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Quarterly Microbial IAQ Assessment Report

Conducted At

**Hobomock Elementary School
81 Learning Lanes
Pembroke, MA 02359**

November, 2024

Prepared For:

Ms. Erin Obey
Superintendent of Schools
Pembroke Public Schools
72 Pilgrim Road
Pembroke, MA 02339

Report Prepared By:

A handwritten signature in black ink that reads "Paul Matuszko".

Paul Matuszko, CIH
Paul Matuszko Environmental Consulting
79 Cedar Street
Walpole, MA 02081

Report Date:

November 26, 2024

PMEC Project #:

24-163.1



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November 26, 2024

Ms. Erin Obey
Superintendent of Schools
Pembroke Public Schools
72 Pilgrim Road
Pembroke, MA 02339

RE: Quarterly Microbial (Mold) IAQ Sampling Report #1 for
Hobomock Elementary School
81 Learning Lane, Pembroke, MA
PMEC Project #24-163.1

Dear Ms. Obey:

Paul Matuszko Environmental Consulting (**PMEC**) is pleased to submit the enclosed report for the quarterly microbial (mold) indoor air quality (IAQ) sampling conducted at the Hobomock Elementary School, 81 Learning Lane, Pembroke, Massachusetts. PMEC conducted the sampling within the school on November 12, 2024. Eight (8) total air samples were collected for microbial laboratory analysis.

This limited microbial assessment report has been prepared for the exclusive use of The Pembroke Public Schools.

Certification:

PMEC certifies that the results and findings provided herein for the Hobomock Elementary School building have been reviewed for accuracy, content, regulatory compliance and quality of presentation.

Should you have any questions regarding this report, please do not hesitate to contact me at (617) 893-4476. Thank you for providing PMEC with the opportunity to provide our services to the Pembroke Public Schools for this project.

Respectively submitted;

A handwritten signature in black ink that reads "Paul Matuszko".

Paul Matuszko, CIH, CIEC
Project Manager
CIH - Certified Industrial Hygienist (American Board of Industrial Hygiene #9236 CP)
CIEC - Certified Indoor Environmental Consultant (American Council for Accredited Certification)





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Attachment B – Hayes Microbial Laboratory Analytical Results (pages 1-6)



**Quarterly Microbial IAQ
Sampling Report #1
For
Hobomock Elementary School
81 Learning Lane
Pembroke, MA**

November, 2024

1.0 Introduction

- A. Paul Matuszko Environmental Consulting (*PMEC*) is pleased to submit this report for the quarterly microbial sampling assessment (#1) conducted at the Hobomock Elementary School, 81 Learning Lane, Pembroke, Massachusetts. *PMEC* was retained by the Pembroke Public Schools to conduct the general air sampling and a general inspection for water and microbial growth within the school building. The microbial spore trap air sampling was conducted by *PMEC* on November 12, 2024. The air sampling was conducted to determine existing spore levels in compliance with the on-going School system IAQ program. A summary of the sampling locations and methods, analysis methods and results are outlined within the report.
- B. *PMEC* was assisted by Pembroke Public Schools Superintendent and maintenance/custodial personnel during the air sampling. Member(s) of the Pembroke Schools Teachers Union were present during the sampling to confirm the locations and sampling methods used. The sampling was conducted to determine existing airborne microbial spore levels in representative areas of each section or wing of the school building.
- C. During sampling, *PMEC* conducted a general visual inspection of representative room surfaces. Additionally, measurements baseline indoor temperature and humidity levels were collected. *PMEC* noted any areas or locations with atypical conditions. The following report summarizes the findings of the assessment, analysis results, and general recommendations.

2.0 Scope of Work

- A. The indoor air quality (IAQ) assessment was conducted in accordance with the following tasks:
 1. General inspection of the representative areas for visible water damaged materials and mold growth in the rooms being sampled.
 2. Conduct representative bioaerosol (microbial) spore trap air sampling for airborne mold (fungal) analysis in designated and representative locations. Seven (7) interior and one (1) exterior/outdoor air samples were collected during the November 12, 2024 after school hours site visit (~3-5 pm).
 3. Prepare a detailed report detailing the findings of the assessment and sampling data.

3.0 Existing Conditions

- A. General Observations
 - Ceiling vents (supply diffusers and exhaust vents) were observed to be clean with no accumulated dust or suspect mold spotting present.
 - Various HEPA filtered air scrubber fan units were operating in miscellaneous classrooms of the school.

- Classrooms horizontal surfaces were generally clean and free of accumulated dust and debris. Ceiling tiles and HVAC ceiling vents were clean and free of excessive water stains and debris.

