



September 26, 2025

Ms. Sheila Downs  
Assistant Superintendent of Operations  
School District U-46  
1460 Sheldon Drive  
Elgin, Illinois 60120

**RE: Mold Air Sampling Report  
Hanover Countryside Elementary School  
6 South Barlett Road, Streamwood, Illinois 60107  
WCG Project No. 4366-321-60-01**

Dear Ms. Downs:

Weaver Consultants Group North Central, LLC (WCG) was retained to conduct mold air sampling throughout Hanover Countryside Elementary School. WCG representatives Mr. Austin E. Lamp and Mr. Nicholas P. Liming conducted the air sampling on September 23, 24, and 25, 2025 for viable and non-viable mold spores under the direction of Mr. Cody R. McNeely, CIH.

## **METHODOLOGY**

### **Airborne Mold Spores (Viable and Non-Viable)**

Total fungal structures (non-culturable) air samples were obtained using Air-O-Cell cassettes connected to a hand-held, battery-powered Bio-Pump. The pump flow rate was established at 15 liters per minute for 5 minutes to collect a total air volume of 75 liters. Following sample collection, the cassettes were sealed and sent along with a chain of custody record to Eurofins Built Environment Testing Central, LLC. (Eurofins) in Naperville, Illinois for analysis. Eurofins is accredited by the American Industrial Hygiene Association (AIHA) Environmental Microbiology Laboratory Accreditation Program (EMLAP). Air-O-Cell cassettes were analyzed by direct microscopy. Results were converted to fungal structures per cubic meter of air (fungal structures/m<sup>3</sup>) based on the duration of sampling and the corresponding volume of air collected. Results were tabulated by rank order of genera to facilitate data interpretation in accordance with the American Conference of Governmental Industrial Hygienists (ACGIH) guideline in "Bioaerosols Assessment and Control" (1999).

## **RESULTS**

Results summarized below reflect conditions at the time of the mold air sampling. See Appendix A Table I for a summary of airborne mold spore sample results and Appendix B for the laboratory analytical report, chain of custody documentation, and laboratory accreditation.

The general evaluation criteria for airborne fungal spores (total fungal structures) in an indoor environment require indoor mold spore counts to be lower than outdoor counts. Such a criteria represents a normal finding in a building furnished with mechanical ventilation and filtration. However, outdoor fungal spores can migrate into buildings when occupants open/close doors and windows. Additionally, indoor fungal types (genera) are typically consistent with the types obtained on contemporaneously collected outdoor air samples or are otherwise common outdoor types.

The total indoor mold air sample results ranged between < 13 and 1,100 fungal structures/m<sup>3</sup>. Results for the exterior samples ranged between 10,000 and 17,000 fungal structures/m<sup>3</sup>. In general, mold air sample results were judged to be normal and typical of an indoor environment and conformed to the aforementioned criteria; however, the presence of *Stachybotrys* spores in sample AL092325-06 (Hallway by Conference Room 3D/Classroom 4) may be an indicator of a historic and/or current indoor source of mold growth. This is not a definitive conclusion and must be used in conjunction with visual observations, water intrusion history, and overall cleanliness and housekeeping.

## CONCLUSIONS

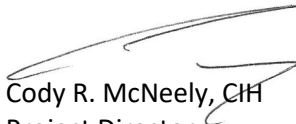
Based on the results of the mold air sampling, WCG makes the following conclusions:

- Mold air sampling throughout the building as well as supplemental mold air sampling in the Main Office and areas adjacent to the Hallway by Conference Room 3D/Classroom 4 indicated the indoor mold spore counts were similar in composition and lower than contemporaneously collected outdoor air samples and represent a normal and typical finding in a building furnished with mechanical ventilation and filtration.

We appreciate the opportunity to assist you on this project. Should you have any questions or need additional information, please do not hesitate to contact me at (312) 806-2235.

Sincerely,

**Weaver Consultants Group North Central, LLC**



Cody R. McNeely, CH  
Project Director

## **APPENDIX A**

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Table I - Summary of Airborne Mold Spore Sample Results

Table I - Summary of Airborne Mold Spores Sample Results  
 Hanover Countryside Elementary School  
 6 South Bartlett Road, Streamwood, Illinois 60107

Sample Number	Sample Location	Laboratory Results	Comments
AL092325-01	Classroom 24	<i>Ascospores, Basidiospores, and Smuts, Periconia, Myxomycetes</i>	Normal and Typical of an Indoor Environment
AL092325-02	Classroom 22	<i>Smuts, Periconia, Myxomycetes</i>	Normal and Typical of an Indoor Environment
AL092325-03	Hallway by Commons 27	<i>Ascospores, Curvularia, Pithomyces, and Smuts, Periconia, Myxomycetes</i>	Normal and Typical of an Indoor Environment
AL092325-04	Classroom 2	<i>Alternaria</i>	Normal and Typical of an Indoor Environment
AL092325-05	Classroom 4	No Spores Detected	Normal and Typical of an Indoor Environment
AL092325-06	Hallway by Conference Room 3D/Classroom 4	<i>Alternaria, Curvularia, Pithomyces, Smuts, Periconia, Myxomycetes, and Stachybotrys</i>	<p>The presence of <i>Stachybotrys</i> spores may be an indicator of a historic and/or current indoor source of mold growth. This is not a definitive conclusion and must be used in conjunction with visual observations, water intrusion history, and overall cleanliness and housekeeping.</p> <p>Reference samples NL092425-18 and NL092425-20 for comparable resample locations</p>
AL092325-07	Staffroom 4D	<i>Smuts, Periconia, Myxomycetes</i>	Normal and Typical of an Indoor Environment
AL092325-08	Classroom 6	<i>Ascospores and Basidiospores</i>	Normal and Typical of an Indoor Environment

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Sample Number	Sample Location	Laboratory Results	Comments
AL092325-09	Classroom 8	<i>Basidiospores, Cladosporium, and Other Brown</i>	Normal and Typical of an Indoor Environment
AL092325-10	Classroom 13	<i>Ascospores and Basidiospores</i>	Normal and Typical of an Indoor Environment
AL092325-11	Hallway by Classroom 8	No Spores Detected	Normal and Typical of an Indoor Environment
AL092325-12	Classroom 41	No Spores Detected	Normal and Typical of an Indoor Environment
AL092325-13	Hallway by Classroom 41	<i>Ascospores, Basidiospores, Cladosporium, and Epicoccum</i>	Normal and Typical of an Indoor Environment
AL092325-14	Classroom 14	No Spores Detected	Normal and Typical of an Indoor Environment
AL092325-15	Classroom 15	No Spores Detected	Normal and Typical of an Indoor Environment
AL092325-16	Building Exterior	<i>Alternaria, Ascospores, Aureobasidium, Basidiospores, Cercospora, Cladosporium, Curvularia, Epicoccum, Pithomyces, Rusts, and Smuts, Periconia, Myxomycetes</i>	N/A
AL092325-17	Building Exterior	<i>Alternaria, Ascospores, Basidiospores, Cercospora, Cladosporium, Epicoccum, Pithomyces, Rusts, and Taeniolella</i>	N/A
NL092425-01	Classroom 23	<i>Cladosporium, Rusts, and Smuts, Periconia, Myxomycetes</i>	Normal and Typical of an Indoor Environment

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Sample Number	Sample Location	Laboratory Results	Comments
NL092425-02	Learning Center 28	<i>Basidiospores</i>	Normal and Typical of an Indoor Environment
NL092425-03	Classroom 25	No Spores Detected	Normal and Typical of an Indoor Environment
NL092425-04	Kitchen 26	<i>Rusts</i>	Normal and Typical of an Indoor Environment
NL092425-05	Classroom 22	No Spores Detected	Normal and Typical of an Indoor Environment
NL092425-06	Classroom 20	<i>Basidiospores, Epicoccum, and Rusts</i>	Normal and Typical of an Indoor Environment
NL092425-07	Commons 27	<i>Ascospores, Basidiospores, and Cladosporium</i>	Normal and Typical of an Indoor Environment
NL092425-08	Maintenance Office (Kitchen 29)	No Spores Detected	Normal and Typical of an Indoor Environment
NL092425-09	Girls Toilet Room 29	<i>Epicoccum, Other Brown, and Smuts, Periconia, Myxomycetes</i>	Normal and Typical of an Indoor Environment
NL092425-10	Boys Toilet Room 30	<i>Basidiospores and Cladosporium</i>	Normal and Typical of an Indoor Environment
NL092425-11	Main Office 1	No Spores Detected	Normal and Typical of an Indoor Environment
NL092425-12	Principals Office 1A	<i>Ascospores, Other Brown, and Pithomyces</i>	Normal and Typical of an Indoor Environment

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Sample Number	Sample Location	Laboratory Results	Comments
NL092425-13	Door 1 Vestibule	<i>Pithomyces</i> and <i>Rusts</i>	Normal and Typical of an Indoor Environment
NL092425-14	Nurses Office 2B	No Spores Detected	Normal and Typical of an Indoor Environment
NL092425-15	Teachers Work Room 3	<i>Pithomyces</i>	Normal and Typical of an Indoor Environment
NL092425-16	Office 1B	<i>Ascospores, Basidiospores, Cladosporium, Curvularia, Epicoccum, Pithomyces, and Rusts</i>	Normal and Typical of an Indoor Environment
NL092425-17	Office 1C	<i>Cladosporium, Pithomyces, Rusts, and Smuts, Periconia, Myxomycetes</i>	Normal and Typical of an Indoor Environment
NL092425-18	Conference Room 3D	<i>Ascospores, Epicoccum, and Pithomyces</i>	Normal and Typical of an Indoor Environment
NL092425-19	Girls Toilet Room 3A	<i>Basidiospores</i> and <i>Pithomyces</i>	Normal and Typical of an Indoor Environment
NL092425-20	Boys Toilet Room 3B	<i>Ascospores</i> and <i>Torula</i>	Normal and Typical of an Indoor Environment
NL092425-21	Gymnasium 100	<i>Basidiospores</i>	Normal and Typical of an Indoor Environment
NL092425-22	Classroom 5	No Spores Detected	Normal and Typical of an Indoor Environment
NL092425-23	Classroom 7	<i>Cladosporium</i>	Normal and Typical of an Indoor Environment

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Sample Number	Sample Location	Laboratory Results	Comments
NL092425-24	Hallway between Classrooms 5 and 7	No Spores Detected	Normal and Typical of an Indoor Environment
NL092425-25	Classroom 9	<i>Ascospores</i>	Normal and Typical of an Indoor Environment
NL092425-26	Classroom 10	<i>Rusts</i>	Normal and Typical of an Indoor Environment
NL092425-27	Classroom 11	No Spores Detected	Normal and Typical of an Indoor Environment
NL092425-28	Classroom 12	<i>Basidiospores</i>	Normal and Typical of an Indoor Environment
NL092425-29	Classroom 17	No Spores Detected	Normal and Typical of an Indoor Environment
NL092425-30	Classroom 19	No Spores Detected	Normal and Typical of an Indoor Environment
NL092425-31	Hallway by Classroom 12	<i>Alternaria</i>	Normal and Typical of an Indoor Environment
NL092425-32	Girls Toilet Room 8B	<i>Curvularia</i> and <i>Pithomyces</i>	Normal and Typical of an Indoor Environment
NL092425-33	Boys Toilet Room 8C	No Spores Detected	Normal and Typical of an Indoor Environment
NL092425-34	Reading Office 4B	No Spores Detected	Normal and Typical of an Indoor Environment

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Sample Number	Sample Location	Laboratory Results	Comments
NL092425-35	S.A.F.E. Office 4A	<i>Basidiospores</i> and <i>Cladosporium</i>	Normal and Typical of an Indoor Environment
NL092425-36	Gymnasium Office 100A	No Spores Detected	Normal and Typical of an Indoor Environment
NL092425-37	Music Room 40	No Spores Detected	Normal and Typical of an Indoor Environment
NL092425-38	Storage Room 42	No Spores Detected	Normal and Typical of an Indoor Environment
NL092425-39	Book Room 43	No Spores Detected	Normal and Typical of an Indoor Environment
NL092425-40	Itinerant Office 45	<i>Basidiospores, Cladosporium, Rusts, and Smuts, Periconia, Myxomycetes</i>	Normal and Typical of an Indoor Environment
NL092425-41	Building Exterior	<i>Alternaria, Ascospores, Basidiospores, Cladosporium, Epicoccum, Other Colorless, and Rusts</i>	N/A
NL092425-42	Building Exterior	<i>Alternaria, Ascospores, Basidiospores, Cladosporium, Epicoccum, and Smuts, Periconia, Myxomycetes</i>	N/A
NL092525-01	Mobile Classroom 1	<i>Smuts, Periconia, Myxomycetes</i>	Normal and Typical of an Indoor Environment
NL092525-02	Mobile Classroom 2	<i>Basidiospores</i>	Normal and Typical of an Indoor Environment

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Sample Number	Sample Location	Laboratory Results	Comments
NL092525-03	Building Exterior	<i>Alternaria, Ascospores, Basidiospores, Cladosporium, Epicoccum, Pithomyces, Rusts, and Smuts, Periconia, Myxomycetes</i>	N/A
NL092525-04	Building Exterior	<i>Alternaria, Ascospores, Basidiospores, Cladosporium, Curvularia, Epicoccum, Pithomyces, and Rusts</i>	N/A

## **APPENDIX B**

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Laboratory Report, Chain of Custody Documentation, and Laboratory Accreditation

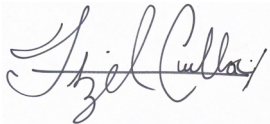
Report for:

**Austin Lamp**  
**Weaver Consultants Group**  
1316 Bond St.  
Suite 108  
Naperville, IL 60563

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Regarding: Eurofins Built Environment Testing Central, LLC  
Project: Hanover Countryside E.S.  
EML ID: 4240790

Approved by:



Business Unit Manager  
Itzel Cuellar

Dates of Analysis:

Spore trap analysis: 09-24-2025

Service SOPs: Spore trap analysis (EB-MY-S-1038)  
AIHA LAP, LLC accredited service, Lab ID #176641

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All samples were received in acceptable condition unless noted in the Report Comments portion in the body of the report. Due to the nature of the analyses performed, field blank correction of results is not applied. The results relate only to the samples as received and tested. Information supplied by the client which can affect the validity of results: sample air volume.

