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Resampling Quarterly IAQ Microbial Assessment Report

Conducted At

Hobomock Elementary School
81 Learning Lane
Pembroke, MA 02359

September 2, 2025
Sampling Session #4 (resampling)

Prepared For: Ms. Erin Obey
Superintendent of Schools
Pembroke Public Schools
72 Pilgrim Road
Pembroke, MA 02339

Report Prepared By: 
Paul Matuszko, CIH
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79 Cedar Street
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Report Date: September 5, 2025

PMEC Project #: 25-144



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September 5, 2025

Ms. Erin Obey
Superintendent of Schools
Pembroke Public Schools
72 Pilgrim Road
Pembroke, MA 02339

RE: Resampling Quarterly Microbial (Mold) IAQ Sampling Report #4 for
Hobomock Elementary School
81 Learning Lane, Pembroke, MA
PMEC Project #25-144

Dear Ms. Obey:

Paul Matuszko Environmental Consulting (**PMEC**) is pleased to submit the enclosed report for the follow-up microbial (mold) sampling session conducted at the Hobomock Elementary School, 81 Learning Lane, Pembroke, Massachusetts. PMEC conducted the spore trap air sampling within the school on September 2, 2025. A total of five (5) spore trap air samples were collected for microbial laboratory analysis.

This limited microbial assessment report has been prepared for the exclusive use of The Pembroke Public Schools.

Certification:

PMEC certifies that the results and findings provided herein for the Hobomock Elementary School building have been reviewed for accuracy, content, regulatory compliance and quality of presentation.

Should you have any questions regarding this report, please do not hesitate to contact me at (617) 893-4476. Thank you for providing PMEC with the opportunity to provide our services to the Pembroke Public Schools for this project.

Respectively submitted;

A handwritten signature in black ink that reads "Paul Matuszko".

Paul Matuszko, CIH, CIEC
Project Manager
CIH - Certified Industrial Hygienist (American Board of Industrial Hygiene #9236 CP)
CIEC - Certified Indoor Environmental Consultant (American Council for Accredited Certification)





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Attachment A - Hayes Microbial Laboratory Analytical Results (pages 1-7)



**Resampling
Microbial IAQ Sampling Report
Session #4
For
Hobomock Elementary School
81 Learning Lane
Pembroke, MA
September 2, 2025**

1.0 Introduction

- A. Paul Matuszko Environmental Consulting (PMEC) is pleased to submit this report for the follow-up resampling quarterly microbial air sampling assessment (session #4) conducted at the Hobomock Elementary School, 81 Learning Lane, Pembroke, Massachusetts. PMEC was retained by the Pembroke Public Schools to conduct the general microbial spore trap air sampling within the school building. The microbial (fungal/mold) spore trap air sampling was conducted by PMEC on September 2, 2025. The sampling was performed as a follow-up to confirm the effectiveness of cleaning efforts within classroom 150 and the large teacher's room. The cleaning efforts were recommended after elevated mold spore levels were detected during the August, 2025 quarterly sampling session. A summary of the sampling locations and methods, analysis methods and results are outlined within the report.
- B. PMEC was assisted by the staff members of the Pembroke Public Schools Superintendent office and maintenance/custodial department during the air sampling. Representative member(s) of the Pembroke Teachers Association (PTA) union and Massachusetts State Teachers Association safety professional were present during the sampling to confirm the locations and sampling methods used. The sampling was conducted to determine existing airborne microbial spore levels in two designated locations within the school building.

2.0 Scope of Work

- A. The microbial spore trap sampling was conducted in accordance with the following tasks:
 - 1. General inspection of the representative sampled areas accessed for visible water damaged materials and mold growth in the rooms being sampled. Additionally, the roof top unit #2 was inspected for potential issues and conditions that may affect the building interior.
 - 2. Collect temperature and relative humidity measurements within the locations sampled.
 - 3. Conduct follow-up bioaerosol (microbial) spore trap air sampling for airborne mold (fungal) analysis in designated and representative locations. The sampling scheme focused on rooms that were previously identified to have elevated spore levels above ambient outdoor levels.
 - 4. Prepare a detailed report detailing the findings of the assessment and sampling data.

3.0 Existing Conditions

- A. Prior to resampling, Classroom 150 was recleaned with ceiling HVAC diffusers wiped with a disinfectant. A HEPA air scrubber (small, blue, portable 500 CFM fan unit) was installed and operates in the room to filter airborne particulates. A Carrier (silver) floor model HEPA fan unit was also installed in the room to continually filter airborne particulates. Both HEPA units were operating prior to and during the sampling on September 2, 2025.

