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**Quarterly
Microbial IAQ
Assessment Report
#2**

Conducted At

**Hobomock Elementary School
81 Learning Lanes
Pembroke, MA 02359**

February, 2025

Prepared For:

Ms. Erin Obey
Superintendent of Schools
Pembroke Public Schools
72 Pilgrim Road
Pembroke, MA 02339

Report Prepared By:

Paul Matuszko, CIH
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Report Date:

February 24, 2025

PMEC Project #:

25-114



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February 24, 2025

Ms. Erin Obey
Superintendent of Schools
Pembroke Public Schools
72 Pilgrim Road
Pembroke, MA 02339

RE: Quarterly Microbial (Mold) IAQ Sampling Report #2 for
Hobomock Elementary School
81 Learning Lane, Pembroke, MA
PMEC Project #24-163.1

Dear Ms. Obey:

Paul Matuszko Environmental Consulting (**PMEC**) is pleased to submit the enclosed report for the quarterly microbial (mold) indoor air quality (IAQ) sampling session conducted at the Hobomock Elementary School, 81 Learning Lane, Pembroke, Massachusetts. PMEC conducted the sampling within the school on February 13, 2025. Nine (9) total air samples were collected for microbial laboratory analysis.

This limited microbial assessment report has been prepared for the exclusive use of The Pembroke Public Schools.

Certification:

PMEC certifies that the results and findings provided herein for the Hobomock Elementary School building have been reviewed for accuracy, content, regulatory compliance and quality of presentation.

Should you have any questions regarding this report, please do not hesitate to contact me at (617) 893-4476. Thank you for providing PMEC with the opportunity to provide our services to the Pembroke Public Schools for this project.

Respectfully submitted;

Paul Matuszko, CIH, CIEC
Project Manager

CIH - Certified Industrial Hygienist (American Board of Industrial Hygiene #9236 CP)

CIEC - Certified Indoor Environmental Consultant (American Council for Accredited Certification)





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**Quarterly Microbial IAQ
Sampling Report
Session #2
For
Hobomock Elementary School
81 Learning Lanes
Pembroke, MA

February, 2025**

1.0 Introduction

- A. Paul Matuszko Environmental Consulting (**PMEC**) is pleased to submit this report for the quarterly microbial sampling assessment (#2) conducted at the Hobomock Elementary School, 81 Learning Lane, Pembroke, Massachusetts. PMEC was retained by the Pembroke Public Schools to conduct the general air sampling and a general inspection for water and microbial growth within the school building. The microbial spore trap air sampling was conducted by PMEC on February 13, 2025. The quarterly sampling session was performed to assess existing spore levels in compliance with the on-going school system indoor air quality (IAQ) program. A summary of the sampling locations and methods, analysis methods and results are outlined within the report.
- B. PMEC was assisted by Pembroke Public Schools Superintendent and maintenance/custodial personnel during the air sampling. Member(s) of the Pembroke Teachers Association (PTA) Union were present during the sampling to confirm the locations and sampling methods used. The sampling was conducted to determine existing airborne microbial spore levels in representative areas of each section or wing of the school building.
- C. During sampling, PMEC conducted a general visual inspection of representative room surfaces. Additionally, measurements baseline indoor temperature and humidity levels were collected. PMEC noted any areas or locations with atypical conditions. The following report summarizes the findings of the assessment, analysis results, and general recommendations.

2.0 Scope of Work

- A. The indoor air quality (IAQ) assessment was conducted in accordance with the following tasks:
 1. General inspection of the representative areas for visible water damaged materials and mold growth in the rooms being sampled.
 2. Conduct representative bioaerosol (microbial) spore trap air sampling for airborne mold (fungal) analysis in designated and representative locations. Eight (8) interior samples and one (1) comparison sample were collected during the February 13, 2025 after school hours site visit (~3:30-5:00 pm).
 - Sampling locations were selected at the request of the PTA for the Music room, library, gym atrium, and SPED classroom #275.
 3. Prepare a detailed report detailing the findings of the assessment and sampling data.