Eurofins Built Environment Testing Central, LLC ("the Company"), a member of the Eurofins Built Environment Testing group of companies, shall have no liability to the client or the client's customer with respect to decisions or recommendations made, actions taken or courses of conduct implemented by either the client or the client's customer as a result of or based upon the Test Results. In no event shall the Company be liable to the client with respect to the Test Results except for the Company's own willful misconduct or gross negligence nor shall the Company be liable for incidental or consequential damages or lost profits or revenues to the fullest extent such liability may be disclaimed by law, even if the Company has been advised of the possibility of such damages, lost profits or lost revenues. In no event shall the Company's liability with respect to the Test Results exceed the amount paid to the Company by the client therefor.

Eurofins Built Environment Testing Central, LLC's LabServe® reporting system includes automated fail-safes to ensure that all AIHA LAP, LLC quality requirements are met and notifications are added to reports when any quality steps remain pending.

Client: Weaver Consultants Group  
 C/O: Austin Lamp  
 Re: Hanover Countryside E.S.

Date of Sampling: 09-23-2025  
 Date of Receipt: 09-24-2025  
 Date of Report: 09-24-2025

**SPORE TRAP REPORT: NON-VIABLE METHODOLOGY**

Location:	AL092325-01: Room 24			AL092325-02: Room 22		
Comments (see below)	None			None		
Lab ID-Version‡:	21210232-1			21210233-1		
Analysis Date:	09/24/2025			09/24/2025		
	raw ct.	% read	spores/m3	raw ct.	% read	spores/m3
Alternaria						
Ascospores	1	25	53			
Aureobasidium						
Basidiospores	3	25	160			
Cercospora						
Chaetomium						
Cladosporium						
Curvularia						
Epicoccum						
Other brown						
Other colorless						
Penicillium/Aspergillus types†						
Pithomyces						
Rusts						
Smuts, Periconia, Myxomycetes	2	100	27	4	100	53
Stachybotrys						
Stemphylium						
Taeniolella						
Torula						
Ulocladium						
Zygomycetes						
Background debris (1-4+)	3+			3+		
Hyphal fragments/m3	< 13			53		
Pollen/m3	< 13			13		
Skin cells (1-4+)	2+			2+		
Sample volume (liters)	75			75		
<b>§ TOTAL SPORES/m3</b>			<b>240</b>			<b>53</b>

**Comments:**

Spore types listed without a count or data entry were not detected during the course of the analysis for the respective sample, indicating a raw count of <1 spore.

† The spores of *Aspergillus* and *Penicillium* (and others such as *Acremonium*, *Paecilomyces*) are small and round with very few distinguishing characteristics. They cannot be differentiated by non-viable sampling methods. Also, some species with very small spores are easily missed, and may be undercounted.

††Background debris indicates the amount of non-biological particulate matter present on the trace (dust in the air) and the resulting visibility for the analyst. It is rated from 1+ (low) to 4+ (high). Counts from areas with 4+ background debris should be regarded as minimal counts and may be higher than reported. It is important to account for samples volumes when evaluating dust levels.

The analytical sensitivity is the spores/m<sup>3</sup> divided by the raw count, expressed in spores/m<sup>3</sup>, per spore and per sample.

For more information regarding analytical sensitivity, please contact QA by calling the laboratory.

‡ A "Version" indicated by -"x" after the Lab ID# with a value greater than 1 indicates a sample with amended data. The revision number is reflected by the value of "x".

§ Total Spores/m<sup>3</sup> has been rounded to two significant figures to reflect analytical precision.

Client: Weaver Consultants Group  
 C/O: Austin Lamp  
 Re: Hanover Countryside E.S.

Date of Sampling: 09-23-2025  
 Date of Receipt: 09-24-2025  
 Date of Report: 09-24-2025

**SPORE TRAP REPORT: NON-VIABLE METHODOLOGY**

Location:	AL092325-03: Hallway by Commons			AL092325-04: Room 2		
Comments (see below)	None			None		
Lab ID-Version‡:	21210234-1			21210235-1		
Analysis Date:	09/24/2025			09/24/2025		
	raw ct.	% read	spores/m3	raw ct.	% read	spores/m3
Alternaria				1	100	13
Ascospores	1	25	53			
Aureobasidium						
Basidiospores						
Cercospora						
Chaetomium						
Cladosporium						
Curvularia	1	100	13			
Epicoccum						
Other brown						
Other colorless						
Penicillium/Aspergillus types†						
Pithomyces	4	100	53			
Rusts						
Smuts, Periconia, Myxomycetes	4	100	53			
Stachybotrys						
Stemphylium						
Taeniolella						
Torula						
Ulocladium						
Zygomycetes						
Background debris (1-4+)	3+			2+		
Hyphal fragments/m3	27			13		
Pollen/m3	27			< 13		
Skin cells (1-4+)	2+			1+		
Sample volume (liters)	75			75		
<b>§ TOTAL SPORES/m3</b>			<b>170</b>			<b>13</b>

**Comments:**

Spore types listed without a count or data entry were not detected during the course of the analysis for the respective sample, indicating a raw count of <1 spore.

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 C/O: Austin Lamp  
 Re: Hanover Countryside E.S.

Date of Sampling: 09-23-2025  
 Date of Receipt: 09-24-2025  
 Date of Report: 09-24-2025

**SPORE TRAP REPORT: NON-VIABLE METHODOLOGY**

Location:	AL092325-05: Room 4			AL092325-06: Hallway by Room 4		
Comments (see below)	None			None		
Lab ID-Version‡:	21210236-1			21210237-1		
Analysis Date:	09/24/2025			09/24/2025		
	raw ct.	% read	spores/m3	raw ct.	% read	spores/m3
Alternaria				2	100	27
Ascospores						
Aureobasidium						
Basidiospores						
Cercospora						
Chaetomium						
Cladosporium						
Curvularia				1	100	13
Epicoccum						
Other brown						
Other colorless						
Penicillium/Aspergillus types†						
Pithomyces				1	100	13
Rusts						
Smuts, Periconia, Myxomycetes				4	100	53
Stachybotrys				1	100	13
Stemphylium						
Taeniolella						
Torula						
Ulocladium						
Zygomycetes						
Background debris (1-4+)	2+			3+		
Hyphal fragments/m3	< 13			27		
Pollen/m3	< 13			13		
Skin cells (1-4+)	2+			2+		
Sample volume (liters)	75			75		
<b>§ TOTAL SPORES/m3</b>			< 13			120

**Comments:**

Spore types listed without a count or data entry were not detected during the course of the analysis for the respective sample, indicating a raw count of <1 spore.

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 C/O: Austin Lamp  
 Re: Hanover Countryside E.S.

Date of Sampling: 09-23-2025  
 Date of Receipt: 09-24-2025  
 Date of Report: 09-24-2025

**SPORE TRAP REPORT: NON-VIABLE METHODOLOGY**

Location:	AL092325-07: Room 4D			AL092325-08: Room 6		
Comments (see below)	None			None		
Lab ID-Version‡:	21210238-1			21210239-1		
Analysis Date:	09/24/2025			09/24/2025		
	raw ct.	% read	spores/m3	raw ct.	% read	spores/m3
Alternaria						
Ascospores				3	25	160
Aureobasidium						
Basidiospores				17	25	910
Cercospora						
Chaetomium						
Cladosporium						
Curvularia						
Epicoccum						
Other brown						
Other colorless						
Penicillium/Aspergillus types†						
Pithomyces						
Rusts						
Smuts, Periconia, Myxomycetes	1	100	13			
Stachybotrys						
Stemphylium						
Taeniolella						
Torula						
Ulocladium						
Zygomycetes						
Background debris (1-4+)	3+			3+		
Hyphal fragments/m3	13			< 13		
Pollen/m3	< 13			< 13		
Skin cells (1-4+)	1+			1+		
Sample volume (liters)	75			75		
<b>§ TOTAL SPORES/m3</b>			<b>13</b>			<b>1,100</b>

**Comments:**

Spore types listed without a count or data entry were not detected during the course of the analysis for the respective sample, indicating a raw count of <1 spore.

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 C/O: Austin Lamp  
 Re: Hanover Countryside E.S.

Date of Sampling: 09-23-2025  
 Date of Receipt: 09-24-2025  
 Date of Report: 09-24-2025

**SPORE TRAP REPORT: NON-VIABLE METHODOLOGY**

Location:	AL092325-09: Room 8			AL092325-10: Room 13		
Comments (see below)	None			None		
Lab ID-Version‡:	21210240-1			21210241-1		
Analysis Date:	09/24/2025			09/24/2025		
	raw ct.	% read	spores/m3	raw ct.	% read	spores/m3
Alternaria						
Ascospores				1	25	53
Aureobasidium						
Basidiospores	8	25	430	1	25	53
Cercospora						
Chaetomium						
Cladosporium	3	25	160			
Curvularia						
Epicoccum						
Other brown	1	25	53			
Other colorless						
Penicillium/Aspergillus types†						
Pithomyces						
Rusts						
Smuts, Periconia, Myxomycetes						
Stachybotrys						
Stemphylium						
Taeniolella						
Torula						
Ulocladium						
Zygomycetes						
Background debris (1-4+)	2+			2+		
Hyphal fragments/m3	< 13			< 13		
Pollen/m3	< 13			< 13		
Skin cells (1-4+)	2+			1+		
Sample volume (liters)	75			75		
<b>§ TOTAL SPORES/m3</b>			<b>640</b>			<b>110</b>

**Comments:**

Spore types listed without a count or data entry were not detected during the course of the analysis for the respective sample, indicating a raw count of <1 spore.

† The spores of *Aspergillus* and *Penicillium* (and others such as *Acremonium*, *Paecilomyces*) are small and round with very few distinguishing characteristics. They cannot be differentiated by non-viable sampling methods. Also, some species with very small spores are easily missed, and may be undercounted.

††Background debris indicates the amount of non-biological particulate matter present on the trace (dust in the air) and the resulting visibility for the analyst. It is rated from 1+ (low) to 4+ (high). Counts from areas with 4+ background debris should be regarded as minimal counts and may be higher than reported. It is important to account for samples volumes when evaluating dust levels.

The analytical sensitivity is the spores/m<sup>3</sup> divided by the raw count, expressed in spores/m<sup>3</sup>, per spore and per sample.

For more information regarding analytical sensitivity, please contact QA by calling the laboratory.

‡ A "Version" indicated by -"x" after the Lab ID# with a value greater than 1 indicates a sample with amended data. The revision number is reflected by the value of "x".

§ Total Spores/m<sup>3</sup> has been rounded to two significant figures to reflect analytical precision.

Client: Weaver Consultants Group  
 C/O: Austin Lamp  
 Re: Hanover Countryside E.S.

Date of Sampling: 09-23-2025  
 Date of Receipt: 09-24-2025  
 Date of Report: 09-24-2025

**SPORE TRAP REPORT: NON-VIABLE METHODOLOGY**

Location:	AL092325-11: Hallway by Room 8			AL092325-12: Room 41		
Comments (see below)	None			None		
Lab ID-Version‡:	21210242-1			21210243-1		
Analysis Date:	09/24/2025			09/24/2025		
	raw ct.	% read	spores/m3	raw ct.	% read	spores/m3
Alternaria						
Ascospores						
Aureobasidium						
Basidiospores						
Cercospora						
Chaetomium						
Cladosporium						
Curvularia						
Epicoccum						
Other brown						
Other colorless						
Penicillium/Aspergillus types†						
Pithomyces						
Rusts						
Smuts, Periconia, Myxomycetes						
Stachybotrys						
Stemphylium						
Taeniolella						
Torula						
Ulocladium						
Zygomycetes						
Background debris (1-4+)	2+			2+		
Hyphal fragments/m3	< 13			< 13		
Pollen/m3	< 13			< 13		
Skin cells (1-4+)	1+			1+		
Sample volume (liters)	75			75		
<b>§ TOTAL SPORES/m3</b>			< 13			< 13

**Comments:**

Spore types listed without a count or data entry were not detected during the course of the analysis for the respective sample, indicating a raw count of <1 spore.

† The spores of *Aspergillus* and *Penicillium* (and others such as *Acremonium*, *Paecilomyces*) are small and round with very few distinguishing characteristics. They cannot be differentiated by non-viable sampling methods. Also, some species with very small spores are easily missed, and may be undercounted.

††Background debris indicates the amount of non-biological particulate matter present on the trace (dust in the air) and the resulting visibility for the analyst. It is rated from 1+ (low) to 4+ (high). Counts from areas with 4+ background debris should be regarded as minimal counts and may be higher than reported. It is important to account for samples volumes when evaluating dust levels.

The analytical sensitivity is the spores/m<sup>3</sup> divided by the raw count, expressed in spores/m<sup>3</sup>, per spore and per sample.

For more information regarding analytical sensitivity, please contact QA by calling the laboratory.

‡ A "Version" indicated by -"x" after the Lab ID# with a value greater than 1 indicates a sample with amended data. The revision number is reflected by the value of "x".