- B. The Large teacher’s room was cleaned and disinfected as needed. One, blue 500 CFM HEPA air scrubber was installed and is operating in the room. Additionally, a duel floor model portable air conditioner and dehumidifier operates continuously in the room.
- C. The roof top air handling unit (AHU) #2 was inspected and found to be clean and free of visible mold growth. The AHU #2 has been operating during the school day since the beginning of the academic year. It should be noted that the roof top AHUs were off during and prior to the August, 2025 sampling session.
- D. Rooms were clean and free of visible suspect mold growth and objectionable odors. Ceiling tiles were clean and free of water stains. HVAC vents were clean and free of accumulated dust and debris.

4.0 Air Quality Measurements

- A. A direct reading, thermohygrometer (Fluke Model 971) was used to collect representative temperature and relative humidity (ratio of water vapor in air) measurements.
- B. ASHRAE recommends that occupied indoor temperatures be maintained between approximately ~68°F-74°F in winter months and 72°F-80°F during the summer months. Relative humidity (RH%) is a measurement of water vapor in the air. RH% levels should be kept between 30-60 percent year-round for optimal thermal comfort. High levels of indoor relative humidity (>60%) may be perceived as “sticky” and will promote the microbial growth (fungal/mold). Low levels of RH% below 20% can cause drying of the eyes, skin, and sinus tissue (mucous membranes).
- C. Recommended season levels are:
 - Temperature (Temp.): Recommended Winter Range: ~68-74 °F
 (degrees Fahrenheit °F) Recommended Summer: ~72-80 °F
 - Relative Humidity (RH%) Recommended Range: between 30%-60%; (<50% goal)
- D. The measurement results and room notes on the assessment date are provided in Table 1 below:

Table 1 Baseline Air Quality Measurements and General Room Conditions			
Location	Temperature	Relative Humidity %	Comments
Classroom # 150	77.5 °F	53.7%	Carrier unit and blue air scrubber on, window closed, AHU supply off HVAC vents clean.
Classroom #155	77.7 °F	51.5%	HVAC vents clean, windows closed, no HEPA fans present
Large teachers room	76.0 °F	49.5%	Portable AC unit present (set to 74 °F), HVAC vents clean, windows closed
Outside classroom 150 (south side in shade) – ambient air	~76.6 °F	49.4%	Warm, clear, sunny summer day

- E. Discussion
 1. Temperatures were typical for a school building during the late summer season. Relative humidity levels were also within recommended levels for the warm summer day.

5.0 Mold Sampling and Analysis Methods

- A. PMEC collected “spore trap” air samples for mold spore analysis using air-o-cell® brand sampling cassettes. Four (4) indoor samples and one (1) outdoor (ambient air) comparison sample were collected for analysis. A calibrated, battery operated Zefon IAQ 15 sampling pump was used to draw air onto the sample cassette’s adhesive slide. PMEC collected the samples at 15 liters of air per minute (LPM) for a five (5) minute sample duration for a total sample collection volume of 75 liters of air. Samples are collected in the middle or back side of each room at an approximate height of four feet high. The airborne aerosols (mold, particulates, pollen, etc.) are trapped on the filter media slide for direct microscopic examination.
- B. The samples were collected on September 2, 2025 at approximately 4:00 pm – 4:45 pm. Ambient conditions were reasonable (~76° F) and clear for the late summer season.
- C. The samples were sent via chain of custody by FedEx to Hayes Microbial Laboratory (Hayes), located in Midlothian, Virginia. Hayes Laboratory is accredited by the American Industrial Hygiene Association (AIHA) for mold and bacteria identification and analysis (AIHA EMPAT Laboratory Accreditation ID # 188863).
- D. Results are reported as Total Fungi Counts in spores per cubic meter of air (C/m³). The samples were analyzed for both non-viable and viable fungi by direct analysis optical microscopy. A summary of analysis criteria of spore trap and direct identification analysis is provided in the lab analysis sheets. The sample results are provided as Attachment A to this report.

6.0 Laboratory Analysis Results

- A. The results of the September 2, 2025 spore trap air sampling are presented in Table 2 below.

