leaf surfaces of living plants. The outdoor numbers are lower in the winter and often relatively high in the summer, especially in high humidity. The outdoor numbers often spike in the late afternoon and evening. Indoors, it can be found growing on textiles, wood, sheetrock, moist window sills and in HVAC supply ducts.
	Health Effects:	A common allergen, producing more than 10 allergenic antigens and a common cause of hypersensitivity pneumonitis.

Analyte Descriptions

Curvularia	Habitat:	They exist in soil and plant debris, and are plant pathogens.
	Health Effects:	They are allergenic and a common cause of allergic fungal sinusitis. An occasional cause of human infection, including keratitis, sinusitis, onychomycosis, mycetoma, pneumonia, endocarditis and disseminated infection, primarily in the immunocompromised.
Epicoccum	Habitat:	It is found in soil and plant litter and is a plant pathogen. It can grow indoors on a variety of substrates, including paper and textiles and is commonly found on wet drywall.
	Health Effects:	It is a common allergen. No cases of infection have been reported in humans.
Myxomycetes/Periconia	Description:	Found on decaying plant material and as a plant pathogen.
	Health Effects:	Some allergenic properties reported, but generally pose no health concerns to humans.
Pithomyces	Habitat:	Common fungus isolated from soil, decaying plant material. Rarely found indoors.
	Health Effects:	Allergenic properties are poorly studied. No cases of infection in humans.



Cleaner environment. Safer workplaces.

Mold Inspection & Moisture Report

For

Ann Antolini Elementary School
30 Antolini Rd New Hartford, CT 06057

Nurse's Room

Prepared For

New Hartford Public Schools 530 Main
Street
New Hartford, CT

Date of Inspection:

July 10, 2026

EnviroMed Project

IH-26-2462

EnviroMed Services, Inc.
470 Murdock Ave., Meriden, CT 06450
telephone (203) 238-4846 • facsimile (203) 238-4243

TABLE OF CONTENTS

Section	Page
I. PROJECT NARRATIVE.....	1
Overview	1
Methods	1
Summary of Results	2
Conclusions and Recommendations	3
II. PHOTOGRAPHS	4
III. LABORATORY ANALYSIS REPORT	8
IV. ADDITIONAL NOTES.....	9

I. PROJECT NARRATIVE

Overview

On July 10, 2026, EnviroMed Services, Inc. conducted a Mold Inspection & moisture inspection at Ann Antolini Elementary School, New Hartford, CT. The purpose of this assessment, as directed by the client, was to survey for airborne fungal contamination within the Nurse's Room.

Methods

The following sampling and assessment scheme was employed:

- Conduct air sampling to test for the presence of airborne fungal structures, including spores, by drawing air through a Zefon Air-O-Cell cassette using a pre-calibrated vacuum pump. In this method, air enters the cassette and particles are collected on the sampling substrate. The manufacturer-recommended flow rate of 15 liters per minute of air, for a duration of 10 minutes, was used to collect each sample. Sample locations were selected based on client direction. The samples were sent to Hayes Microbial Consulting in Midlothian, VA, for analysis by optical microscopy. Hayes Microbial Consulting is approved by the American Industrial Hygiene Association (AIHA Cert# 188863) and the State of Connecticut Department of Public Health. The results are represented in counts of fungal structures and fungal structures per cubic meter of air (spores/m³). The results are compared to the Baseline sample that was collected outdoors.
- Monitoring for moisture was performed using a General Tools MM8 Pinless Moisture Meter. The scan function of the meter was used, which measures readings in REL (relative content). REL – Readings on the relative content are used for wallboard and wood. The REL value is a theoretical moisture level indicating potential severity of the moisture content with the material ranging from 0 to 100%. The dry area is below 25%, moderately wet area is between 26% and 75% and excessively wet is greater than 76%.

Summary of Results

Analyte Descriptions

Acronium: Found in soil and decaying plant debris. This species is a type I and Type III allergen. It may cause mycetoma, infections of the nails, corneal ulcers, meningitis, and endocarditis.