§ Total Spores/m<sup>3</sup> has been rounded to two significant figures to reflect analytical precision.

Client: Weaver Consultants Group  
 C/O: Austin Lamp  
 Re: Hanover Countryside E.S.

Date of Sampling: 09-23-2025  
 Date of Receipt: 09-24-2025  
 Date of Report: 09-24-2025

**SPORE TRAP REPORT: NON-VIABLE METHODOLOGY**

Location:	AL092325-13: Hallway by Room 41			AL092325-14: Room 14		
Comments (see below)	None			None		
Lab ID-Version‡:	21210244-1			21210245-1		
Analysis Date:	09/24/2025			09/24/2025		
	raw ct.	% read	spores/m3	raw ct.	% read	spores/m3
Alternaria						
Ascospores	2	25	110			
Aureobasidium						
Basidiospores	1	25	53			
Cercospora						
Chaetomium						
Cladosporium	1	25	53			
Curvularia						
Epicoccum	1	100	13			
Other brown						
Other colorless						
Penicillium/Aspergillus types†						
Pithomyces						
Rusts						
Smuts, Periconia, Myxomycetes						
Stachybotrys						
Stemphylium						
Taeniolella						
Torula						
Ulocladium						
Zygomycetes						
Background debris (1-4+)	3+			2+		
Hyphal fragments/m3	< 13			< 13		
Pollen/m3	< 13			< 13		
Skin cells (1-4+)	2+			1+		
Sample volume (liters)	75			75		
<b>§ TOTAL SPORES/m3</b>			<b>230</b>			<b>&lt; 13</b>

**Comments:**

Spore types listed without a count or data entry were not detected during the course of the analysis for the respective sample, indicating a raw count of <1 spore.

† The spores of *Aspergillus* and *Penicillium* (and others such as *Acremonium*, *Paecilomyces*) are small and round with very few distinguishing characteristics. They cannot be differentiated by non-viable sampling methods. Also, some species with very small spores are easily missed, and may be undercounted.

††Background debris indicates the amount of non-biological particulate matter present on the trace (dust in the air) and the resulting visibility for the analyst. It is rated from 1+ (low) to 4+ (high). Counts from areas with 4+ background debris should be regarded as minimal counts and may be higher than reported. It is important to account for samples volumes when evaluating dust levels.

The analytical sensitivity is the spores/m<sup>3</sup> divided by the raw count, expressed in spores/m<sup>3</sup>, per spore and per sample.

For more information regarding analytical sensitivity, please contact QA by calling the laboratory.

‡ A "Version" indicated by -"x" after the Lab ID# with a value greater than 1 indicates a sample with amended data. The revision number is reflected by the value of "x".

§ Total Spores/m<sup>3</sup> has been rounded to two significant figures to reflect analytical precision.

Client: Weaver Consultants Group  
 C/O: Austin Lamp  
 Re: Hanover Countryside E.S.

Date of Sampling: 09-23-2025  
 Date of Receipt: 09-24-2025  
 Date of Report: 09-24-2025

**SPORE TRAP REPORT: NON-VIABLE METHODOLOGY**

Location:	AL092325-15: Room 15			AL092325-16: Building Exterior		
Comments (see below)	None			None		
Lab ID-Version‡:	21210246-1			21210247-1		
Analysis Date:	09/24/2025			09/24/2025		
	raw ct.	% read	spores/m3	raw ct.	% read	spores/m3
Alternaria				4	100	53
Ascospores				12	12.5	1,300
Aureobasidium				1	100	13
Basidiospores				66	12.5	7,000
Cercospora				7	100	93
Chaetomium						
Cladosporium				14	12.5	1,500
Curvularia				1	100	13
Epicoccum				7	100	93
Other brown						
Other colorless						
Penicillium/Aspergillus types†						
Pithomyces				6	100	80
Rusts				4	100	53
Smuts, Periconia, Myxomycetes				3	100	40
Stachybotrys						
Stemphylium						
Taeniolella						
Torula						
Ulocladium						
Zygomycetes						
Background debris (1-4+)	3+			3+		
Hyphal fragments/m3	< 13			67		
Pollen/m3	< 13			53		
Skin cells (1-4+)	1+			< 1+		
Sample volume (liters)	75			75		
<b>§ TOTAL SPORES/m3</b>			< 13			10,000

**Comments:**

Spore types listed without a count or data entry were not detected during the course of the analysis for the respective sample, indicating a raw count of <1 spore.

† The spores of *Aspergillus* and *Penicillium* (and others such as *Acremonium*, *Paecilomyces*) are small and round with very few distinguishing characteristics. They cannot be differentiated by non-viable sampling methods. Also, some species with very small spores are easily missed, and may be undercounted.

††Background debris indicates the amount of non-biological particulate matter present on the trace (dust in the air) and the resulting visibility for the analyst. It is rated from 1+ (low) to 4+ (high). Counts from areas with 4+ background debris should be regarded as minimal counts and may be higher than reported. It is important to account for samples volumes when evaluating dust levels.

The analytical sensitivity is the spores/m<sup>3</sup> divided by the raw count, expressed in spores/m<sup>3</sup>, per spore and per sample.

For more information regarding analytical sensitivity, please contact QA by calling the laboratory.

‡ A "Version" indicated by -"x" after the Lab ID# with a value greater than 1 indicates a sample with amended data. The revision number is reflected by the value of "x".

§ Total Spores/m<sup>3</sup> has been rounded to two significant figures to reflect analytical precision.

Client: Weaver Consultants Group  
 C/O: Austin Lamp  
 Re: Hanover Countryside E.S.

Date of Sampling: 09-23-2025  
 Date of Receipt: 09-24-2025  
 Date of Report: 09-24-2025

**SPORE TRAP REPORT: NON-VIABLE METHODOLOGY**

Location:	AL092325-17: Building Exterior		
Comments (see below)	A		
Lab ID-Version‡:	21210248-1		
Analysis Date:	09/24/2025		
	raw ct.	% read	spores/m3
Alternaria	13	100	170
Ascospores	20	12.5	2,100
Aureobasidium			
Basidiospores	78	12.5	8,300
Cercospora	6	100	80
Chaetomium			
Cladosporium	16/23	12.5/100	2,000
Curvularia			
Epicoccum	16	100	210
Other brown			
Other colorless			
Penicillium/Aspergillus types†			
Pithomyces	29	100	390
Rusts	5	100	67
Smuts, Periconia, Myxomycetes			
Stachybotrys			
Stemphylium			
Taeniolella	1	100	13
Torula			
Ulocladium			
Zygomycetes			
Background debris (1-4+)	4+		
Hyphal fragments/m3	67		
Pollen/m3	93		
Skin cells (1-4+)	< 1+		
Sample volume (liters)	75		
<b>§ TOTAL SPORES/m3</b>			<b>13,000</b>

**Comments:**A) 23 of the raw count *Cladosporium* spores were present as a single clump.

Spore types listed without a count or data entry were not detected during the course of the analysis for the respective sample, indicating a raw count of <1 spore.

† The spores of *Aspergillus* and *Penicillium* (and others such as *Acremonium*, *Paecilomyces*) are small and round with very few distinguishing characteristics. They cannot be differentiated by non-viable sampling methods. Also, some species with very small spores are easily missed, and may be undercounted.

††Background debris indicates the amount of non-biological particulate matter present on the trace (dust in the air) and the resulting visibility for the analyst. It is rated from 1+ (low) to 4+ (high). Counts from areas with 4+ background debris should be regarded as minimal counts and may be higher than reported. It is important to account for samples volumes when evaluating dust levels.

The analytical sensitivity is the spores/m<sup>3</sup> divided by the raw count, expressed in spores/m<sup>3</sup>, per spore and per sample.

For more information regarding analytical sensitivity, please contact QA by calling the laboratory.

‡ A "Version" indicated by -"x" after the Lab ID# with a value greater than 1 indicates a sample with amended data. The revision number is reflected by the value of "x".

§ Total Spores/m<sup>3</sup> has been rounded to two significant figures to reflect analytical precision.

**Eurofins Built Environment Testing Central, LLC**  
1815 West Diehl Road, Suite 800, Naperville, IL 60563  
(866) 871-1984 www.eurofinsus.com/Built

Client: Weaver Consultants Group  
C/O: Austin Lamp  
Re: Hanover Countryside E.S.

Date of Sampling: 09-23-2025  
Date of Receipt: 09-24-2025  
Date of Report: 09-24-2025

**SPORE TRAP REPORT: NON-VIABLE METHODOLOGY**

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**PROJECT ANALYST AND SIGNATORY REPORT**

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**Project Analyst**



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**Analyst:** Kimberly Bugarin

The test report shall not be reproduced except in full, without written approval of the laboratory. The report must not be used by the client to claim product certification, approval, or endorsement by AIHA LAP, LLC, or any agency of the federal government. The Company reserves the right to dispose of all samples after a period of thirty (30) days, according to all state and federal guidelines, unless otherwise specified.

‡ A "Version" indicated by "-x" after the Lab ID# with a value greater than 1 indicates a sample with amended data. The revision number is reflected by the value of "x".

East: (866) 871-1984  
 Central: (800) 651-4802  
 West: (866) 888-6653

WEATHER		Fog	Rain	Snow	Wind	Clear
LEVEL	None					
	Light					
	Moderate					
	Heavy					

REQUESTED SERVICES

Non-Culturable		Culturable		Other Requests
Spore Trap	Tape, Swab, Bulk	BioCassette™ Swab	Andersen, SAS	



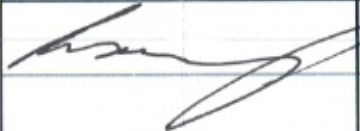
CONTACT INFORMATION		
Company:	Weaver Consultants Group	Address: 1316 Bond Street, Suite 108, Naperville, IL 60563
Contact:	Austin Lamp	Special Instructions: E-mail results to: ih@wcgrp.com & alamp@wcgrp.com
Phone:	224-828-2197	E-mail invoice to: ih@wcgrp.com

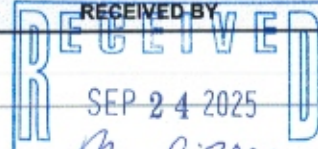
PROJECT INFORMATION			TURN AROUND TIME CODES - (TAT)	
Project ID:	Hanover Countryside E.S.		STD - Standard (Default)	Rushes received after 2pm or on weekends, will be considered received the next business day. Please alert us in advance of weekend analysis needs.
Project Description:			ND - Next Business Day	
Project Zip Code:	Sampling Date/Time:	9/23/25	SD - Same Business Day	
PO Number:	Sampled By:	A. Lamp	WH - Weekend/Holiday/ASAP	

SAMPLE ID	DESCRIPTION	Sample Type (Below)	TAT (Above)	Total Volume/Area (as applicable)	NOTES (Time of day, Temp, RH, etc.)
AL092325-01	Room 24	ST	WH	75L	
-02	Room 22				
-03	Hallway by Commons				
-04	Room 2				
-05	Room 4				
-06	Hallway by Room 4				
-07	Room 4 D				
-08	Room 6				
-09	Room 8				
-10	Room 13				
-11	Hallway by Room 8				
-12	Room 41				
-13	Hallway by Room 41				
-14	Room 14				
-15	Room 15				

Spore Trap Analysis	Other biological particles - supplement	Direct Microscopic Exam (Qualitative)	Quantitative spore count direct exam	Dust Characterization	1-Media Surface Fungi (Genus ID + Asp. spp.)	Culturable Air Fungi (Genus ID + Asp. spp.)	Gram Stain and Counts (Culturable Air and St)	Legionella culture	Total Coliform, E. coli (Presence/Absence)	QuantiTray-Sewage Screen	OTHER: (please specify test)	Asbestos in Air - PCM Airborne Fiber Count (	Asbestos Bulk - PLM	Lead (Pb) - Flame AA	PCR (please specify test)	Allergens (please specify test)
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SAMPLE TYPE CODES			
BC - BioCassette™	CP - Contact Plate	T - Tape	O - Other:
A1S - Andersen	ST - Spore Trap	SW - Swab	
SAS - Surface Air Sampler	B - Bulk	SO - Soil	
NP - Non-potable Water	P - Potable Water	D - Dust	

RELINQUISHED BY	DATE & TIME
	9/23/25 9 PM

RECEIVED BY	DATE & TIME
 SEP 24 2025 By: 	

WEATHER		Fog	Rain	Snow	Wind	Clear
LEVEL	None					
	Light					
	Moderate					
	Heavy					

REQUESTED SERVICES

Non-Culturable		Culturable		Other Requests
Spore Trap	Tape, Swab, Bulk	BioCassette™ Andersen, SAS, Swa		



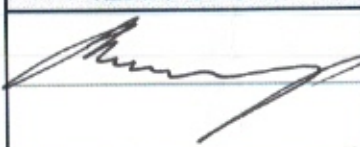
CONTACT INFORMATION		
Company:	Weaver Consultants Group	Address: 1316 Bond Street, Suite 108, Naperville, IL 60563
Contact:	Austin Lamp	Special Instructions: E-mail results to: ih@wcgrp.com & alamp@wcgrp.com
Phone:	224-828-2197	E-mail invoice to: ih@wcgrp.com

PROJECT INFORMATION		TURN AROUND TIME CODES - (TAT)	
Project ID:	Hanover Countryside ES	STD - Standard (Default)	Rushes received after 2pm or on weekends, will be considered received the next business day. Please alert us in advance of weekend analysis needs.
Project Description:		ND - Next Business Day	
Project Zip Code:		SD - Same Business Day	
PO Number:		WH - Weekend/Holiday/ASAP	
Sampling Date/Time:	9/23/25		
Sampled By:	A. Lamp		