Table 2 Spore Trap Air Sample Analysis Results September 2, 2025				
Sample #	Sample Location	Total Fungi (C/m ³)	Specific Species & levels of note	Comments
3961 2296 (01)	Classroom 150 (Initial “a” sample)	26 C/m ³	Ascospores – 13 C/m ³ Cladosporium – 13 C/m ³	Very low and acceptable. Recleaning efforts completed.
3961 2304 (02)	Classroom #155 (across hallway comparison sample)	79 C/m ³	Ascospores – 13 C/m ³ Aspergillus/Penicillium - 53 C/m ³ Curvularia – 13 C/m ³	Very low and acceptable. No issues present.
3961 2306 (03)	Outside Ambient Air (Outside classroom 150 south side on grass)	18,400 C/m ³	Ascospores – 2,800 C/m ³ Basidiospores – 1,600 C/m ³ Cladosporium – 4,200 C/m ³ Curvularia – 2,600 C/m ³ Epicoccum – 4,900 C/m ³ Pithomyces – 2,300 C/m ³	Typical high late summer spore level.
3961 2319 (04)	Classroom # 150 (2 nd “b” sample)	80 C/m ³	Ascospores – 13 C/m ³ Curvularia – 27 C/m ³ Epicoccum – 27 C/m ³ Pithomyces – 13 C/m ³	Very low and acceptable. Results confirm cleaning efforts successful.
3961 2292 (05)	Large Teacher’s Room	80 C/m ³	Ascospores – 27 C/m ³ Curvularia – 27 C/m ³ Epicoccum – 13 C/m ³ Pithomyces – 13 C/m ³	Low and acceptable. Cleaning efforts successful

- Notes:**
- Additional information on species types are provided in the Laboratory Analysis results.
 - C/m³ = spore counts per cubic meter of air
 - Aspergillus/penicillium spores are two different species but cannot be differentiated at the level of magnification used during analysis.
 - Samples collected adjacent to teachers desk or back middle of room
 - Results in **Bold** (if present) = species or levels of concern.

7.0 Discussion of Analysis Results

- A. The analysis results for the September 2, 2025 follow up resampling session indicate low and acceptable levels for both classroom #150 and the large teacher's room. The current levels indicate that the operating HVAC system (AHU #2 and smaller teachers room zone AHU) is properly filtering microbial spores and other airborne particulates. Additionally, the cleaning and additional on-going HEPA air scrubbing has successfully lowered spore levels to acceptable levels.
- B. The spore types found within the classroom #150, classroom #155, and large teacher's room are commonly found at low levels and are similar in type to the outdoor ambient sample analysis results. The current airborne spore levels on the sampling date are not representative of an amplified airborne spore condition in the locations sampled.

8.0 Background Mold Information

- A. Currently, there are no standards or regulations to indicate acceptable numerical levels of airborne fungal spores derived from indoor environments. Results are also assessed for specific target species that may induce allergic reactions. Specific species are reviewed for their known potential to cause allergic reactions or as an indicator of potential water damage and moisture issues. In general, indoor mold levels should be equivalent to or lower than outdoor levels or non-complaint areas with similar types and percentages of mold species. Please note that airborne mold spores are present in most indoor environments at low levels.
- B. There are no regulations for acceptable levels of mold in the indoor environment; therefore, exact numerical limits are not supported at this time. Specific fungal species of concern are recommended to have much lower levels in the indoor environment. Existing conditions and symptoms are unique to each season, building and occupant. Airborne mold spore levels may vary greatly by location, time of day and weather conditions. However, health impacts cannot be predicted based on this information; individuals experience varying levels of allergic and non-allergic response to mold. Controlling moisture, in each form, is critical to the prevention of indoor mold growth issues.

9.0 Summary Conclusions

- A. The September 2, 2025 resampling session indicate the laboratory analysis results were low and acceptable in the rooms sampled. **The current levels in these five locations indicate a Level 1 condition (normal indoor fungal ecology) as defined by the Institute of Inspection, Cleaning, and Restoration Certification (IICRC).**
- B. No elevated spore levels were identified in classroom 150 and the large teacher's room.
- C. Temperature and relative humidity levels were within recommended levels during the sampling period. These levels are typical for the summer season with warm and dry conditions.

10.0 Limitations

- A. The assessment provided herein is based on the professional judgment of P MEC using approved industry standards and guidelines. Not all areas of the school building were accessed for inspection

during the representative general spore trap air sampling. Assessment findings are based on the investigator's careful consideration of field observations and interpretation of analysis results in accordance with industry standards, including, but not limited to, IICRC S520 guidelines for Condition 1 – normal fungal ecology, 2008 AIHA (Green Book) publication "Recognition, Evaluation, and Control of Indoor Mold", and the ACGIH 1999 book "Bioaerosols – Assessment and Control".