Ascospores: A Large group consisting of more than 3000 species of fungi. Common plant pathogens and outdoor numbers become very high following rain. Most of the genera are indistinguishable by spore trap analysis and are combined on the report. Health effects are poorly studied, but many are likely to be allergenic.

Aspergillus/Penicillium: The most common fungi isolated from the environment. Very common in soil and on decaying plant material. Are able to grow well indoors on a wide variety of substrates. This group contains common allergens, and many can cause hypersensitive pneumonitis.

Basidiospores: A common group of fungi that includes the mushroom and bracket fungi. They are saprophytes and plant pathogens. In wet conditions, they can cause structural damage to buildings. Effects are that they are common allergens and are also associated with hypersensitive pneumonitis.

Cladosporium: A common genus worldwide. Found in soil and plant debris and on the leaf surfaces of living plants. Indoors, it can be found growing on textiles, wood, sheetrock, moist windowsills and in HVAC supply ducts. It is a common allergen, producing more than 10 allergenic antigens and a common cause of hypersensitivity pneumonitis.

Curvularia: Found in soil and plant debris and are plant pathogens. This species is allergenic and a common cause of allergic fungal sinusitis. They are an occasional cause for infections in human that result in keratitis, sinusitis, onychomycosis, mycetoma, pneumonia, endocarditis, and disseminated infection.

Myxomycetes: Found on decaying plant material and as a plant pathogen. Some allergenic properties are reported, but generally pose no health concerns to humans.

Stachybotrys: Commonly found in soil and on decaying plant material. Allergenic properties are poorly studied and no cases of infection have been reported in humans. They do however produce potent tricothecene mycotoxins. The toxins produced by this fungus can suppress the immune system affecting the lymphoid tissue and the bone marrow. The mycotoxin is also reported to be a liver and kidney carcinogen.

Air-O-Cell Spore Trap Sampling Results:

There are currently no federal or State of Connecticut regulatory standards for concentrations of mold in air. However, some commonly used guidelines have been established. Levels of mold within the building are routinely compared to the baseline levels outside the building to detect any amplifications or unusual mold species as compared to the natural airborne conditions of that day.

Section III of this report contains a copy of the analytical report.

Sample ID: A1**Sample Location: Outdoor Baseline**

The prominent fungal genera present was *Ascospores*. The other fungal genera present were *Alternaria*, *Aspergillus/Penicillium*, *Basidiospores*, *Cladosporium*, *Helicomyces*, and *Oidium*.

Sample ID: A2**Sample Location: Nurse's Room**

The prominent fungal genera present was *Cladosporium*. The other fungal genera present were *Ascospores*, *Basidiospores*, *Epicoccum*, *Helicomyces* and *Myxomycetes/Periconia*.

There was no amplification detected for any of the fungal genera as compared to the outdoor baseline sample.

Sample ID: A3**Sample Location: Nurse's Room (2)**

The prominent fungal genera present was *Cladosporium*. The other fungal genera present were *Alternaria*, *Ascospores*, *Aspergillus/Penicillium*, *Basidiospores*, and *Epicoccum*.

Conclusions and Findings

Based on the results obtained and observations made during the assessment, EnviroMed Services, Inc. has come to the following conclusions.

Conclusions

Air Samples

- There is no evidence of airborne toxigenic mold in the Nurse's Room.
- There is no evidence of airborne mold amplification in the Nurse's Room.

Based on our review of the airborne mold samples collected within the Nurse's Room at Ann Antolini School, located at 30 Antolini Road, New Hartford, Ct on July 10, 2026, EnviroMed Services, Inc. has determined that the areas tested are fit for occupancy. Laboratory analysis identifies no evidence of airborne mold amplification or the presence of toxigenic mold within areas evaluated.

II. PHOTOGRAPHS



Figure 1. Photograph of the Outdoor Baseline sample, A1.



Figure 2. Photograph of the Nurse's Office sample, A2.



Figure 3. Photograph of the Nurse's Office sample, A3.

III. LABORATORY ANALYSIS REPORT

Analysis Report prepared for

EnviroMed Services, Inc.