SAMPLE ID	DESCRIPTION	Sample Type (Below)	TAT (Above)	Total Volume/Area (as applicable)	NOTES (Time of day, Temp, RH, etc.)
AL092325-16	Building Exterior	ST	WH	75L	
-17	Building Exterior	L	L	L	
-18	Main Office - Wood Stud	T	L	N/A	
-19	Principals office - Fridge Condenser	T	L	N/A	
-20	Principals Office - Wood Stud	T	L	NA	

Spore Trap Analysis	Other biological particles - supplement	Direct Microscopic Exam (Qualitative)	Quantitative spore count direct exam	Dust Characterization	1-Media Surface Fungi (Genus ID + Asp. spp.)	Culturable Air Fungi (Genus ID + Asp. spp.)	Gram Stain and Counts (Culturable Air and Su)	Legionella culture	Total Coliform, E.coli (Presence/Absence)	Quantl Tray-Sewage Screen	OTHER: (please specify test)	Asbestos in Air - PCM Airborne Fiber Count	Asbestos Bulk - PLM	Lead (Pb) - Flame AA	PCR (please specify test)	Allergens (please specify test)
✓																
✓																
		✓														
		✓														
		✓														

SAMPLE TYPE CODES			
BC - BioCassette™	CP - Contact Plate	T - Tape	O - Other:
A1S - Andersen	ST - Spore Trap	SW - Swab	
SAS - Surface Air Sampler	B - Bulk	SO - Soil	
NP - Non-potable Water	P - Potable Water	D - Dust	

RELINQUISHED BY	DATE & TIME
	9/23/25 9 pm

RECEIVED BY	DATE & TIME

Report for:

**Nicholas Liming**  
**Weaver Consultants Group**  
1316 Bond St.  
Suite 108  
Naperville, IL 60563

---

Regarding: Eurofins Built Environment Testing Central, LLC  
Project: Hanover Countryside  
EML ID: 4242712

Approved by:



Business Unit Manager  
Itzel Cuellar

Dates of Analysis:

Spore trap analysis: 09-25-2025

Service SOPs: Spore trap analysis (EB-MY-S-1038)  
AIHA LAP, LLC accredited service, Lab ID #176641

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All samples were received in acceptable condition unless noted in the Report Comments portion in the body of the report. Due to the nature of the analyses performed, field blank correction of results is not applied. The results relate only to the samples as received and tested. Information supplied by the client which can affect the validity of results: sample air volume.

Eurofins Built Environment Testing Central, LLC ("the Company"), a member of the Eurofins Built Environment Testing group of companies, shall have no liability to the client or the client's customer with respect to decisions or recommendations made, actions taken or courses of conduct implemented by either the client or the client's customer as a result of or based upon the Test Results. In no event shall the Company be liable to the client with respect to the Test Results except for the Company's own willful misconduct or gross negligence nor shall the Company be liable for incidental or consequential damages or lost profits or revenues to the fullest extent such liability may be disclaimed by law, even if the Company has been advised of the possibility of such damages, lost profits or lost revenues. In no event shall the Company's liability with respect to the Test Results exceed the amount paid to the Company by the client therefor.

Eurofins Built Environment Testing Central, LLC's LabServe® reporting system includes automated fail-safes to ensure that all AIHA LAP, LLC quality requirements are met and notifications are added to reports when any quality steps remain pending.

Client: Weaver Consultants Group  
 C/O: Nicholas Liming  
 Re: Hanover Countryside

Date of Sampling: 09-24-2025  
 Date of Receipt: 09-25-2025  
 Date of Report: 09-25-2025

**SPORE TRAP REPORT: NON-VIABLE METHODOLOGY**

Location:	NL092425-01: Classroom 23			NL092425-02: Learning Center		
Comments (see below)	None			None		
Lab ID-Version‡:	21221469-1			21221470-1		
Analysis Date:	09/25/2025			09/25/2025		
	raw ct.	% read	spores/m3	raw ct.	% read	spores/m3
Alternaria						
Ascospores						
Basidiospores				2	25	110
Chaetomium						
Cladosporium	2	25	110			
Curvularia						
Epicoccum						
Fusarium						
Myrothecium						
Nigrospora						
Other brown						
Other colorless						
Penicillium/Aspergillus types†						
Pithomyces						
Rusts	5	100	67			
Smuts, Periconia, Myxomycetes	2	100	27			
Stachybotrys						
Stemphylium						
Torula						
Ulocladium						
Zygomycetes						
Background debris (1-4+)	1+			1+		
Hyphal fragments/m3	27			< 13		
Pollen/m3	< 13			< 13		
Skin cells (1-4+)	< 1+			< 1+		
Sample volume (liters)	75			75		
<b>§ TOTAL SPORES/m3</b>			<b>200</b>			<b>110</b>

**Comments:**

Spore types listed without a count or data entry were not detected during the course of the analysis for the respective sample, indicating a raw count of <1 spore.

† The spores of *Aspergillus* and *Penicillium* (and others such as *Acremonium*, *Paecilomyces*) are small and round with very few distinguishing characteristics. They cannot be differentiated by non-viable sampling methods. Also, some species with very small spores are easily missed, and may be undercounted.

††Background debris indicates the amount of non-biological particulate matter present on the trace (dust in the air) and the resulting visibility for the analyst. It is rated from 1+ (low) to 4+ (high). Counts from areas with 4+ background debris should be regarded as minimal counts and may be higher than reported. It is important to account for samples volumes when evaluating dust levels.

The analytical sensitivity is the spores/m<sup>3</sup> divided by the raw count, expressed in spores/m<sup>3</sup>, per spore and per sample.

For more information regarding analytical sensitivity, please contact QA by calling the laboratory.

‡ A "Version" indicated by -"x" after the Lab ID# with a value greater than 1 indicates a sample with amended data. The revision number is reflected by the value of "x".

§ Total Spores/m<sup>3</sup> has been rounded to two significant figures to reflect analytical precision.

Client: Weaver Consultants Group  
 C/O: Nicholas Liming  
 Re: Hanover Countryside

Date of Sampling: 09-24-2025  
 Date of Receipt: 09-25-2025  
 Date of Report: 09-25-2025

**SPORE TRAP REPORT: NON-VIABLE METHODOLOGY**

Location:	NL092425-03: Classroom 25			NL092425-04: Kitchen 26		
Comments (see below)	None			None		
Lab ID-Version‡:	21221471-1			21221472-1		
Analysis Date:	09/25/2025			09/25/2025		
	raw ct.	% read	spores/m3	raw ct.	% read	spores/m3
Alternaria						
Ascospores						
Basidiospores						
Chaetomium						
Cladosporium						
Curvularia						
Epicoccum						
Fusarium						
Myrothecium						
Nigrospora						
Other brown						
Other colorless						
Penicillium/Aspergillus types†						
Pithomyces						
Rusts				1	100	13
Smuts, Periconia, Myxomycetes						
Stachybotrys						
Stemphylium						
Torula						
Ulocladium						
Zygomycetes						
Background debris (1-4+)	1+			1+		
Hyphal fragments/m3	< 13			< 13		
Pollen/m3	< 13			< 13		
Skin cells (1-4+)	< 1+			< 1+		
Sample volume (liters)	75			75		
<b>§ TOTAL SPORES/m3</b>			< 13			13

**Comments:**

Spore types listed without a count or data entry were not detected during the course of the analysis for the respective sample, indicating a raw count of <1 spore.

† The spores of *Aspergillus* and *Penicillium* (and others such as *Acremonium*, *Paecilomyces*) are small and round with very few distinguishing characteristics. They cannot be differentiated by non-viable sampling methods. Also, some species with very small spores are easily missed, and may be undercounted.

††Background debris indicates the amount of non-biological particulate matter present on the trace (dust in the air) and the resulting visibility for the analyst. It is rated from 1+ (low) to 4+ (high). Counts from areas with 4+ background debris should be regarded as minimal counts and may be higher than reported. It is important to account for samples volumes when evaluating dust levels.

The analytical sensitivity is the spores/m<sup>3</sup> divided by the raw count, expressed in spores/m<sup>3</sup>, per spore and per sample.

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§ Total Spores/m<sup>3</sup> has been rounded to two significant figures to reflect analytical precision.

Client: Weaver Consultants Group  
 C/O: Nicholas Liming  
 Re: Hanover Countryside

Date of Sampling: 09-24-2025  
 Date of Receipt: 09-25-2025  
 Date of Report: 09-25-2025

**SPORE TRAP REPORT: NON-VIABLE METHODOLOGY**

Location:	NL092425-05: Classroom 22			NL092425-06: Classroom 20		
Comments (see below)	None			None		
Lab ID-Version‡:	21221473-1			21221474-1		
Analysis Date:	09/25/2025			09/25/2025		
	raw ct.	% read	spores/m3	raw ct.	% read	spores/m3
Alternaria						
Ascospores						
Basidiospores				1	25	53
Chaetomium						
Cladosporium						
Curvularia						
Epicoccum				1	100	13
Fusarium						
Myrothecium						
Nigrospora						
Other brown						
Other colorless						
Penicillium/Aspergillus types†						
Pithomyces						
Rusts				3	100	40
Smuts, Periconia, Myxomycetes						
Stachybotrys						
Stemphylium						
Torula						
Ulocladium						
Zygomycetes						
Background debris (1-4+)	2+			1+		
Hyphal fragments/m3	< 13			< 13		
Pollen/m3	< 13			< 13		
Skin cells (1-4+)	< 1+			< 1+		
Sample volume (liters)	75			75		
<b>§ TOTAL SPORES/m3</b>			< 13			110

**Comments:**

Spore types listed without a count or data entry were not detected during the course of the analysis for the respective sample, indicating a raw count of <1 spore.

† The spores of *Aspergillus* and *Penicillium* (and others such as *Acremonium*, *Paecilomyces*) are small and round with very few distinguishing characteristics. They cannot be differentiated by non-viable sampling methods. Also, some species with very small spores are easily missed, and may be undercounted.

††Background debris indicates the amount of non-biological particulate matter present on the trace (dust in the air) and the resulting visibility for the analyst. It is rated from 1+ (low) to 4+ (high). Counts from areas with 4+ background debris should be regarded as minimal counts and may be higher than reported. It is important to account for samples volumes when evaluating dust levels.

The analytical sensitivity is the spores/m<sup>3</sup> divided by the raw count, expressed in spores/m<sup>3</sup>, per spore and per sample.

For more information regarding analytical sensitivity, please contact QA by calling the laboratory.

‡ A "Version" indicated by -"x" after the Lab ID# with a value greater than 1 indicates a sample with amended data. The revision number is reflected by the value of "x".

§ Total Spores/m<sup>3</sup> has been rounded to two significant figures to reflect analytical precision.

Client: Weaver Consultants Group  
 C/O: Nicholas Liming  
 Re: Hanover Countryside

Date of Sampling: 09-24-2025  
 Date of Receipt: 09-25-2025  
 Date of Report: 09-25-2025

**SPORE TRAP REPORT: NON-VIABLE METHODOLOGY**

Location:	NL092425-07: Commons 27			NL092425-08: Kitchen 29		
Comments (see below)	None			None		
Lab ID-Version‡:	21221475-1			21221476-1		
Analysis Date:	09/25/2025			09/25/2025		
	raw ct.	% read	spores/m3	raw ct.	% read	spores/m3
Alternaria						
Ascospores	1	25	53			
Basidiospores	1	25	53			
Chaetomium						
Cladosporium	2	25	110			
Curvularia						
Epicoccum						
Fusarium						
Myrothecium						
Nigrospora						
Other brown						
Other colorless						
Penicillium/Aspergillus types†						
Pithomyces						
Rusts						
Smuts, Periconia, Myxomycetes						
Stachybotrys						
Stemphylium						
Torula						
Ulocladium						
Zygomycetes						
Background debris (1-4+)	2+			3+		
Hyphal fragments/m3	< 13			27		
Pollen/m3	< 13			< 13		
Skin cells (1-4+)	< 1+			< 1+		
Sample volume (liters)	75			75		
<b>§ TOTAL SPORES/m3</b>			<b>210</b>			<b>&lt; 13</b>

**Comments:**

Spore types listed without a count or data entry were not detected during the course of the analysis for the respective sample, indicating a raw count of <1 spore.

† The spores of *Aspergillus* and *Penicillium* (and others such as *Acremonium*, *Paecilomyces*) are small and round with very few distinguishing characteristics. They cannot be differentiated by non-viable sampling methods. Also, some species with very small spores are easily missed, and may be undercounted.

††Background debris indicates the amount of non-biological particulate matter present on the trace (dust in the air) and the resulting visibility for the analyst. It is rated from 1+ (low) to 4+ (high). Counts from areas with 4+ background debris should be regarded as minimal counts and may be higher than reported. It is important to account for samples volumes when evaluating dust levels.

The analytical sensitivity is the spores/m<sup>3</sup> divided by the raw count, expressed in spores/m<sup>3</sup>, per spore and per sample.

For more information regarding analytical sensitivity, please contact QA by calling the laboratory.

‡ A "Version" indicated by -"x" after the Lab ID# with a value greater than 1 indicates a sample with amended data. The revision number is reflected by the value of "x".

§ Total Spores/m<sup>3</sup> has been rounded to two significant figures to reflect analytical precision.