- B. The analysis results are only representative of the conditions of the date and time of sample collection and are considered a "snapshot in time". PMEC's results and discussion listed herein represent the conditions present at the time of inspection and sampling.

ATTACHMENTS

Attachment A – Hayes Microbial – September 2, 2025 Sample Analysis Results (pages 1-7)

Attachment A

Laboratory Analysis Results

**Hayes Microbial Laboratory
September 2, 2025 Resampling Sampling Session
(pages 1-7)**



#25042621

Analysis Report prepared for

Paul Matuszko Environmental Consulting

79 Cedar Street
Walpole, MA 02081

Phone: (617) 893-4476

25-144
Hobomock Elementary School
81 Learning Ln.
Pembroke, MA 02359

Collected: **September 2, 2025**
Received: **September 4, 2025**
Reported: **September 5, 2025**



EPA Laboratory ID: VA01419



Lab ID: #188863



DPH License: #PH-0198

We would like to thank you for trusting Hayes Microbial for your analytical needs!
We received 5 samples by FedEx in good condition for this project on September 4th, 2025.

The results in this analysis pertain only to this job, collected on the stated date, and should not be used in the interpretation of any other job. Information supplied by the customer can affect the validity of results. These results apply only to the samples as received. This report may not be duplicated, except in full, without the written consent of Hayes Microbial Consulting, LLC.

All information provided to Hayes Microbial is confidential information relating to our customers and their clients. We will not disclose, copy, or distribute any information verbally or written, except to those designated by the customer(s). We take confidentiality very seriously. No changes to the distribution list will be made without the express consent of the customer.

This laboratory bears no responsibility for sample collection activities, analytical method limitations, or your use of the test results. Interpretation and use of test results are your responsibility. Any reference to health effects or interpretation of mold levels is strictly the opinion of Hayes Microbial. In no event, shall Hayes Microbial or any of its employees be liable for lost profits or any special, incidental or consequential damages arising out of the use of these test results.

A handwritten signature in black ink that reads 'Stephen N. Hayes'.

Steve Hayes, BSMT (ASCP)
Laboratory Director
Hayes Microbial Consulting, LLC.

Sample Number*	1 3961 2296			2 3961 2304			3 3961 2306			4 3961 2319		
Sample Name*	Classroom 150A			Classroom 155			Outside Ambient Air			Classroom 150B		
Sample Volume*	75 L											
Reporting Limit	13 spores/m ³											
Background	2			2			2			2		
Fragments	ND			ND			ND			ND		
Organism	Raw Count	Count / m ³	% of Total	Raw Count	Count / m ³	% of Total	Raw Count	Count / m ³	% of Total	Raw Count	Count / m ³	% of Total
Alternaria												
Ascospores	1	13	50.0%	1	13	16.7%	211	2800	15.2%	1	13	16.7%
Aspergillus Penicillium				4	53	66.7%						
Basidiospores							122	1600	8.8%			
Bipolaris Drechslera												
Chaetomium												
Cladosporium	1	13	50.0%				314	4200	22.6%			
Curvularia				1	13	16.7%	198	2600	14.3%	2	27	33.3%
Epicoccum							371	4900	26.7%	2	27	33.3%
Fusarium												
Memnoniella												
Myxomycetes												
Pithomyces							173	2300	12.5%	1	13	16.7%
Stachybotrys												
Stemphylium												
Torula												
Ulocladium												
Total	2	26	100%	6	79	100%	1389	18400	100%	6	80	100%

Water Damage Indicator	Common Allergen	Slightly Higher than Baseline	Significantly Higher than Baseline	Ratio Abnormality
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* indicates data provided by the customer

Collected: **Sep 2, 2025**

Received: **Sep 4, 2025**

Reported: **Sep 5, 2025**



Project Analyst:
 Ramesh Poluri, PhD

P. Ramesh

Date:
09 - 05 - 2025

Reviewed By:
 David McDonald, PHR

David McDonald

Date:
09 - 05 - 2025

Sample Number*	5	3961 2292			
Sample Name*	Large Teacher's Room				
Sample Volume*	75 L				
Reporting Limit	13 spores/m ³				
Background	2				
Fragments	ND				
Organism	Raw Count	Count / m³	% of Total		
Alternaria					
Ascospores	2	27	33.3%		
Aspergillus Penicillium					
Basidiospores					
Bipolaris Drechslera					
Chaetomium					
Cladosporium					
Curvularia	2	27	33.3%		
Epicoccum	1	13	16.7%		
Fusarium					
Memnoniella					
Myxomycetes					
Pithomyces	1	13	16.7%		
Stachybotrys					
Stemphylium					
Torula					
Ulocladium					
Total	6	80	100%		