470 Murdock Avenue
Meriden, CT 06450

Phone: (203) 238-4846

IH-26-2462
New Hartford Public Schools
Ann Antolini Elementary School

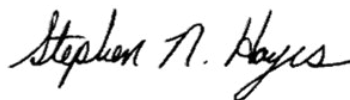
Collected: **July 10, 2026**
Received: **July 13, 2026**
Reported: **July 13, 2026**

We would like to thank you for trusting Hayes Microbial for your analytical needs!
We received 3 samples by FedEx in good condition for this project on July 13th, 2026.

The results in this analysis pertain only to this job, collected on the stated date, and should not be used in the interpretation of any other job. Information supplied by the customer can affect the validity of results. These results apply only to the samples as received. This report may not be duplicated, except in full, without the written consent of Hayes Microbial Consulting, LLC.

All information provided to Hayes Microbial is confidential information relating to our customers and their clients. We will not disclose, copy, or distribute any information verbally or written, except to those designated by the customer(s). We take confidentiality very seriously. No changes to the distribution list will be made without the express consent of the customer.

This laboratory bears no responsibility for sample collection activities, analytical method limitations, or your use of the test results. Interpretation and use of test results are your responsibility. Any reference to health effects or interpretation of mold levels is strictly the opinion of Hayes Microbial. In no event, shall Hayes Microbial or any of its employees be liable for lost profits or any special, incidental or consequential damages arising out of the use of these test results.



Steve Hayes, BSMT (ASCP)
Laboratory Director
Hayes Microbial Consulting, LLC.



EPA Laboratory ID: VA01419



Lab ID: #188863



DPH License: #PH-0198

Sample Number*	1	A1		2	A2		3	A3				
Sample Name*	Outdoor Baseline			Nurse's Room			Nurse's Room					
Sample Volume*	150 L			150 L			150 L					
Reporting Limit	7 spores/m³			7 spores/m³			7 spores/m³					
Background	2			2			2					
Fragments	7/m³			ND			ND					
Organism	Raw Count	Count / m³	% of Total	Raw Count	Count / m³	% of Total	Raw Count	Count / m³	% of Total			
Alternaria	1	7	<1%				1	7	2.0%			
Ascospores	245	1600	46.1%	5	33	12.2%	8	53	16.3%			
Aspergillus Penicillium	4	27	<1%				2	13	4.1%			
Basidiospores	154	1000	29.0%	3	20	7.3%	6	40	12.2%			
Bipolaris Drechslera												
Chaetomium												
Cladosporium	120	800	22.6%	28	190	68.3%	31	210	63.3%			
Curvularia												
Epicoccum				2	13	4.9%	1	7	2.0%			
Fusarium												
Helicomyces	6	40	1.1%	1	7	2.4%						
Memnoniella												
Myxomycetes/Periconia				2	13	4.9%						
Oidium	1	7	<1%									
Pithomyces												
Rusts/Smuts												
Stachybotrys												
Stemphylium												
Torula												
Ulocladium												
Total	531	3481	100%	41	276	100%	49	330	100%			

Water Damage Indicator	Common Allergen	Slightly Higher than Baseline	Significantly Higher than Baseline	Ratio Abnormality
------------------------	-----------------	-------------------------------	------------------------------------	-------------------