Client: Weaver Consultants Group  
 C/O: Nicholas Liming  
 Re: Hanover Countryside

Date of Sampling: 09-24-2025  
 Date of Receipt: 09-25-2025  
 Date of Report: 09-25-2025

**SPORE TRAP REPORT: NON-VIABLE METHODOLOGY**

Location:	NL092425-09: Girls Toilet 29			NL092425-10: Boys Toilet 30		
Comments (see below)	None			None		
Lab ID-Version‡:	21221477-1			21221478-1		
Analysis Date:	09/25/2025			09/25/2025		
	raw ct.	% read	spores/m3	raw ct.	% read	spores/m3
Alternaria						
Ascospores						
Basidiospores				1	25	53
Chaetomium						
Cladosporium				1	25	53
Curvularia						
Epicoccum	1	100	13			
Fusarium						
Myrothecium						
Nigrospora						
Other brown	1	100	13			
Other colorless						
Penicillium/Aspergillus types†						
Pithomyces						
Rusts						
Smuts, Periconia, Myxomycetes	1	100	13			
Stachybotrys						
Stemphylium						
Torula						
Ulocladium						
Zygomycetes						
Background debris (1-4+)	3+			3+		
Hyphal fragments/m3	< 13			< 13		
Pollen/m3	< 13			< 13		
Skin cells (1-4+)	< 1+			< 1+		
Sample volume (liters)	75			75		
<b>§ TOTAL SPORES/m3</b>			<b>40</b>			<b>110</b>

**Comments:**

Spore types listed without a count or data entry were not detected during the course of the analysis for the respective sample, indicating a raw count of <1 spore.

† The spores of *Aspergillus* and *Penicillium* (and others such as *Acremonium*, *Paecilomyces*) are small and round with very few distinguishing characteristics. They cannot be differentiated by non-viable sampling methods. Also, some species with very small spores are easily missed, and may be undercounted.

††Background debris indicates the amount of non-biological particulate matter present on the trace (dust in the air) and the resulting visibility for the analyst. It is rated from 1+ (low) to 4+ (high). Counts from areas with 4+ background debris should be regarded as minimal counts and may be higher than reported. It is important to account for samples volumes when evaluating dust levels.

The analytical sensitivity is the spores/m<sup>3</sup> divided by the raw count, expressed in spores/m<sup>3</sup>, per spore and per sample.

For more information regarding analytical sensitivity, please contact QA by calling the laboratory.

‡ A "Version" indicated by -"x" after the Lab ID# with a value greater than 1 indicates a sample with amended data. The revision number is reflected by the value of "x".

§ Total Spores/m<sup>3</sup> has been rounded to two significant figures to reflect analytical precision.

Client: Weaver Consultants Group  
 C/O: Nicholas Liming  
 Re: Hanover Countryside

Date of Sampling: 09-24-2025  
 Date of Receipt: 09-25-2025  
 Date of Report: 09-25-2025

**SPORE TRAP REPORT: NON-VIABLE METHODOLOGY**

Location:	NL092425-11: Office 1			NL092425-12: Principal 1A		
Comments (see below)	None			None		
Lab ID-Version‡:	21221479-1			21221480-1		
Analysis Date:	09/25/2025			09/25/2025		
	raw ct.	% read	spores/m3	raw ct.	% read	spores/m3
Alternaria						
Ascospores				1	25	53
Basidiospores						
Chaetomium						
Cladosporium						
Curvularia						
Epicoccum						
Fusarium						
Myrothecium						
Nigrospora						
Other brown				1	100	13
Other colorless						
Penicillium/Aspergillus types†						
Pithomyces				1	100	13
Rusts						
Smuts, Periconia, Myxomycetes						
Stachybotrys						
Stemphylium						
Torula						
Ulocladium						
Zygomycetes						
Background debris (1-4+)	3+			3+		
Hyphal fragments/m3	< 13			< 13		
Pollen/m3	< 13			< 13		
Skin cells (1-4+)	< 1+			< 1+		
Sample volume (liters)	75			75		
<b>§ TOTAL SPORES/m3</b>			< 13			80

**Comments:**

Spore types listed without a count or data entry were not detected during the course of the analysis for the respective sample, indicating a raw count of <1 spore.

† The spores of *Aspergillus* and *Penicillium* (and others such as *Acremonium*, *Paecilomyces*) are small and round with very few distinguishing characteristics. They cannot be differentiated by non-viable sampling methods. Also, some species with very small spores are easily missed, and may be undercounted.

††Background debris indicates the amount of non-biological particulate matter present on the trace (dust in the air) and the resulting visibility for the analyst. It is rated from 1+ (low) to 4+ (high). Counts from areas with 4+ background debris should be regarded as minimal counts and may be higher than reported. It is important to account for samples volumes when evaluating dust levels.

The analytical sensitivity is the spores/m<sup>3</sup> divided by the raw count, expressed in spores/m<sup>3</sup>, per spore and per sample.

For more information regarding analytical sensitivity, please contact QA by calling the laboratory.

‡ A "Version" indicated by -"x" after the Lab ID# with a value greater than 1 indicates a sample with amended data. The revision number is reflected by the value of "x".

§ Total Spores/m<sup>3</sup> has been rounded to two significant figures to reflect analytical precision.

Client: Weaver Consultants Group  
 C/O: Nicholas Liming  
 Re: Hanover Countryside

Date of Sampling: 09-24-2025  
 Date of Receipt: 09-25-2025  
 Date of Report: 09-25-2025

**SPORE TRAP REPORT: NON-VIABLE METHODOLOGY**

Location:	NL092425-13: Door 1 Vestibule			NL092425-14: Nurses Office		
Comments (see below)	None			None		
Lab ID-Version‡:	21221481-1			21221482-1		
Analysis Date:	09/25/2025			09/25/2025		
	raw ct.	% read	spores/m3	raw ct.	% read	spores/m3
Alternaria						
Ascospores						
Basidiospores						
Chaetomium						
Cladosporium						
Curvularia						
Epicoccum						
Fusarium						
Myrothecium						
Nigrospora						
Other brown						
Other colorless						
Penicillium/Aspergillus types†						
Pithomyces	2	100	27			
Rusts	4	100	53			
Smuts, Periconia, Myxomycetes						
Stachybotrys						
Stemphylium						
Torula						
Ulocladium						
Zygomycetes						
Background debris (1-4+)	3+			3+		
Hyphal fragments/m3	< 13			< 13		
Pollen/m3	< 13			< 13		
Skin cells (1-4+)	< 1+			< 1+		
Sample volume (liters)	75			75		
<b>§ TOTAL SPORES/m3</b>			<b>80</b>			<b>&lt; 13</b>

**Comments:**

Spore types listed without a count or data entry were not detected during the course of the analysis for the respective sample, indicating a raw count of <1 spore.

† The spores of *Aspergillus* and *Penicillium* (and others such as *Acremonium*, *Paecilomyces*) are small and round with very few distinguishing characteristics. They cannot be differentiated by non-viable sampling methods. Also, some species with very small spores are easily missed, and may be undercounted.

††Background debris indicates the amount of non-biological particulate matter present on the trace (dust in the air) and the resulting visibility for the analyst. It is rated from 1+ (low) to 4+ (high). Counts from areas with 4+ background debris should be regarded as minimal counts and may be higher than reported. It is important to account for samples volumes when evaluating dust levels.

The analytical sensitivity is the spores/m<sup>3</sup> divided by the raw count, expressed in spores/m<sup>3</sup>, per spore and per sample.

For more information regarding analytical sensitivity, please contact QA by calling the laboratory.

‡ A "Version" indicated by -"x" after the Lab ID# with a value greater than 1 indicates a sample with amended data. The revision number is reflected by the value of "x".

§ Total Spores/m<sup>3</sup> has been rounded to two significant figures to reflect analytical precision.

Client: Weaver Consultants Group  
 C/O: Nicholas Liming  
 Re: Hanover Countryside

Date of Sampling: 09-24-2025  
 Date of Receipt: 09-25-2025  
 Date of Report: 09-25-2025

**SPORE TRAP REPORT: NON-VIABLE METHODOLOGY**

Location:	NL092425-15: Teachers Workroom 3			NL092425-16: Office 1B		
Comments (see below)	None			None		
Lab ID-Version‡:	21221483-1			21221484-1		
Analysis Date:	09/25/2025			09/25/2025		
	raw ct.	% read	spores/m3	raw ct.	% read	spores/m3
Alternaria						
Ascospores				1	25	53
Basidiospores				1	25	53
Chaetomium						
Cladosporium				1	25	53
Curvularia				1	100	13
Epicoccum				1	100	13
Fusarium						
Myrothecium						
Nigrospora						
Other brown						
Other colorless						
Penicillium/Aspergillus types†						
Pithomyces	2	100	27	1	100	13
Rusts				1	100	13
Smuts, Periconia, Myxomycetes						
Stachybotrys						
Stemphylium						
Torula						
Ulocladium						
Zygomycetes						
Background debris (1-4+)	3+			2+		
Hyphal fragments/m3	< 13			< 13		
Pollen/m3	< 13			< 13		
Skin cells (1-4+)	< 1+			< 1+		
Sample volume (liters)	75			75		
<b>§ TOTAL SPORES/m3</b>			<b>27</b>			<b>210</b>

**Comments:**

Spore types listed without a count or data entry were not detected during the course of the analysis for the respective sample, indicating a raw count of <1 spore.

† The spores of *Aspergillus* and *Penicillium* (and others such as *Acremonium*, *Paecilomyces*) are small and round with very few distinguishing characteristics. They cannot be differentiated by non-viable sampling methods. Also, some species with very small spores are easily missed, and may be undercounted.

††Background debris indicates the amount of non-biological particulate matter present on the trace (dust in the air) and the resulting visibility for the analyst. It is rated from 1+ (low) to 4+ (high). Counts from areas with 4+ background debris should be regarded as minimal counts and may be higher than reported. It is important to account for samples volumes when evaluating dust levels.

The analytical sensitivity is the spores/m<sup>3</sup> divided by the raw count, expressed in spores/m<sup>3</sup>, per spore and per sample.

For more information regarding analytical sensitivity, please contact QA by calling the laboratory.

‡ A "Version" indicated by -"x" after the Lab ID# with a value greater than 1 indicates a sample with amended data. The revision number is reflected by the value of "x".

§ Total Spores/m<sup>3</sup> has been rounded to two significant figures to reflect analytical precision.

Client: Weaver Consultants Group  
 C/O: Nicholas Liming  
 Re: Hanover Countryside

Date of Sampling: 09-24-2025  
 Date of Receipt: 09-25-2025  
 Date of Report: 09-25-2025

**SPORE TRAP REPORT: NON-VIABLE METHODOLOGY**

Location:	NL092425-17: Office 1C			NL092425-18: Conference 3D		
Comments (see below)	None			None		
Lab ID-Version‡:	21221485-1			21221486-1		
Analysis Date:	09/25/2025			09/25/2025		
	raw ct.	% read	spores/m3	raw ct.	% read	spores/m3
Alternaria						
Ascospores				2	25	110
Basidiospores						
Chaetomium						
Cladosporium	1	25	53			
Curvularia						
Epicoccum				1	100	13
Fusarium						
Myrothecium						
Nigrospora						
Other brown						
Other colorless						
Penicillium/Aspergillus types†						
Pithomyces	1	100	13	1	100	13
Rusts	2	100	27			
Smuts, Periconia, Myxomycetes	1	100	13			
Stachybotrys						
Stemphylium						
Torula						
Ulocladium						
Zygomycetes						
Background debris (1-4+)	3+			3+		
Hyphal fragments/m3	< 13			< 13		
Pollen/m3	< 13			< 13		
Skin cells (1-4+)	< 1+			< 1+		
Sample volume (liters)	75			75		
<b>§ TOTAL SPORES/m3</b>			<b>110</b>			<b>130</b>

**Comments:**

Spore types listed without a count or data entry were not detected during the course of the analysis for the respective sample, indicating a raw count of <1 spore.

† The spores of *Aspergillus* and *Penicillium* (and others such as *Acremonium*, *Paecilomyces*) are small and round with very few distinguishing characteristics. They cannot be differentiated by non-viable sampling methods. Also, some species with very small spores are easily missed, and may be undercounted.

††Background debris indicates the amount of non-biological particulate matter present on the trace (dust in the air) and the resulting visibility for the analyst. It is rated from 1+ (low) to 4+ (high). Counts from areas with 4+ background debris should be regarded as minimal counts and may be higher than reported. It is important to account for samples volumes when evaluating dust levels.

The analytical sensitivity is the spores/m<sup>3</sup> divided by the raw count, expressed in spores/m<sup>3</sup>, per spore and per sample.

For more information regarding analytical sensitivity, please contact QA by calling the laboratory.

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§ Total Spores/m<sup>3</sup> has been rounded to two significant figures to reflect analytical precision.

Client: Weaver Consultants Group  
 C/O: Nicholas Liming  
 Re: Hanover Countryside

Date of Sampling: 09-24-2025  
 Date of Receipt: 09-25-2025  
 Date of Report: 09-25-2025

**SPORE TRAP REPORT: NON-VIABLE METHODOLOGY**

Location:	NL092425-19: Girls Toilet 3A			NL092425-20: Boys Toilet 3B		
Comments (see below)	None			None		
Lab ID-Version‡:	21221487-1			21221488-1		
Analysis Date:	09/25/2025			09/25/2025		
	raw ct.	% read	spores/m3	raw ct.	% read	spores/m3
Alternaria						
Ascospores				1	25	53
Basidiospores	1	25	53			
Chaetomium						
Cladosporium						
Curvularia						
Epicoccum						
Fusarium						
Myrothecium						
Nigrospora						
Other brown						
Other colorless						
Penicillium/Aspergillus types†						
Pithomyces	1	100	13			
Rusts						
Smuts, Periconia, Myxomycetes						
Stachybotrys						
Stemphylium						
Torula				1	100	13
Ulocladium						
Zygomycetes						
Background debris (1-4+)	3+			3+		
Hyphal fragments/m3	< 13			13		
Pollen/m3	< 13			< 13		
Skin cells (1-4+)	< 1+			< 1+		
Sample volume (liters)	75			75		
<b>§ TOTAL SPORES/m3</b>			<b>67</b>			<b>67</b>

**Comments:**

Spore types listed without a count or data entry were not detected during the course of the analysis for the respective sample, indicating a raw count of <1 spore.