3.0 Air Quality Measurements

- A. A direct reading, thermohygrometer (Fluke Model 971) was used to collect representative temperature and relative humidity (ratio of water vapor in air) measurements.

B. ASHRAE recommends that occupied indoor temperatures be maintained between approximately ~68°F-74°F in winter months and 72°F-80°F during the summer months. Relative humidity (RH%) is a measurement of water vapor in the air. RH% levels should be kept between 30-60 percent year-round for optimal thermal comfort. High levels of indoor relative humidity (>60%) may be perceived as “sticky” and will promote the microbial growth (fungal/mold). Low levels of RH% below 20% can cause drying of the eyes, skin, and sinus tissue (mucous membranes).

C. Recommended season levels are:

- Temperature (Temp.): Recommended Winter Range: ~68-74 °F
(degrees Fahrenheit °F) Recommended Summer: ~72-80 °F
- Relative Humidity (RH%) Recommended Range: between 30%-60%;
(<50% goal)

D. The measurement results on the assessment date are provided in Table 1 below:

Table 1 Baseline Air Quality Measurements			
Location	Temperature	Relative Humidity %	Comments
Classroom # 105	68.8 °F	37.3%	Stained ceiling tile back left corner
Classroom #160	70.5 °F	37.1%	-
Hallway at Room #130 (outside girl's room)	70.8 °F	33.3%	No odor present
Classroom #220	72.5 °F	30.4%	-
SPED Classroom #275	76.2 °F	28.7%	Warmer interior core room
Music Room	73.5 °F	32.4%	No odor present
Gym Atrium (entrance lobby)	69.8 °F	35.9%	No odor present
Inside vestibule of Gym entrance	40.6 °F	54.7%	Comparison sample. Snow cover on ground
Library – right side	73.0 °F	30.6 %	No odor present

E. Discussion

1. Temperature were typical for a school building during the mid-winter season. Relative humidity levels were within recommended levels for the winter season. RH% will be lower in warmer rooms such as SPED #275. However, the temperatures are typical of occupied buildings during a dry winter day.

4.0 Mold Sampling and Analysis Methods

- A. PMEC collected “spore trap” air samples for mold spore analysis using air-o-cell® brand sampling cassettes. Eight (8) indoor samples and one (1) outdoor (vestibule) comparison sample were collected for analysis. A calibrated, battery operated Zefon IAQ 15 sampling pump was used to draw air onto the sample cassette’s adhesive slide. PMEC collected the samples at 15 liters of air per minute (LPM) for a five (5) minute sample duration for a total sample collection volume of 75 liters of air. Samples are collected in the middle or back side of each room at an approximate height of four feet high. The airborne aerosols (mold, particulates, pollen, etc.) are trapped on the filter media slide for direct microscopic examination.

- B. The samples were collected on February 13, 2025 at approximately 3:30 pm – 5:00 pm. Ambient conditions were cool (~39-40° F) and clear with snow cover on exterior surfaces. Indoor conditions were typical of the season.
- C. The samples were sent via chain of custody by Fed Ex to Hayes Microbial Laboratory (Hayes), located in Midlothian, Virginia. Hayes Laboratory is accredited by the American Industrial Hygiene Association (AIHA) for mold and bacteria identification and analysis (AIHA EMPAT Laboratory Accreditation ID # 188863).
- D. Results are reported as Total Fungi Counts in spores per cubic meter of air (**C/m³**). The samples were analyzed for both non-viable and viable fungi by direct analysis optical microscopy. A summary of analysis criteria of spore trap and direct identification analysis is provided in the lab analysis sheets. The sample results are provided as Attachment B to this report.

5.0 Laboratory Analysis Results

- A. The results of the February 13, 2025 spore trap air sampling are presented in Table 2 below.