Water Damage Indicator	Common Allergen	Slightly Higher than Baseline	Significantly Higher than Baseline	Ratio Abnormality
------------------------	-----------------	-------------------------------	------------------------------------	-------------------

* indicates data provided by the customer



Collected: **Sep 2, 2025**

Received: **Sep 4, 2025**

Reported: **Sep 5, 2025**

Project Analyst:
 Ramesh Poluri, PhD *P. Ramesh*

Date:
09 - 05 - 2025

Reviewed By:
 David McDonald, PHR *David McDonald*

Date:
09 - 05 - 2025

Spore Trap Information

Reporting Limit	The Reporting Limit is the lowest number of spores that can be detected based on the total volume of the sample collected and the percentage of the slide that is counted. At Hayes Microbial, 100% of the slide is read so the LOD is based solely on the total volume. Raw spore counts that exceed 500 spores will be estimated.					
Blanks	Results have not been corrected for field or laboratory blanks.					
Background	<p>The Background is the amount of debris that is present in the sample. This debris consists of skin cells, dirt, dust, pollen, drywall dust and other organic and non-organic matter. As the background density increases, the likelihood of spores, especially small spores such as those of <i>Aspergillus</i> and <i>Penicillium</i> may be obscured. The background is rated on a scale of 1 to 5 and each level is determined as follows:</p> <p>NBD: No background detected due to possible pump or cassette malfunction. Recollect sample. (Field Blanks will display NBD)</p> <p>1 : <5% of field occluded. No spores will be uncountable.</p> <p>2 : 5-25% of field occluded.</p> <p>3 : 25-75% of field occluded.</p> <p>4 : 75-90% of field occluded.</p> <p>5 : >90% of field occluded. Suggested recollection of sample.</p>					
Fragments	Fragments are small pieces of fungal mycelium or spores. They are not identifiable as to type and when present in very large numbers, may indicate the presence of mold amplification.					
Control Comparisons	There are no national standards for the numbers of fungal spores that may be present in the indoor environment. As a general rule and guideline that is widely accepted in the indoor air quality field, the numbers and types of spores that are present in the indoor environment should not exceed those that are present outdoors at any given time. There will always be some mold spores present in "normal" indoor environments. The purpose of sampling and counting spores is to help determine whether an abnormal condition exists within the indoor environment and if it does, to help pinpoint the area of contamination. Spore counts should not be used as the sole determining factor of mold contamination. There are many factors that can cause anomalies in the comparison of indoor and outdoor samples due to the dynamic nature of both of those environments.					
<table border="1"> <tr><td>Water Damage Indicator</td></tr> <tr><td>Common Allergen</td></tr> <tr><td>Slightly Higher than Baseline</td></tr> <tr><td>Significantly Higher than Baseline</td></tr> <tr><td>Ratio Abnormality</td></tr> </table>	Water Damage Indicator	Common Allergen	Slightly Higher than Baseline	Significantly Higher than Baseline	Ratio Abnormality	<p>Blue: These molds are commonly seen in conditions of prolonged water intrusion and usually indicate a problem.</p> <p>Green: Although all molds are potential allergens, these are the most common allergens that may be found indoors.</p> <p>Orange: The spore count is slightly higher than the outside count and may or may not indicate a source of contamination.</p> <p>Red: The spore count is significantly higher than the baseline count and probably indicates a source of contamination.</p> <p>Violet: The types of spores found indoors should be similar to the ones that were identified in the baseline sample. Significant increases (more than 25%) in the ratio of a particular spore type may indicate the presence of abnormal levels of mold, even if the total number of spores of that type is lower in the indoor environment than it was outdoors.</p>
Water Damage Indicator						
Common Allergen						
Slightly Higher than Baseline						
Significantly Higher than Baseline						
Ratio Abnormality						
Color Coding	Fungi that are present in indoor samples at levels lower than 200 per cubic meter are not color coded on the report, unless they are one of the water damage indicators.					
Significant Figures	Raw counts and column totals may reflect more than 2 significant figures, but results should only be considered significant to 2 figures.					

Ascospores	Habitat:	A large group consisting of more than 3000 species of fungi. Common plant pathogens and outdoor numbers become very high following rain. Most of the genera are indistinguishable by spore trap analysis and are combined on the report.
	Health Effects:	Health affects are poorly studied, but many are likely to be allergenic.