† The spores of *Aspergillus* and *Penicillium* (and others such as *Acremonium*, *Paecilomyces*) are small and round with very few distinguishing characteristics. They cannot be differentiated by non-viable sampling methods. Also, some species with very small spores are easily missed, and may be undercounted.

††Background debris indicates the amount of non-biological particulate matter present on the trace (dust in the air) and the resulting visibility for the analyst. It is rated from 1+ (low) to 4+ (high). Counts from areas with 4+ background debris should be regarded as minimal counts and may be higher than reported. It is important to account for samples volumes when evaluating dust levels.

The analytical sensitivity is the spores/m<sup>3</sup> divided by the raw count, expressed in spores/m<sup>3</sup>, per spore and per sample.

For more information regarding analytical sensitivity, please contact QA by calling the laboratory.

‡ A "Version" indicated by -"x" after the Lab ID# with a value greater than 1 indicates a sample with amended data. The revision number is reflected by the value of "x".

§ Total Spores/m<sup>3</sup> has been rounded to two significant figures to reflect analytical precision.

Client: Weaver Consultants Group  
 C/O: Nicholas Liming  
 Re: Hanover Countryside

Date of Sampling: 09-24-2025  
 Date of Receipt: 09-25-2025  
 Date of Report: 09-25-2025

**SPORE TRAP REPORT: NON-VIABLE METHODOLOGY**

Location:	NL092425-21: Gymnasium			NL092425-22: Classroom 5		
Comments (see below)	None			None		
Lab ID-Version‡:	21221489-1			21221490-1		
Analysis Date:	09/25/2025			09/25/2025		
	raw ct.	% read	spores/m3	raw ct.	% read	spores/m3
Alternaria						
Ascospores						
Basidiospores	2	25	110			
Chaetomium						
Cladosporium						
Curvularia						
Epicoccum						
Fusarium						
Myrothecium						
Nigrospora						
Other brown						
Other colorless						
Penicillium/Aspergillus types†						
Pithomyces						
Rusts						
Smuts, Periconia, Myxomycetes						
Stachybotrys						
Stemphylium						
Torula						
Ulocladium						
Zygomycetes						
Background debris (1-4+)	< 1+			2+		
Hyphal fragments/m3	< 13			< 13		
Pollen/m3	< 13			< 13		
Skin cells (1-4+)	< 1+			1+		
Sample volume (liters)	75			75		
<b>§ TOTAL SPORES/m3</b>			<b>110</b>			<b>&lt; 13</b>

**Comments:**

Spore types listed without a count or data entry were not detected during the course of the analysis for the respective sample, indicating a raw count of <1 spore.

† The spores of *Aspergillus* and *Penicillium* (and others such as *Acremonium*, *Paecilomyces*) are small and round with very few distinguishing characteristics. They cannot be differentiated by non-viable sampling methods. Also, some species with very small spores are easily missed, and may be undercounted.

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The analytical sensitivity is the spores/m<sup>3</sup> divided by the raw count, expressed in spores/m<sup>3</sup>, per spore and per sample.

For more information regarding analytical sensitivity, please contact QA by calling the laboratory.

‡ A "Version" indicated by -"x" after the Lab ID# with a value greater than 1 indicates a sample with amended data. The revision number is reflected by the value of "x".

§ Total Spores/m<sup>3</sup> has been rounded to two significant figures to reflect analytical precision.

Client: Weaver Consultants Group  
 C/O: Nicholas Liming  
 Re: Hanover Countryside

Date of Sampling: 09-24-2025  
 Date of Receipt: 09-25-2025  
 Date of Report: 09-25-2025

**SPORE TRAP REPORT: NON-VIABLE METHODOLOGY**

Location:	NL092425-23: Classroom 7			NL092425-24: Hallway by Classroom 5/7		
Comments (see below)	None			None		
Lab ID-Version‡:	21221491-1			21221492-1		
Analysis Date:	09/25/2025			09/25/2025		
	raw ct.	% read	spores/m3	raw ct.	% read	spores/m3
Alternaria						
Ascospores						
Basidiospores						
Chaetomium						
Cladosporium	2	25	110			
Curvularia						
Epicoccum						
Fusarium						
Myrothecium						
Nigrospora						
Other brown						
Other colorless						
Penicillium/Aspergillus types†						
Pithomyces						
Rusts						
Smuts, Periconia, Myxomycetes						
Stachybotrys						
Stemphylium						
Torula						
Ulocladium						
Zygomycetes						
Background debris (1-4+)	2+			2+		
Hyphal fragments/m3	< 13			< 13		
Pollen/m3	< 13			< 13		
Skin cells (1-4+)	1+			1+		
Sample volume (liters)	75			75		
<b>§ TOTAL SPORES/m3</b>			<b>110</b>			<b>&lt; 13</b>

**Comments:**

Spore types listed without a count or data entry were not detected during the course of the analysis for the respective sample, indicating a raw count of <1 spore.

† The spores of *Aspergillus* and *Penicillium* (and others such as *Acremonium*, *Paecilomyces*) are small and round with very few distinguishing characteristics. They cannot be differentiated by non-viable sampling methods. Also, some species with very small spores are easily missed, and may be undercounted.

††Background debris indicates the amount of non-biological particulate matter present on the trace (dust in the air) and the resulting visibility for the analyst. It is rated from 1+ (low) to 4+ (high). Counts from areas with 4+ background debris should be regarded as minimal counts and may be higher than reported. It is important to account for samples volumes when evaluating dust levels.

The analytical sensitivity is the spores/m<sup>3</sup> divided by the raw count, expressed in spores/m<sup>3</sup>, per spore and per sample.

For more information regarding analytical sensitivity, please contact QA by calling the laboratory.

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§ Total Spores/m<sup>3</sup> has been rounded to two significant figures to reflect analytical precision.

Client: Weaver Consultants Group  
 C/O: Nicholas Liming  
 Re: Hanover Countryside

Date of Sampling: 09-24-2025  
 Date of Receipt: 09-25-2025  
 Date of Report: 09-25-2025

**SPORE TRAP REPORT: NON-VIABLE METHODOLOGY**

Location:	NL092425-25: Classroom 9			NL092425-26: Classroom 10		
Comments (see below)	None			None		
Lab ID-Version‡:	21221493-1			21221494-1		
Analysis Date:	09/25/2025			09/25/2025		
	raw ct.	% read	spores/m3	raw ct.	% read	spores/m3
Alternaria						
Ascospores	1	25	53			
Basidiospores						
Chaetomium						
Cladosporium						
Curvularia						
Epicoccum						
Fusarium						
Myrothecium						
Nigrospora						
Other brown						
Other colorless						
Penicillium/Aspergillus types†						
Pithomyces						
Rusts				1	100	13
Smuts, Periconia, Myxomycetes						
Stachybotrys						
Stemphylium						
Torula						
Ulocladium						
Zygomycetes						
Background debris (1-4+)	2+			2+		
Hyphal fragments/m3	< 13			< 13		
Pollen/m3	< 13			< 13		
Skin cells (1-4+)	1+			1+		
Sample volume (liters)	75			75		
<b>§ TOTAL SPORES/m3</b>			<b>53</b>			<b>13</b>

**Comments:**

Spore types listed without a count or data entry were not detected during the course of the analysis for the respective sample, indicating a raw count of <1 spore.

† The spores of *Aspergillus* and *Penicillium* (and others such as *Acremonium*, *Paecilomyces*) are small and round with very few distinguishing characteristics. They cannot be differentiated by non-viable sampling methods. Also, some species with very small spores are easily missed, and may be undercounted.

††Background debris indicates the amount of non-biological particulate matter present on the trace (dust in the air) and the resulting visibility for the analyst. It is rated from 1+ (low) to 4+ (high). Counts from areas with 4+ background debris should be regarded as minimal counts and may be higher than reported. It is important to account for samples volumes when evaluating dust levels.

The analytical sensitivity is the spores/m<sup>3</sup> divided by the raw count, expressed in spores/m<sup>3</sup>, per spore and per sample.

For more information regarding analytical sensitivity, please contact QA by calling the laboratory.

‡ A "Version" indicated by -"x" after the Lab ID# with a value greater than 1 indicates a sample with amended data. The revision number is reflected by the value of "x".

§ Total Spores/m<sup>3</sup> has been rounded to two significant figures to reflect analytical precision.

Client: Weaver Consultants Group  
 C/O: Nicholas Liming  
 Re: Hanover Countryside

Date of Sampling: 09-24-2025  
 Date of Receipt: 09-25-2025  
 Date of Report: 09-25-2025

**SPORE TRAP REPORT: NON-VIABLE METHODOLOGY**

Location:	NL092425-27: Classroom 11			NL092425-28: Classroom 12		
Comments (see below)	None			None		
Lab ID-Version‡:	21221495-1			21221496-1		
Analysis Date:	09/25/2025			09/25/2025		
	raw ct.	% read	spores/m3	raw ct.	% read	spores/m3
Alternaria						
Ascospores						
Basidiospores				1	25	53
Chaetomium						
Cladosporium						
Curvularia						
Epicoccum						
Fusarium						
Myrothecium						
Nigrospora						
Other brown						
Other colorless						
Penicillium/Aspergillus types†						
Pithomyces						
Rusts						
Smuts, Periconia, Myxomycetes						
Stachybotrys						
Stemphylium						
Torula						
Ulocladium						
Zygomycetes						
Background debris (1-4+)	2+			2+		
Hyphal fragments/m3	< 13			< 13		
Pollen/m3	< 13			< 13		
Skin cells (1-4+)	1+			1+		
Sample volume (liters)	75			75		
<b>§ TOTAL SPORES/m3</b>			< 13			53

**Comments:**

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Client: Weaver Consultants Group  
 C/O: Nicholas Liming  
 Re: Hanover Countryside

Date of Sampling: 09-24-2025  
 Date of Receipt: 09-25-2025  
 Date of Report: 09-25-2025

**SPORE TRAP REPORT: NON-VIABLE METHODOLOGY**

Location:	NL092425-29: Classroom 17			NL092425-30: Classroom 19		
Comments (see below)	None			None		
Lab ID-Version‡:	21221497-1			21221498-1		
Analysis Date:	09/25/2025			09/25/2025		
	raw ct.	% read	spores/m3	raw ct.	% read	spores/m3
Alternaria						
Ascospores						
Basidiospores						
Chaetomium						
Cladosporium						
Curvularia						
Epicoccum						
Fusarium						
Myrothecium						
Nigrospora						
Other brown						
Other colorless						
Penicillium/Aspergillus types†						
Pithomyces						
Rusts						
Smuts, Periconia, Myxomycetes						
Stachybotrys						
Stemphylium						
Torula						
Ulocladium						
Zygomycetes						
Background debris (1-4+)	2+			2+		
Hyphal fragments/m3	< 13			< 13		
Pollen/m3	< 13			< 13		
Skin cells (1-4+)	1+			1+		
Sample volume (liters)	75			75		
<b>§ TOTAL SPORES/m3</b>			< 13			< 13

**Comments:**

Spore types listed without a count or data entry were not detected during the course of the analysis for the respective sample, indicating a raw count of <1 spore.

† The spores of *Aspergillus* and *Penicillium* (and others such as *Acremonium*, *Paecilomyces*) are small and round with very few distinguishing characteristics. They cannot be differentiated by non-viable sampling methods. Also, some species with very small spores are easily missed, and may be undercounted.

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Client: Weaver Consultants Group  
 C/O: Nicholas Liming  
 Re: Hanover Countryside

Date of Sampling: 09-24-2025  
 Date of Receipt: 09-25-2025  
 Date of Report: 09-25-2025

**SPORE TRAP REPORT: NON-VIABLE METHODOLOGY**

Location:	NL092425-31: Hallway by Classroom 12			NL092425-32: Girls Toilet 8B		
Comments (see below)	None			None		
Lab ID-Version‡:	21221499-1			21221500-1		
Analysis Date:	09/25/2025			09/25/2025		
	raw ct.	% read	spores/m3	raw ct.	% read	spores/m3
Alternaria	1	100	13			
Ascospores						
Basidiospores						
Chaetomium						
Cladosporium						
Curvularia				1	100	13
Epicoccum						
Fusarium						
Myrothecium						
Nigrospora						
Other brown						
Other colorless						
Penicillium/Aspergillus types†						
Pithomyces				1	100	13
Rusts						
Smuts, Periconia, Myxomycetes						
Stachybotrys						
Stemphylium						
Torula						
Ulocladium						
Zygomycetes						
Background debris (1-4+)	2+			2+		
Hyphal fragments/m3	< 13			< 13		
Pollen/m3	< 13			< 13		
Skin cells (1-4+)	1+			1+		
Sample volume (liters)	75			75		
<b>§ TOTAL SPORES/m3</b>			<b>13</b>			<b>27</b>

**Comments:**

Spore types listed without a count or data entry were not detected during the course of the analysis for the respective sample, indicating a raw count of <1 spore.