Table 2 Spore Trap Air Sample Analysis Results February 21, 2025 Quarterly IAQ Sampling Session #2				
Sample #	Sample Location	Total Fungi (C/m³)	Specific Species & levels of note	Comments
3961 2261 (01)	Classroom 105	13 C/m ³	Ascospores – 13 C/m ³	Very low and acceptable
3961 2263 (02)	Classroom #160	143 C/m ³	Aspergillus/Penicillium -130 C/m ³ Bipolaris / Drechslera – 13 C/m ³	Low and acceptable
3961 2275 (03)	Hallway near #130	13 C/m ³	Ascospores – 13 C/m ³	Very low and acceptable
3961 2273 (04)	Classroom #220	27 C/m ³	Ascospores – 27 C/m ³	Very low and acceptable
3961 2259 (05)	SPED Classroom #275	13 C/m ³	Ascospores – 13 C/m ³	Very low and acceptable
3911 3613 (06)	Music Room	26 C/m ³	Ascospores – 13 C/m ³ Cladosporium – 13 C/m ³	Very low and acceptable
39611 3649 (07)	Gym Atrium (entrance lobby)	26 C/m ³	Ascospores – 13 C/m ³ Basidiospores – 13 C/m ³	Very low and acceptable
3961 2262 (08)	Vestibule entrance at Gym entrance (comparison sample)	27 C/m ³	Ascospores – 27 C/m ³	Typical winter level with snow cover on ground
3961 2283 (09)	Library – At right side tables	13 C/m ³	Ascospores – 13 C/m ³	Very low and acceptable

Notes:

- Additional information on species types are provided in the Laboratory Analysis results.
- C/m³ = spore counts per cubic meter of air
- Samples collected in middle or back middle of room unless noted
- Results in **Bold** (if present) = species or levels of concern.

6.0 Discussion of Analysis Results

- A. The analysis results for the February 13, 2025 sampling session indicate very low and acceptable airborne spore levels. The species identified in these rooms are typically found indoors at low levels and are not a cause for concern.
- B. The indoor sample species identified are similar to the species present on the vestibule comparison sample analysis. The current airborne spore levels on the sampling date are not representative of an amplified airborne spore condition in the locations sampled.
- C. Low levels of aspergillus/penicillium (Asp/Pen) species were identified in Classroom 160. The presence of Asp/Pen spores were not identified in other locations. However, the levels identified are currently low at the time of sampling. Their presence is noted but are not considered to be elevated or a cause for concern at this time.
- D. The results are typical for the mid-winter season with snow cover on exterior surfaces which results in limited to no exterior spore levels.

7.0 Background Mold Information

- A. Currently, there are no standards or regulations to indicate acceptable numerical levels of airborne fungal spores derived from indoor environments. Results are also assessed for specific target species that may induce allergic reactions. Specific species are reviewed for their known potential to cause allergic reactions or as an indicator of potential water damage and moisture issues. In general, indoor mold levels should be equivalent to or lower than outdoor levels or non-complaint areas with similar types and percentages of mold species. Please note that airborne mold spores are present in most indoor environments at low levels.
- B. There are no regulations for acceptable levels of mold in the indoor environment; therefore, exact numerical limits are not supported at this time. Specific fungal species of concern are recommended to have much lower levels in the indoor environment. Existing conditions and symptoms are unique to each season, building and occupant. Airborne mold spore levels may vary greatly by location, time of day and weather conditions. However, health impacts cannot be predicted based on this information; individuals experience varying levels of allergic and non-allergic response to mold. Controlling moisture, in each form, is critical to the prevention of indoor mold growth issues.

8.0 Summary Conclusions

- A. The February 13, 2025 laboratory analysis results for sampling session #2 indicate that the airborne mold spore levels in the locations sampled were low and acceptable. The analyses did not identify any areas with elevated levels that would indicate an amplification of mold spores. The levels indicate a Level 1 condition (normal indoor fungal ecology) as defined by the Institute of Inspection, Cleaning, and Restoration Certification (IICRC).
- B. Relative humidity levels were generally within recommended levels (>30%) during the sampling period. These levels are typical for the winter season with dry conditions.

9.0 Limitations

- A. The assessment provided herein is based on the professional judgment of PMEC using approved industry standards and guidelines. Not all areas of the school building were accessed for inspection during the representative general spore trap air sampling. Assessment findings are based on the investigator's careful consideration of field observations and interpretation of analysis results in

accordance with industry standards, including, but not limited to, IICRC S520 guidelines for Condition 1 – normal fungal ecology, 2008 AIHA (Green Book) publication “Recognition, Evaluation, and Control of Indoor Mold”, and the ACGIH 1999 book “Bioaerosols – Assessment and Control”.