* indicates data provided by the customer

Spore Trap Information

Reporting Limit	The Reporting Limit is the lowest number of spores that can be detected based on the total volume of the sample collected and the percentage of the slide that is counted. At Hayes Microbial, 100% of the slide is read so the LOD is based solely on the total volume. Raw spore counts that exceed 500 spores will be estimated.
Blanks	Results have not been corrected for field or laboratory blanks.
Background	The Background is the amount of debris that is present in the sample. This debris consists of skin cells, dirt, dust, pollen, drywall dust and other organic and non-organic matter. As the background density increases, the likelihood of spores, especially small spores such as those of <i>Aspergillus</i> and <i>Penicillium</i> may be obscured. The background is rated on a scale of 1 to 5 and each level is determined as follows: NBD: No background detected due to possible pump or cassette malfunction. Recollect sample. (Field Blanks will display NBD) 1 : <5% of field occluded. No spores will be uncountable. 2 : 5-25% of field occluded. 3 : 25-75% of field occluded. 4 : 75-90% of field occluded. 5 : >90% of field occluded. Suggested recollection of sample.
Fragments	Fragments are small pieces of fungal mycelium or spores. They are not identifiable as to type and when present in very large numbers, may indicate the presence of mold amplification.
Control Comparisons	There are no national standards for the numbers of fungal spores that may be present in the indoor environment. As a general rule and guideline that is widely accepted in the indoor air quality field, the numbers and types of spores that are present in the indoor environment should not exceed those that are present outdoors at any given time. There will always be some mold spores present in "normal" indoor environments. The purpose of sampling and counting spores is to help determine whether an abnormal condition exists within the indoor environment and if it does, to help pinpoint the area of contamination. Spore counts should not be used as the sole determining factor of mold contamination. There are many factors that can cause anomalies in the comparison of indoor and outdoor samples due to the dynamic nature of both of those environments.
Water Damage Indicator	Blue: These molds are commonly seen in conditions of prolonged water intrusion and usually indicate a problem.
Common Allergen	Green: Although all molds are potential allergens, these are the most common allergens that may be found indoors.
Slightly Higher than Baseline	Orange: The spore count is slightly higher than the outside count and may or may not indicate a source of contamination.
Significantly Higher than Baseline	Red: The spore count is significantly higher than the baseline count and probably indicates a source of contamination.
Ratio Abnormality	Violet: The types of spores found indoors should be similar to the ones that were identified in the baseline sample. Significant increases (more than 25%) in the ratio of a particular spore type may indicate the presence of abnormal levels of mold, even if the total number of spores of that type is lower in the indoor environment than it was outdoors.
Color Coding	Fungi that are present in indoor samples at levels lower than 200 per cubic meter are not color coded on the report, unless they are one of the water damage indicators.
Significant Figures	Raw counts and column totals may reflect more than 2 significant figures, but results should only be considered significant to 2 figures.

Analyte Descriptions

Alternaria	Habitat:	Commonly found outdoors in soil and decaying plants. Indoors, it is commonly found on window sills and other horizontal surfaces.
	Health Effects:	A common allergen and has been associated with hypersensitivity pneumonitis. Alternaria is capable of producing toxic metabolites which may be associated with disease in humans or animals. Occasionally an agent of onychomycosis, ulcerated cutaneous infection and chronic sinusitis, principally in the immunocompromised patient.
Ascospores	Habitat:	A large group consisting of more than 3000 species of fungi. Common plant pathogens and outdoor numbers become very high following rain. Most of the genera are indistinguishable by spore trap analysis and are combined on the report.
	Health Effects:	Health affects are poorly studied, but many are likely to be allergenic.
Aspergillus Penicillium	Habitat:	The most common fungi isolated from the environment. Very common in soil and on decaying plant material. Are able to grow well indoors on a wide variety of substrates.
	Health Effects:	This group contains common allergens and many can cause hypersensitivity pneumonitis. They may cause extrinsic asthma, and many are opportunistic pathogens. Many species produce mycotoxins which may be associated with disease in humans and other animals. Toxin production is dependent on the species, the food source, competition with other organisms, and other environmental conditions.
Basidiospores	Habitat:	A common group of Fungi that includes the mushrooms and bracket fungi. They are saprophytes and plant pathogens. In wet conditions they can cause structural damage to buildings.
	Health Effects:	Common allergens and are also associated with hypersensitivity pneumonitis.
Cladosporium	Habitat:	One of the most common genera worldwide. Found in soil and plant debris and on the leaf surfaces of living plants. The outdoor numbers are lower in the winter and often relatively high in the summer, especially in high humidity. The outdoor numbers often spike in the late afternoon and evening. Indoors, it can be found growing on textiles, wood, sheetrock, moist window sills and in HVAC supply ducts.
	Health Effects:	A common allergen, producing more than 10 allergenic antigens and a common cause of hypersensitivity pneumonitis.
Epicoccum	Habitat:	It is found in soil and plant litter and is a plant pathogen. It can grow indoors on a variety of substrates, including paper and textiles and is commonly found on wet drywall.
	Health Effects:	It is a common allergen. No cases of infection have been reported in humans.