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Client: Weaver Consultants Group  
 C/O: Nicholas Liming  
 Re: Hanover Countryside

Date of Sampling: 09-24-2025  
 Date of Receipt: 09-25-2025  
 Date of Report: 09-25-2025

**SPORE TRAP REPORT: NON-VIABLE METHODOLOGY**

Location:	NL092425-33: Boys Toilet 8C			NL092425-34: Reading 4B		
Comments (see below)	None			None		
Lab ID-Version‡:	21221501-1			21221502-1		
Analysis Date:	09/25/2025			09/25/2025		
	raw ct.	% read	spores/m3	raw ct.	% read	spores/m3
Alternaria						
Ascospores						
Basidiospores						
Chaetomium						
Cladosporium						
Curvularia						
Epicoccum						
Fusarium						
Myrothecium						
Nigrospora						
Other brown						
Other colorless						
Penicillium/Aspergillus types†						
Pithomyces						
Rusts						
Smuts, Periconia, Myxomycetes						
Stachybotrys						
Stemphylium						
Torula						
Ulocladium						
Zygomycetes						
Background debris (1-4+)	2+			2+		
Hyphal fragments/m3	< 13			< 13		
Pollen/m3	< 13			< 13		
Skin cells (1-4+)	2+			1+		
Sample volume (liters)	75			75		
<b>§ TOTAL SPORES/m3</b>			< 13			< 13

**Comments:**

Spore types listed without a count or data entry were not detected during the course of the analysis for the respective sample, indicating a raw count of <1 spore.

† The spores of *Aspergillus* and *Penicillium* (and others such as *Acremonium*, *Paecilomyces*) are small and round with very few distinguishing characteristics. They cannot be differentiated by non-viable sampling methods. Also, some species with very small spores are easily missed, and may be undercounted.

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Client: Weaver Consultants Group  
 C/O: Nicholas Liming  
 Re: Hanover Countryside

Date of Sampling: 09-24-2025  
 Date of Receipt: 09-25-2025  
 Date of Report: 09-25-2025

**SPORE TRAP REPORT: NON-VIABLE METHODOLOGY**

Location:	NL092425-35: Safe Office 4A			NL092425-36: Gymnasium Office		
Comments (see below)	None			None		
Lab ID-Version‡:	21221503-1			21221504-1		
Analysis Date:	09/25/2025			09/25/2025		
	raw ct.	% read	spores/m3	raw ct.	% read	spores/m3
Alternaria						
Ascospores						
Basidiospores	1	25	53			
Chaetomium						
Cladosporium	5	25	270			
Curvularia						
Epicoccum						
Fusarium						
Myrothecium						
Nigrospora						
Other brown						
Other colorless						
Penicillium/Aspergillus types†						
Pithomyces						
Rusts						
Smuts, Periconia, Myxomycetes						
Stachybotrys						
Stemphylium						
Torula						
Ulocladium						
Zygomycetes						
Background debris (1-4+)	3+			3+		
Hyphal fragments/m3	40			< 13		
Pollen/m3	13			< 13		
Skin cells (1-4+)	2+			2+		
Sample volume (liters)	75			75		
<b>§ TOTAL SPORES/m3</b>			<b>320</b>			<b>&lt; 13</b>

**Comments:**

Spore types listed without a count or data entry were not detected during the course of the analysis for the respective sample, indicating a raw count of <1 spore.

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Client: Weaver Consultants Group  
 C/O: Nicholas Liming  
 Re: Hanover Countryside

Date of Sampling: 09-24-2025  
 Date of Receipt: 09-25-2025  
 Date of Report: 09-25-2025

**SPORE TRAP REPORT: NON-VIABLE METHODOLOGY**

Location:	NL092425-37: Music Room 40			NL092425-38: Storage 42		
Comments (see below)	None			None		
Lab ID-Version‡:	21221505-1			21221506-1		
Analysis Date:	09/25/2025			09/25/2025		
	raw ct.	% read	spores/m3	raw ct.	% read	spores/m3
Alternaria						
Ascospores						
Basidiospores						
Chaetomium						
Cladosporium						
Curvularia						
Epicoccum						
Fusarium						
Myrothecium						
Nigrospora						
Other brown						
Other colorless						
Penicillium/Aspergillus types†						
Pithomyces						
Rusts						
Smuts, Periconia, Myxomycetes						
Stachybotrys						
Stemphylium						
Torula						
Ulocladium						
Zygomycetes						
Background debris (1-4+)	3+			2+		
Hyphal fragments/m3	< 13			< 13		
Pollen/m3	< 13			< 13		
Skin cells (1-4+)	2+			1+		
Sample volume (liters)	75			75		
<b>§ TOTAL SPORES/m3</b>			< 13			< 13

**Comments:**

Spore types listed without a count or data entry were not detected during the course of the analysis for the respective sample, indicating a raw count of <1 spore.

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Client: Weaver Consultants Group  
 C/O: Nicholas Liming  
 Re: Hanover Countryside

Date of Sampling: 09-24-2025  
 Date of Receipt: 09-25-2025  
 Date of Report: 09-25-2025

**SPORE TRAP REPORT: NON-VIABLE METHODOLOGY**

Location:	NL092425-39: Book Room 43			NL092425-40: Itinerant 45		
Comments (see below)	None			None		
Lab ID-Version‡:	21221507-1			21221508-1		
Analysis Date:	09/25/2025			09/25/2025		
	raw ct.	% read	spores/m3	raw ct.	% read	spores/m3
Alternaria						
Ascospores						
Basidiospores				2	25	110
Chaetomium						
Cladosporium				1	25	53
Curvularia						
Epicoccum						
Fusarium						
Myrothecium						
Nigrospora						
Other brown						
Other colorless						
Penicillium/Aspergillus types†						
Pithomyces						
Rusts				1	100	13
Smuts, Periconia, Myxomycetes				1	100	13
Stachybotrys						
Stemphylium						
Torula						
Ulocladium						
Zygomycetes						
Background debris (1-4+)	2+			3+		
Hyphal fragments/m3	< 13			13		
Pollen/m3	< 13			53		
Skin cells (1-4+)	1+			1+		
Sample volume (liters)	75			75		
<b>§ TOTAL SPORES/m3</b>			< 13			190

**Comments:**

Spore types listed without a count or data entry were not detected during the course of the analysis for the respective sample, indicating a raw count of <1 spore.

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Client: Weaver Consultants Group  
 C/O: Nicholas Liming  
 Re: Hanover Countryside

Date of Sampling: 09-24-2025  
 Date of Receipt: 09-25-2025  
 Date of Report: 09-25-2025

**SPORE TRAP REPORT: NON-VIABLE METHODOLOGY**

Location:	NL092425-41: Building Exterior			NL092425-42: Building Exterior		
Comments (see below)	None			None		
Lab ID-Version‡:	21221509-1			21221510-1		
Analysis Date:	09/25/2025			09/25/2025		
	raw ct.	% read	spores/m3	raw ct.	% read	spores/m3
Alternaria	3	100	40	2	100	27
Ascospores	57	25	3,000	46	25	2,500
Basidiospores	138	25	7,400	158	25	8,400
Chaetomium						
Cladosporium	34	25	1,800	48	25	2,600
Curvularia						
Epicoccum	2	100	27	2	100	27
Fusarium						
Myrothecium						
Nigrospora						
Other brown						
Other colorless	4	25	210			
Penicillium/Aspergillus types†						
Pithomyces						
Rusts	1	100	13			
Smuts, Periconia, Myxomycetes				2	100	27
Stachybotrys						
Stemphylium						
Torula						
Ulocladium						
Zygomycetes						
Background debris (1-4+)	3+			3+		
Hyphal fragments/m3	< 13			< 13		
Pollen/m3	< 13			< 13		
Skin cells (1-4+)	< 1+			< 1+		
Sample volume (liters)	75			75		
<b>§ TOTAL SPORES/m3</b>			<b>13,000</b>			<b>14,000</b>

**Comments:**

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Client: Weaver Consultants Group  
C/O: Nicholas Liming  
Re: Hanover Countryside

Date of Sampling: 09-24-2025  
Date of Receipt: 09-25-2025  
Date of Report: 09-25-2025

**SPORE TRAP REPORT: NON-VIABLE METHODOLOGY**

**PROJECT ANALYSTS AND SIGNATORY REPORT**

---

**Project Analysts**



---

**Analyst:** Kimberly Bugarin



---

**Analyst:** Amber Rutter

The test report shall not be reproduced except in full, without written approval of the laboratory. The report must not be used by the client to claim product certification, approval, or endorsement by AIHA LAP, LLC, or any agency of the federal government. The Company reserves the right to dispose of all samples after a period of thirty (30) days, according to all state and federal guidelines, unless otherwise specified.

‡ A "Version" indicated by "-x" after the Lab ID# with a value greater than 1 indicates a sample with amended data. The revision number is reflected by the value of "x".



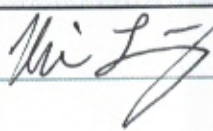
WEATHER	Fog	Rain	Snow	Wind	Clear
LEVEL	None				X
	Light				
	Moderate				
	Heavy				


RE		004242712	
Non-Culturable	Spore Trap	Tape, Swab, Bulk	BioC. Swab, Water, Bulk, Dust, Contact Plate
Spore Trap Analysis	Other biological particles - supplement	Direct Microscopic Exam (Qualitative)	Quantitative spore count direct exam
Dust Characterization	1-Media Surface Fungi (Genus ID + Asp. spp.)	Culturable Air Fungi (Genus ID + Asp. spp.)	Gram Stain and Counts (Culturable Air and Surface Bacteria)
Legionella culture	Total Coliform, E. coli (Presence/Absence)	Quantitative Sewage Screen	OTHER: (please specify test)
Asbestos in Air - PCM Airborne Fiber Count (NIOSH 7400)	Asbestos Bulk - PLM	Lead (Pb) - Flame AA	PCR (please specify test)
Allergens (please specify test)			

CONTACT INFORMATION		
Company:	Weaver Consultants Group	Address: 1316 Bond Street, Suite 108, Naperville, IL 60563
Contact:	Nick Liming	Special Instructions:
Phone:	630-870-9656	E-mail results to: ih@wcgrp.com & nliming@wcgrp.com
		E-mail invoice to: ih@wcgrp.com

PROJECT INFORMATION		TURN AROUND TIME CODES - (TAT)	
Project ID:	Hanover Countryside	STD - Standard (Default)	Rushes received after 2pm or on weekends, will be considered received the next business day. Please alert us in advance of weekend analysis needs.
Project Description:		ND - Next Business Day	
Project Zip Code:		SD - Same Business Day	
PO Number:		WH - Weekend/Holiday/ASAP	
Sampling Date/Time:	9/24/25		
Sampled By:	N. Liming		

SAMPLE ID	DESCRIPTION	Sample Type (Below)	TAT (Above)	Total Volume/Area (as applicable)	NOTES (Time of day, Temp, RH, etc.)
NL092425-01	Classroom 23	ST	WH	75L	
-02	Learning Center				
-03	Classroom 25				
-04	Kitchen 26				
-05	Classroom 22				
-06	Classroom 20				
-07	Commons 27				
-08	Kitchen 29				
-09	Girls Toilet 29				
-10	Boys Toilet 30				
-11	Office 1				
-12	Principal 1A				
-13	Door 1 Vestibule				
-14	Nurses Office				
-15	Teachers Workroom 3				

SAMPLE TYPE CODES				RELINQUISHED BY	DATE & TIME
BC - BioCassette™	CP - Contact Plate	T - Tape	O - Other:		9/24/25
A1S - Andersen	ST - Spore Trap	SW - Swab			
SAS - Surface Air Sampler	B - Bulk	SO - Soil			
NP - Non-potable Water	P - Potable Water	D - Dust			

RECEIVED BY	DATE & TIME
	SEP 25 2025

By submitting this Chain of Custody, you agree to be bound by the terms and conditions set forth at: <https://www.eurofinsus.com/environment-testing/built-environment/resources/sampling-guides-and-forms>



East: (866) 871-1984  
 Central: (800) 651-4802  
 West: (866) 888-6653

WEATHER		Fog	Rain	Snow	Wind	Clear
LEVEL	None					
	Light					
	Moderate					
	Heavy					X

**REQUESTED SERVICES**

Non-Culturable

Spore Trap  Tape, Swab, Bulk  BioCasse Swab, Wt

004242712

**CONTACT INFORMATION**

Company: Weaver Consultants Group Address: 1316 Bond Street, Suite 108, Naperville, IL 60563

Contact: Nick Liming Special Instructions: E-mail results to: ih@wcgrp.com & nliming@wcgrp.com

Phone: 630-870-9656 E-mail invoice to: ih@wcgrp.com

**PROJECT INFORMATION**

Project ID: Harbor Countryside

Project Description: Harbor Countryside

Project Zip Code: N. Liming

PO Number: N. Liming

**TURN AROUND TIME CODES - (TAT)**

STD - Standard (Default) Rushes received after 2pm or on weekends, will be considered received the next business day. Please alert us in advance of weekend analysis needs.