- B. The analysis results are only representative of the conditions of the date and time of sample collection and are considered a “snapshot in time”. PMEC’s results and discussion listed herein represent the conditions present at the time of inspection and sampling.

ATTACHMENTS

Attachment A – Sampling Floor Plan (1 page)

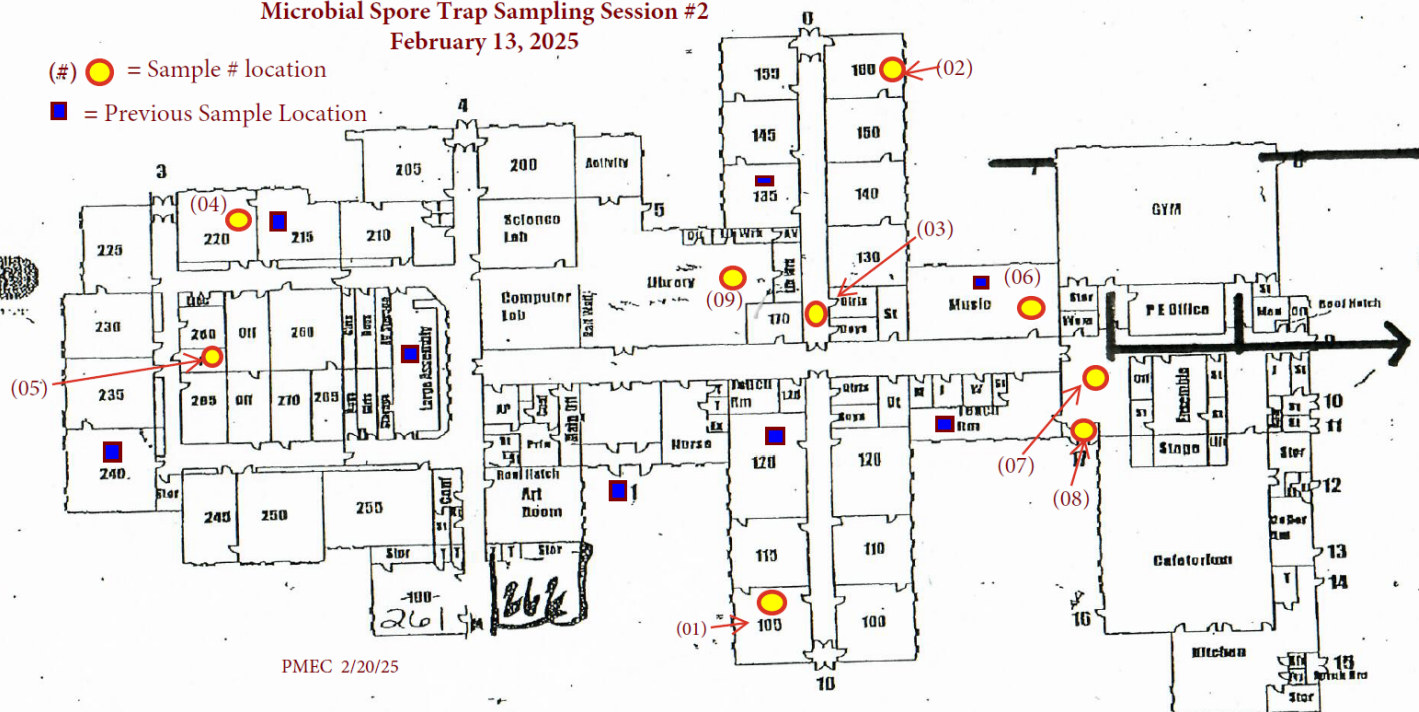
Attachment B – Hayes Microbial – February 13, 2025 Sample Analysis Results (pages 1-6)