Analyte Descriptions

Helicomycetes	Habitat:	Found on wood and decaying plant matter.
	Health Effects:	Health effects are poorly studied.
Myxomycetes/Periconia	Description:	Found on decaying plant material and as a plant pathogen.
	Health Effects:	Some allergenic properties reported, but generally pose no health concerns to humans.
Oidium	Habitat:	Plant pathogen commonly known as powdery mildew.
	Health Effects:	Health effects are poorly studied.

IV. ADDITIONAL NOTES

Airborne mold concentrations (levels) and/or surfaces may change over a period of time. The sample results are a representation of the indoor air quality and/or surfaces at the time the test was performed. Mold may grow in indoor environments without producing airborne mold spores or hyphae. EnviroMed cannot guarantee the identification of all/every mold growth area based on airborne testing. EnviroMed Services Inc. is not liable for inaccessible areas, illness, allergies, or damage to your property as a result of this report's findings. EnviroMed Services Inc. is not liable for the outcome of the remediation process/completion as a result of this report's findings.



AIHA Laboratory Accreditation Programs, LLC
acknowledges that
Hayes Microbial Consulting
3005 E. Boundary Terrace, Suite F Midlothian, VA 23112
Laboratory ID: LAP-188863

along with all premises from which key activities are performed, as listed above, has fulfilled the requirements of the AIHA Laboratory Accreditation Programs, LLC (AIHA LAP) accreditation to the ISO/IEC 17025:2017 international standard, General Requirements for the Competence of Testing and Calibration Laboratories in the following:

LABORATORY ACCREDITATION PROGRAMS

☐	INDUSTRIAL HYGIENE	Accreditation Expires:
☐	ENVIRONMENTAL LEAD	Accreditation Expires:
☒	ENVIRONMENTAL MICROBIOLOGY	Accreditation Expires: April 01, 2027
☐	FOOD	Accreditation Expires:
☐	UNIQUE SCOPES	Accreditation Expires:
☐	BE FIELD/MOBILE	Accreditation Expires:

Specific Field(s) of Testing/Method(s) within each Accreditation Program for which the above named laboratory maintains accreditation is outlined on the attached Scope of Accreditation. Continued accreditation is contingent upon successful on-going compliance with ISO/IEC 17025:2017 and AIHA LAP requirements. This certificate is not valid without the attached Scope of Accreditation. Please review the AIHA LAP website (www.aihaaccreditedlabs.org) for the most current Scope.

A handwritten signature in black ink that reads 'Cheryl O. Morton'.

Cheryl O Morton
Managing Director, AIHA Laboratory Accreditation Programs, LLC



AIHA Laboratory Accreditation Programs, LLC

SCOPE OF ACCREDITATION

Hayes Microbial Consulting

3005 E. Boundary Terrace, Suite F Midlothian, VA
23112

Laboratory ID: LAP-188863

Issue Date: 05/01/2023

The laboratory is approved for those specific field(s) of testing/methods listed in the table below. Clients are urged to verify the laboratory's current accreditation status for the particular field(s) of testing/Methods, since these can change due to proficiency status, suspension and/or withdrawal of accreditation.

Environmental Microbiology Laboratory Accreditation Program (EMLAP)

Initial Accreditation Date: 08/01/2010

EMLAP Scope Category	Field of Testing (FOT)	Component, parameter or characteristic tested	Method	Method Description (for internal methods only)
Fungal	Air - Culturable	Air	HMC-#103	In House: Viable Air Culture
Fungal	Air - Direct Examination	Spore Trap	HMC-#101	In House: Spore Trap Analysis
Fungal	Bulk - Culturable	Bulk Material	HMC-#104	In House: Bulk Culture Analysis
Fungal	Bulk - Direct Examination	Bulk Material	HMC-#102	In House: Direct Identification Analysis
Fungal	Surface - Culturable	Swab	HMC-#105	In House: Swab Culture Analysis
Fungal	Surface - Direct Examination	Bio-Tape, Swab	HMC-#102	In House: Direct Identification Analysis

A complete listing of currently accredited EMLAP laboratories is available on the AIHA LAP, LLC website at:
<http://www.aihaaccreditedlabs.org>