ND - Next Business Day

SD - Same Business Day

WH - Weekend/Holiday/ASAP

SAMPLE ID	DESCRIPTION	Sample Type (Below)	TAT (Above)	Total Volume/Area (as applicable)	NOTES (Time of day, Temp, RH, etc.)
NL092425-31	Hallway by Classroom 12	ST	WH	75L	
-32	Girls Toilet 2B				
-33	Boys Toilet 2C				
-34	Reading 4B				
-35	Safe Office 4A				
-36	Gymnasium Office				
-37	Music Room 40				
-38	Storage 42				
-39	Book Room 43				
-40	Itinerant 45				
-41	Building Exterior				
-42	Building Exterior				

Spore Trap Analysis	Other biological particles - supplement	Direct Microscopic Exam (Qualitative)	Quantitative spore count direct exam	Dust Characterization	1-Media Surface Fungi (Genus ID + Asp. spp.)	Culturable Air Fungi (Genus ID + Asp. spp.)	Gram Stain and Counts (Culturable Air and Surface Balantia)	Legionella culture	Total Coliform, E.coli (Presence/Absence)	QuantTray Sewage Screen	OTHER: (please specify test)	Ashbestos in Air - PCM Aircrome Fiber Count (NIOSH 74)	Asbestos Bulk - PLM	Lead (Pb) - Flame AA	PCR (please specify test)	Allergens (please specify test)
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**SAMPLE TYPE CODES**

BC - BioCassette™	CP - Contact Plate	T - Tape	O - Other:
A1S - Andersen	ST - Spore Trap	SW - Swab	
SAS - Surface Air Sampler	B - Bulk	SO - Soil	
NP - Non-potable Water	P - Potable Water	D - Dust	

**RELINQUISHED BY**

*Nick Liming*

**DATE & TIME**

9/24/25

**RECEIVED BY**

**DATE & TIME**

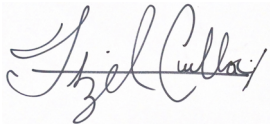
Report for:

**Nicholas Liming**  
**Weaver Consultants Group**  
1316 Bond St.  
Suite 108  
Naperville, IL 60563

---

Regarding: Eurofins Built Environment Testing Central, LLC  
Project: Hanover Countryside; Mobile Unit  
EML ID: 4244488

Approved by:



Business Unit Manager  
Itzel Cuellar

Dates of Analysis:

Spore trap analysis: 09-26-2025

Service SOPs: Spore trap analysis (EB-MY-S-1038)  
AIHA LAP, LLC accredited service, Lab ID #176641

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All samples were received in acceptable condition unless noted in the Report Comments portion in the body of the report. Due to the nature of the analyses performed, field blank correction of results is not applied. The results relate only to the samples as received and tested. Information supplied by the client which can affect the validity of results: sample air volume.

Eurofins Built Environment Testing Central, LLC ("the Company"), a member of the Eurofins Built Environment Testing group of companies, shall have no liability to the client or the client's customer with respect to decisions or recommendations made, actions taken or courses of conduct implemented by either the client or the client's customer as a result of or based upon the Test Results. In no event shall the Company be liable to the client with respect to the Test Results except for the Company's own willful misconduct or gross negligence nor shall the Company be liable for incidental or consequential damages or lost profits or revenues to the fullest extent such liability may be disclaimed by law, even if the Company has been advised of the possibility of such damages, lost profits or lost revenues. In no event shall the Company's liability with respect to the Test Results exceed the amount paid to the Company by the client therefor.

Eurofins Built Environment Testing Central, LLC's LabServe® reporting system includes automated fail-safes to ensure that all AIHA LAP, LLC quality requirements are met and notifications are added to reports when any quality steps remain pending.

Client: Weaver Consultants Group  
 C/O: Nicholas Liming  
 Re: Hanover Countryside; Mobile Unit

Date of Sampling: 09-25-2025  
 Date of Receipt: 09-26-2025  
 Date of Report: 09-26-2025

**SPORE TRAP REPORT: NON-VIABLE METHODOLOGY**

Location:	NL092525-01: Room M-1			NL092525-02: Room M-2		
Comments (see below)	None			None		
Lab ID-Version‡:	21230558-1			21230559-1		
Analysis Date:	09/26/2025			09/26/2025		
	raw ct.	% read	spores/m3	raw ct.	% read	spores/m3
Alternaria						
Ascospores						
Basidiospores				2	25	110
Chaetomium						
Cladosporium						
Curvularia						
Epicoccum						
Fusarium						
Myrothecium						
Nigrospora						
Other colorless						
Penicillium/Aspergillus types†						
Pithomyces						
Rusts						
Smuts, Periconia, Myxomycetes	1	100	13			
Stachybotrys						
Stemphylium						
Torula						
Ulocladium						
Zygomycetes						
Background debris (1-4+)	1+			3+		
Hyphal fragments/m3	< 13			< 13		
Pollen/m3	< 13			< 13		
Skin cells (1-4+)	< 1+			1+		
Sample volume (liters)	75			75		
<b>§ TOTAL SPORES/m3</b>			<b>13</b>			<b>110</b>

**Comments:**

Spore types listed without a count or data entry were not detected during the course of the analysis for the respective sample, indicating a raw count of <1 spore.

† The spores of *Aspergillus* and *Penicillium* (and others such as *Acremonium*, *Paecilomyces*) are small and round with very few distinguishing characteristics. They cannot be differentiated by non-viable sampling methods. Also, some species with very small spores are easily missed, and may be undercounted.

††Background debris indicates the amount of non-biological particulate matter present on the trace (dust in the air) and the resulting visibility for the analyst. It is rated from 1+ (low) to 4+ (high). Counts from areas with 4+ background debris should be regarded as minimal counts and may be higher than reported. It is important to account for samples volumes when evaluating dust levels.

The analytical sensitivity is the spores/m<sup>3</sup> divided by the raw count, expressed in spores/m<sup>3</sup>, per spore and per sample.

For more information regarding analytical sensitivity, please contact QA by calling the laboratory.

‡ A "Version" indicated by -"x" after the Lab ID# with a value greater than 1 indicates a sample with amended data. The revision number is reflected by the value of "x".

§ Total Spores/m<sup>3</sup> has been rounded to two significant figures to reflect analytical precision.

Client: Weaver Consultants Group  
 C/O: Nicholas Liming  
 Re: Hanover Countryside; Mobile Unit

Date of Sampling: 09-25-2025  
 Date of Receipt: 09-26-2025  
 Date of Report: 09-26-2025

**SPORE TRAP REPORT: NON-VIABLE METHODOLOGY**

Location:	NL092525-03: Building Exterior			NL092525-04: Building Exterior		
Comments (see below)	None			None		
Lab ID-Version‡:	21230560-1			21230561-1		
Analysis Date:	09/26/2025			09/26/2025		
	raw ct.	% read	spores/m3	raw ct.	% read	spores/m3
Alternaria	14	100	190	14	100	190
Ascospores	16	25	850	16	12.5	1,700
Basidiospores	129	25	6,900	107	12.5	11,000
Chaetomium						
Cladosporium	98	25	5,200	31	12.5	3,300
Curvularia				2	100	27
Epicoccum	15	100	200	30	100	400
Fusarium						
Myrothecium						
Nigrospora						
Other colorless						
Penicillium/Aspergillus types†						
Pithomyces	2	100	27	1	100	13
Rusts	11	100	150	5	100	67
Smuts, Periconia, Myxomycetes	1	100	13			
Stachybotrys						
Stemphylium						
Torula						
Ulocladium						
Zygomycetes						
Background debris (1-4+)	4+			3+		
Hyphal fragments/m3	40			130		
Pollen/m3	< 13			27		
Skin cells (1-4+)	< 1+			< 1+		
Sample volume (liters)	75			75		
<b>§ TOTAL SPORES/m3</b>			<b>14,000</b>			<b>17,000</b>

**Comments:**

Spore types listed without a count or data entry were not detected during the course of the analysis for the respective sample, indicating a raw count of <1 spore.

† The spores of *Aspergillus* and *Penicillium* (and others such as *Acremonium*, *Paecilomyces*) are small and round with very few distinguishing characteristics. They cannot be differentiated by non-viable sampling methods. Also, some species with very small spores are easily missed, and may be undercounted.

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The analytical sensitivity is the spores/m<sup>3</sup> divided by the raw count, expressed in spores/m<sup>3</sup>, per spore and per sample.

For more information regarding analytical sensitivity, please contact QA by calling the laboratory.

‡ A "Version" indicated by -"x" after the Lab ID# with a value greater than 1 indicates a sample with amended data. The revision number is reflected by the value of "x".

§ Total Spores/m<sup>3</sup> has been rounded to two significant figures to reflect analytical precision.

**Eurofins Built Environment Testing Central, LLC**  
1815 West Diehl Road, Suite 800, Naperville, IL 60563  
(866) 871-1984 www.eurofinsus.com/Built

Client: Weaver Consultants Group  
C/O: Nicholas Liming  
Re: Hanover Countryside; Mobile Unit

Date of Sampling: 09-25-2025  
Date of Receipt: 09-26-2025  
Date of Report: 09-26-2025

**SPORE TRAP REPORT: NON-VIABLE METHODOLOGY**

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**PROJECT ANALYST AND SIGNATORY REPORT**

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**Project Analyst**



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**Analyst:** Kimberly Bugarin

The test report shall not be reproduced except in full, without written approval of the laboratory. The report must not be used by the client to claim product certification, approval, or endorsement by AIHA LAP, LLC, or any agency of the federal government. The Company reserves the right to dispose of all samples after a period of thirty (30) days, according to all state and federal guidelines, unless otherwise specified.

‡ A "Version" indicated by "-x" after the Lab ID# with a value greater than 1 indicates a sample with amended data. The revision number is reflected by the value of "x".

East: (866) 871-1984  
 Central: (800) 651-4802  
 West: (866) 888-6653


WEATHER		Fog	Rain	Snow	Wind	Clear
LEVEL	None					
	Light					
	Moderate					
	Heavy					X

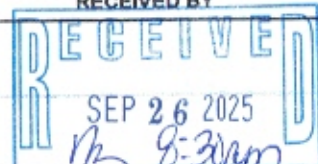
		<b>REQUESTED SERVICES</b>	
		<b>Non-Culturable</b>	<b>Culturable</b>
Spore Trap	Tape, Swab, Bulk	BioCassette™ Swa	Andersen, SAS
		 004244488	
Spore Trap Analysis	Other biological particles - supplement	1-Media Surface Fungi (Genus ID + Asp. spp.)	Asbestos in Air - PCM Airborne Fiber Count (NI)
Direct Microscopic Exam (Qualitative)	Quantitative spore count direct exam	Culturable Air Fungi (Genus ID + Asp. spp.)	Asbestos Bulk - PLM
Dust Characterization		Gram Stain and Counts (Culturable Air and Sur)	Lead (Pb) - Flame AA
		Legionella culture	PCR (please specify test)
		Total Coliform, E. coli (Presence/Absence)	Allergens (please specify test)
		Quant. Tray-Sewage Screen	
		OTHER: (please specify test)	

CONTACT INFORMATION		
Company:	Weaver Consultants Group	Address: 1316 Bond Street, Suite 108, Naperville, IL 60563
Contact:	Nick Liming	Special Instructions:
Phone:	630-870-9656	E-mail results to: ih@wcgrp.com & nliming@wcgrp.com
		E-mail invoice to: ih@wcgrp.com

PROJECT INFORMATION		TURN AROUND TIME CODES - (TAT)	
Project ID:	Hanover Countryside	STD - Standard (Default)	Rushes received after 2pm or on weekends, will be considered received the next business day. Please alert us in advance of weekend analysis needs.
Project Description:	Mobile Unit	ND - Next Business Day	
Project Zip Code:		SD - Same Business Day	
PO Number:		WH - Weekend/Holiday/ASAP	
Sampling Date/Time:	9/25/25		
Sampled By:	N. Liming		

SAMPLE ID	DESCRIPTION	Sample Type (Below)	TAT (Above)	Total Volume/Area (as applicable)	NOTES (Time of day, Temp, RH, etc.)
NL092525-01	Room M-1	ST	WH	75 L	
-02	Room M-2	↓	↓	↓	
-03	Building Exterior	↓	↓	↓	
-04	Building Exterior	↓	↓	↓	

SAMPLE TYPE CODES				RELINQUISHED BY	DATE & TIME
BC - BioCassette™	CP - Contact Plate	T - Tape	O - Other:		9/25/25
A1S - Andersen	ST - Spore Trap	SW - Swab			
SAS - Surface Air Sampler	B - Bulk	SO - Soil			
NP - Non-potable Water	P - Potable Water	D - Dust			

RECEIVED BY	DATE & TIME
 RECEIVED SEP 26 2025 9:30am	



**AIHA Laboratory Accreditation Programs, LLC**  
*acknowledges that*  
**Eurofins Built Environment Testing Central- Chicago, IL.**  
**Eurofins Built Environment Testing Central, LLC.**  
**1815 West Diehl Rd, Suite 800, Naperville, IL 60563-6421**  
**Laboratory ID: LAP-176641**

along with all premises from which key activities are performed, as listed above, has fulfilled the requirements of the AIHA Laboratory Accreditation Programs, LLC (AIHA LAP) accreditation to the ISO/IEC 17025:2017 international standard, General Requirements for the Competence of Testing and Calibration Laboratories in the following:

**LABORATORY ACCREDITATION PROGRAMS**

- |                                     |                            |   |
|-------------------------------------|----------------------------|---|
| <input type="checkbox"/>            | INDUSTRIAL HYGIENE         | Accreditation Expires:                    |
| <input type="checkbox"/>            | ENVIRONMENTAL LEAD         | Accreditation Expires:                    |
| <input checked="" type="checkbox"/> | ENVIRONMENTAL MICROBIOLOGY | Accreditation Expires: September 01, 2027 |
| <input type="checkbox"/>            | FOOD                       | Accreditation Expires:                    |
| <input type="checkbox"/>            | UNIQUE SCOPES              | Accreditation Expires:                    |
| <input type="checkbox"/>            | BE FIELD/MOBILE            | Accreditation Expires:                    |

Specific Field(s) of Testing/Method(s) within each Accreditation Program for which the above named laboratory maintains accreditation is outlined on the attached Scope of Accreditation. Continued accreditation is contingent upon successful on-going compliance with ISO/IEC 17025:2017 and AIHA LAP requirements. This certificate is not valid without the attached Scope of Accreditation. Please review the AIHA LAP website ([www.aihaaccreditedlabs.org](http://www.aihaaccreditedlabs.org)) for the most current Scope.

A handwritten signature in black ink that reads 'Cheryl O. Morton'.

Cheryl O Morton  
Managing Director, AIHA Laboratory Accreditation Programs, LLC