Attachment A

Sample Location Floor Plan

Single Door
Window

(#) = Sample # location
 = Previous Sample Location



Attachment B

Laboratory Analysis Results

**Hayes Microbial –
February 20, 2025 Sampling Session #2
(pages 1-6)**

Analysis Report prepared for

Paul Matuszko Environmental Consulting

79 Cedar Street
Walpole, MA 02081

Phone: (617) 893-4476

25-114
Hobomock Elem. School
81 Learning Ln.
Pembroke, MA

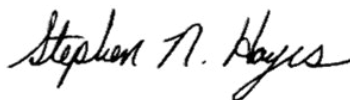
Collected: **February 13, 2025**
Received: **February 17, 2025**
Reported: **February 17, 2025**

We would like to thank you for trusting Hayes Microbial for your analytical needs!
We received 9 samples by FedEx in good condition for this project on February 17th, 2025.

The results in this analysis pertain only to this job, collected on the stated date, and should not be used in the interpretation of any other job. Information supplied by the customer can affect the validity of results. These results apply only to the samples as received. This report may not be duplicated, except in full, without the written consent of Hayes Microbial Consulting, LLC.

All information provided to Hayes Microbial is confidential information relating to our customers and their clients. We will not disclose, copy, or distribute any information verbally or written, except to those designated by the customer(s). We take confidentiality very seriously. No changes to the distribution list will be made without the express consent of the customer.

This laboratory bears no responsibility for sample collection activities, analytical method limitations, or your use of the test results. Interpretation and use of test results are your responsibility. Any reference to health effects or interpretation of mold levels is strictly the opinion of Hayes Microbial. In no event, shall Hayes Microbial or any of its employees be liable for lost profits or any special, incidental or consequential damages arising out of the use of these test results.



Steve Hayes, BSMT (ASCP)
Laboratory Director
Hayes Microbial Consulting, LLC.



EPA Laboratory ID: VA01419



Lab ID: #188863



DPH License: #PH-0198

Sample Number*	1	3961 2261		2	3961 2263		3	3961 2275		4	3961 2273	
Sample Name*	Classroom 105			Classroom 160			Hallway Near 130			Classroom 220		
Sample Volume*	75 L			75 L			75 L			75 L		
Reporting Limit	13 spores/m³			13 spores/m³			13 spores/m³			13 spores/m³		
Background	2			2			2			2		
Fragments	ND			ND			ND			ND		
Organism	Raw Count	Count / m³	% of Total	Raw Count	Count / m³	% of Total	Raw Count	Count / m³	% of Total	Raw Count	Count / m³	% of Total
Alternaria												
Ascospores	1	13	100.0%				1	13	100.0%	2	27	100.0%
Aspergillus Penicillium				10	130	90.9%						
Basidiospores												
Bipolaris Drechslera				1	13	9.1%						
Chaetomium												
Cladosporium												
Curvularia												
Epicoccum												
Fusarium												
Memnoniella												
Myxomycetes												
Pithomyces												
Stachybotrys												
Stemphylium												
Torula												
Ulocladium												
Total	1	13	100%	11	143	100%	1	13	100%	2	27	100%

Water Damage Indicator

Common Allergen

* indicates data provided by the customer



Sample Number*	5	3961 2259		6	3911 3613		7	3911 3649		8	3961 2262	
Sample Name*	SPED Room 275			Music Room			Gym Atrium			Gym Entrance Vestibule - Comparison		
Sample Volume*	75 L			75 L			75 L			75 L		
Reporting Limit	13 spores/m³			13 spores/m³			13 spores/m³			13 spores/m³		
Background	2			2			2			2		
Fragments	ND			ND			ND			ND		
Organism	Raw Count	Count / m³	% of Total	Raw Count	Count / m³	% of Total	Raw Count	Count / m³	% of Total	Raw Count	Count / m³	% of Total
Alternaria												
Ascospores	1	13	100.0%	1	13	50.0%	1	13	50.0%	2	27	100.0%
Aspergillus Penicillium												
Basidiospores							1	13	50.0%			
Bipolaris Drechslera												
Chaetomium												
Cladosporium				1	13	50.0%						
Curvularia												
Epicoccum												
Fusarium												
Memnoniella												
Myxomycetes												
Pithomyces												
Stachybotrys												
Stemphylium												
Torula												
Ulocladium												
Total	1	13	100%	2	26	100%	2	26	100%	2	27	100%