Aspergillus Penicillium	Habitat:	The most common fungi isolated from the environment. Very common in soil and on decaying plant material. Are able to grow well indoors on a wide variety of substrates.
	Health Effects:	This group contains common allergens and many can cause hypersensitivity pneumonitis. They may cause extrinsic asthma, and many are opportunistic pathogens. Many species produce mycotoxins which may be associated with disease in humans and other animals. Toxin production is dependent on the species, the food source, competition with other organisms, and other environmental conditions.

Basidiospores	Habitat:	A common group of Fungi that includes the mushrooms and bracket fungi. They are saprophytes and plant pathogens. In wet conditions they can cause structural damage to buildings.
	Health Effects:	Common allergens and are also associated with hypersensitivity pneumonitis.

Cladosporium	Habitat:	One of the most common genera worldwide. Found in soil and plant debris and on the leaf surfaces of living plants. The outdoor numbers are lower in the winter and often relatively high in the summer, especially in high humidity. The outdoor numbers often spike in the late afternoon and evening. Indoors, it can be found growing on textiles, wood, sheetrock, moist window sills and in HVAC supply ducts.
	Health Effects:	A common allergen, producing more than 10 allergenic antigens and a common cause of hypersensitivity pneumonitis.

Curvularia	Habitat:	They exist in soil and plant debris, and are plant pathogens.
	Health Effects:	They are allergenic and a common cause of allergic fungal sinusitis. An occasional cause of human infection, including keratitis, sinusitis, onychomycosis, mycetoma, pneumonia, endocarditis and disseminated infection, primarily in the immunocompromised.

Epicoccum	Habitat:	It is found in soil and plant litter and is a plant pathogen. It can grow indoors on a variety of substrates, including paper and textiles and is commonly found on wet drywall.
	Health Effects:	It is a common allergen. No cases of infection have been reported in humans.

Pithomyces

Habitat: Common fungus isolated from soil, decaying plant material. Rarely found indoors.
Health Effects: Allergenic properties are poorly studied. No cases of infection in humans.



Paul Matuszko Environmental Consulting
 79 Cedar Street
 Walpole, MA 02081

N

SHIP: FEDEX - ENV 50
 DATE: 09-04-2025



25042621

8175 9179 7924



Job Number: 25-144	Job Name: Hobomock Elementary School 81 Learning Lane Pembroke, MA 02359	Mobile: 617 893-4476	Email: pmatuszko@pmecsolutions.co
Collector: Paul Matuszko		Note: <i>Background Resampling Session</i>	
Date Collected: 9/2/25 ✓			

Analysis Type	Analysis Description	Turnaround	Accepted Media Types
Spore Trap	S Identification & Enumeration of Fungal Spores	24 Hour	Air Cassettes, Impact Slides
	S+ Spore Trap Analysis with Dander, Fiber, and Pollen counts	24 Hour	Air Cassettes, Impact Slides
Direct ID	D ID & Semi-Quantative Enumeration of spores and mycelium	24 Hour	Bio-Tape, Tape, Swab, Bulk, Agar Plate
	D+ Direct Analysis with Fully Quantitative spore count	24 Hour	Bio-Tape, Tape, Swab, Bulk, Agar Plate
Culture	C1 Identification & Enumeration of Mold only	7 Day	Air Plate, Agar Plate, Swab, Bulk
	C2 Identification & Enumeration of Bacteria only	4 Day	Air Plate, Agar Plate, Swab, Bulk
	C3 Identification & Enumeration of Mold and Bacteria	7 Day	Air Plate, Agar Plate, Swab, Bulk
	C5 Coliform Screen for Sewage Bacteria	2 Day	Agar Plate, Swab, Bulk
Particle	TPA Total Particulate Analysis, ID & Count (Does Not Include Mold)	24 Hour	Air Cassettes, Impact Slides, Bio-Tape

#	Number	Sample	Analysis	Volume	Notes
1	39612296	classroom 150a	S	75L	5min @ 15LPM
2	39612304	classroom 155	S		
3	39612306	outside Ambient Air	S		on grass outside Room 155
4	39612319	classroom 150b	S		
5	39612292	Large Teacher's Room	S		
6					
7					
8					
9					
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15					
16					

Released by: Paul Matuszko <i>PM</i>	Date: 9/3/25 <i>PM</i>	Received By: <i>PM</i>	Date: 9/4
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