Water Damage Indicator

Common Allergen

* indicates data provided by the customer



Sample Number*	9	3961 2283							
Sample Name*	Library Right Side								
Sample Volume*	75 L								
Reporting Limit	13 spores/m³								
Background	2								
Fragments	ND								
Organism	Raw Count	Count / m³	% of Total						
Alternaria									
Ascospores	1	13	100.0%						
Aspergillus Penicillium									
Basidiospores									
Bipolaris Drechslera									
Chaetomium									
Cladosporium									
Curvularia									
Epicoccum									
Fusarium									
Memnoniella									
Myxomycetes									
Pithomyces									
Stachybotrys									
Stemphylium									
Torula									
Ulocladium									
Total	1	13	100%						

Water Damage Indicator

Common Allergen

* indicates data provided by the customer

Collected:Feb 13, 2025

Received: Feb 17, 2025

Reported: **Feb 17, 2025**



Project Analyst:
Ramesh Poluri, PhD

Date: **02 - 17 - 2025**

Reviewed By:
Steve Hayes, BSMT

Date: 02 - 17 - 2025

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Page: 4 of 6

Spore Trap Information

Reporting Limit	The Reporting Limit is the lowest number of spores that can be detected based on the total volume of the sample collected and the percentage of the slide that is counted. At Hayes Microbial, 100% of the slide is read so the LOD is based solely on the total volume. Raw spore counts that exceed 500 spores will be estimated.
Blanks	Results have not been corrected for field or laboratory blanks.
Background	The Background is the amount of debris that is present in the sample. This debris consists of skin cells, dirt, dust, pollen, drywall dust and other organic and non-organic matter. As the background density increases, the likelihood of spores, especially small spores such as those of <i>Aspergillus</i> and <i>Penicillium</i> may be obscured. The background is rated on a scale of 1 to 5 and each level is determined as follows: NBD: No background detected due to possible pump or cassette malfunction. Recollect sample. (Field Blanks will display NBD) 1 : <5% of field occluded. No spores will be uncountable. 2 : 5-25% of field occluded. 3 : 25-75% of field occluded. 4 : 75-90% of field occluded. 5 : >90% of field occluded. Suggested recollection of sample.
Fragments	Fragments are small pieces of fungal mycelium or spores. They are not identifiable as to type and when present in very large numbers, may indicate the presence of mold amplification.
Control Comparisons	There are no national standards for the numbers of fungal spores that may be present in the indoor environment. As a general rule and guideline that is widely accepted in the indoor air quality field, the numbers and types of spores that are present in the indoor environment should not exceed those that are present outdoors at any given time. There will always be some mold spores present in "normal" indoor environments. The purpose of sampling and counting spores is to help determine whether an abnormal condition exists within the indoor environment and if it does, to help pinpoint the area of contamination. Spore counts should not be used as the sole determining factor of mold contamination. There are many factors that can cause anomalies in the comparison of indoor and outdoor samples due to the dynamic nature of both of those environments.
Water Damage Indicator Common Allergen	Blue: These molds are commonly seen in conditions of prolonged water intrusion and usually indicate a problem. Green: Although all molds are potential allergens, these are the most common allergens that may be found indoors.
Color Coding	Fungi that are present in indoor samples at levels lower than 200 per cubic meter are not color coded on the report, unless they are one of the water damage indicators.
Significant Figures	Raw counts and column totals may reflect more than 2 significant figures, but results should only be considered significant to 2 figures.

Analyte Descriptions

Ascospores	Habitat:	A large group consisting of more than 3000 species of fungi. Common plant pathogens and outdoor numbers become very high following rain. Most of the genera are indistinguishable by spore trap analysis and are combined on the report.
	Health Effects:	Health affects are poorly studied, but many are likely to be allergenic.
Aspergillus Penicillium	Habitat:	The most common fungi isolated from the environment. Very common in soil and on decaying plant material. Are able to grow well indoors on a wide variety of substrates.
	Health Effects:	This group contains common allergens and many can cause hypersensitivity pneumonitis. They may cause extrinsic asthma, and many are opportunistic pathogens. Many species produce mycotoxins which may be associated with disease in humans and other animals. Toxin production is dependent on the species, the food source, competition with other organisms, and other environmental conditions.
Basidiospores	Habitat:	A common group of Fungi that includes the mushrooms and bracket fungi. They are saprophytes and plant pathogens. In wet conditions they can cause structural damage to buildings.
	Health Effects:	Common allergens and are also associated with hypersensitivity pneumonitis.
Bipolaris Drechslera	Habitat:	They are found in soil and as plant pathogens. Can grow indoors on a variety of substrates.
	Health Effects:	They may be allergenic and are very commonly involved in allergic fungal sinusitis. They are opportunistic pathogens but occasionally infect healthy individuals, causing keratitis, sinusitis and osteomyelitis.
Cladosporium	Habitat:	One of the most common genera worldwide. Found in soil and plant debris and on the leaf surfaces of living plants. The outdoor numbers are lower in the winter and often relatively high in the summer, especially in high humidity. The outdoor numbers often spike in the late afternoon and evening. Indoors, it can be found growing on textiles, wood, sheetrock, moist window sills and in HVAC supply ducts.
	Health Effects:	A common allergen, producing more than 10 allergenic antigens and a common cause of hypersensitivity pneumonitis.



Paul Matuszko Environmental Consulting
79 Cedar Street
Walpole, MA 02081

N

SHIP: FEDEX - ENV 50
DATE: 02-17-2025



25007143

Pembroke Public Schools

Job Number: 25-114		Job Name: Hobomock Elem School		Mobile: 6178934476		Email: pmatuszko@pmecsolutions.	
Collector: Paul Matuszko		81 Learning Lanes		Note: <i>pos. rain, 39°F, 70%, Background air sampling</i>			
Date Collected: 2/13/25		Pembroke, MA					
Analysis Type		Analysis Description		Turnaround		Accepted Media Types	
Spore Trap	S	Identification & Enumeration of Fungal Spores		24 Hour	Air Cassettes, Impact Slides		
	S+	Spore Trap Analysis with Dander, Fiber, and Pollen counts		24 Hour	Air Cassettes, Impact Slides		
Direct ID	D	ID & Semi-Quantative Enumeration of spores and mycelium		24 Hour	Bio-Tape, Tape, Swab, Bulk, Agar Plate		
	D+	Direct Analysis with Fully Quantitative spore count		24 Hour	Bio-Tape, Tape, Swab, Bulk, Agar Plate		
Culture	C1	Identification & Enumeration of Mold only		7 Day	Air Plate, Agar Plate, Swab, Bulk		
	C2	Identification & Enumeration of Bacteria only		4 Day	Air Plate, Agar Plate, Swab, Bulk		
	C3	Identification & Enumeration of Mold and Bacteria		7 Day	Air Plate, Agar Plate, Swab, Bulk		
	C5	Coliform Screen for Sewage Bacteria		2 Day	Agar Plate, Swab, Bulk		
Particle	TPA	Total Particulate Analysis, ID & Count (Does Not Include Mold)		24 Hour	Air Cassettes, Impact Slides, Bio-Tape		
#	Number	Sample	Analysis	Volume	Notes		
1	3961 2261	Classroom 105	S	75 L	5 min @ 15 LPM		
2	3961 2263	Classroom 160					
3	3961 2275	Hallway near 130					
4	3961 2273	Classroom 220					
5	3961 2259	SPEO Room #275			Carpet		
6	3911 3613	Music Room					
7	3911 3649	Gym Atrium					
8	3961 2262	Gym Entrance vestibule - Comparison			SNOW COVER ON GROUND		
9	3961 2283	Library - Right side					
10							
11							
12							
13							
14							
15							
16							
Released by: <i>Paul Matuszko</i>		Date: 2/14/25		Received By: <i>Am</i>		Date: 2/17	