4.0 Air Quality Measurements

A. A direct reading, thermohygrometer (Fluke Model 971) was used to collect representative temperature and relative humidity (ratio of water vapor in air) measurements. Recommended season levels are:

- Temperature (Temp.): Recommended Winter Range: 63-72 °F
 (degrees Fahrenheit °F) Recommended Summer: 68-78 °F
- Relative Humidity (RH%) Recommended Range: between 30%-60%; (<50% goal)

B. The measurement results on the assessment date are provided in Table 1 below:

Table 1 Baseline Air Quality Measurements			
Location	Temperature	Relative Humidity %	Comments
LIR assembly room	70.4 °F	34.3%	Slightly low RH% level
Classroom #240	66.7 °F	29.3%	Slightly low RH% level, open window
Classroom #215	68.7 °F	28.3%	Slightly low RH% level
Classroom #125	70.5 °F	28.1%	Slightly low RH% level
Classroom #135	70.0 °F	27.0%	HEPA fan on, Slightly low RH% level
Music Room	70.6 °F	27.3%	Slightly low RH% level, carpet removed
Faculty dining room	68.3 °F	27.2%	Slightly low RH% level, carpet removed
Outside ambient air (outside front entrance)	44.6 °F	49.2%	Cool, clear, dry, typical late fall day

C. Discussion

1. Temperature were typical for a school building during the late fall season. Relative humidity levels were at or slightly lower than recommended levels. However, this is typical of occupied buildings during a dry and cool late fall season day.

5.0 Mold Sampling and Analysis Methods

A. PMEC collected “spore trap” air samples for mold spore analysis using air-o-cell® brand sampling cassettes. Seven (7) indoor samples and one (1) outdoor comparison sample were collected for analysis. A calibrated, battery operated Zefon IAQ 15 sampling pump was used to draw air onto the sample cassette’s adhesive slide. PMEC collected the samples at 15 liters of air per minute (LPM) for a five (5) minute sample duration for a total sample collection volume of 75 liters of air. The airborne aerosols (mold, particulates, pollen, etc.) are trapped on the filter media slide for direct microscopic examination.

B. The samples were collected on November 12, 2024 at approximately 4:00 pm – 5:15 pm. Ambient conditions were cool (~46° F) and dry. Indoor conditions were typical of the season.

- C. The samples were sent via chain of custody by Fed Ex to Hayes Microbial Laboratory (Hayes), located in Midlothian, Virginia. Hayes Laboratory is accredited by the American Industrial Hygiene Association (AIHA) for mold and bacteria identification and analysis (AIHA EMPAT Laboratory Accreditation ID # 188863).
- D. Results are reported as Total Fungi Counts in spores per cubic meter of air (**C/m³**). The samples were analyzed for both non-viable and viable fungi by direct analysis optical microscopy. A summary of analysis criteria of spore trap and direct identification analysis is provided in the lab analysis sheets. The sample results are provided as Attachment B to this report.

6.0 Laboratory Analysis Results

- A. The results of the November 12, 2024 spore trap air sampling are presented in Table 2 below.

Table 2 Spore Trap Air Sample Analysis Results November 12, 2024 Quarterly IAQ Sampling Session #1				
Sample #	Sample Location	Total Fungi (C/m ³)	Specific Species & levels of note	Comments
3782 1665 (01)	LIR Assembly Room	39 C/m ³	Ascospores – 13 C/m ³ Basidiospores – 13 C/m ³ Myxomycetes – 13 C/m ³	Low and acceptable
3782 1612 (02)	Classroom #240	80 C/m ³	Ascospores – 40 C/m ³ Basidiospores – 13 C/m ³ Myxomycetes – 27 C/m ³	Low and acceptable
3782 1670 (03)	Classroom #215	13 C/m ³	Cladosporium – 13 C/m ³	Low and acceptable
3782 1589 (04)	Classroom #125	40 C/m ³	Ascospores – 27 C/m ³ Curvularia – 13 C/m ³	Low and acceptable
3782 1587 (05)	Classroom #135	53 C/m ³	Ascospores – 13 C/m ³ Basidiospores – 27 C/m ³ Curvularia – 13 C/m ³	Low and acceptable
3782 1637 (06)	Music Room	26 C/m ³	Ascospores – 13 C/m ³ Rusts/Smuts – 13 C/m ³	Low and acceptable
3782 1601 (07)	Faculty Dining Room	93 C/m ³	Ascospores – 53 C/m ³ Cladosporium – 27 C/m ³ Myxomycetes – 13 C/m ³	Low and acceptable
3782 1593 (08)	Outside ambient air – (Outside main entrance)	226 C/m ³	Ascospores – 53 C/m ³ Basidiospores – 40 C/m ³ Cladosporium – 93 C/m ³ Myxomycetes – 13 C/m ³ Rusts/Smuts – 27 C/m ³	Typical fall season levels

- Notes:**
- Additional information on species types are provided in the Laboratory Analysis results.
 - C/m³ = spore counts per cubic meter of air
 - Samples collected in middle or back middle of room unless noted
 - Results in **Bold** = species or levels of concern.

7.0 Discussion of Analysis Results

- A. The analysis results for the November 12, 2024 sampling session indicate very low and acceptable airborne spore levels. The species identified in these rooms are typically found indoors at low levels and are not a cause for concern.

- B. The indoor sample species identified are similar to the species present on the outdoor ambient air sample analysis. The current airborne spore levels on the sampling date are not representative of an amplified airborne spore condition in the locations sampled.
- C. No spores of aspergillus/penicillium or other water damage indicator species were identified on any of the sample results.

8.0 Background Mold Information

- A. Currently, there are no standards or regulations to indicate acceptable numerical levels of airborne fungal spores derived from indoor environments. Results are also assessed for specific target species that may induce allergic reactions. Specific species are reviewed for their known potential to cause allergic reactions or as an indicator of potential water damage and moisture issues. In general, indoor mold levels should be equivalent to or lower than outdoor levels or non-complaint areas with similar types and percentages of mold species. Please note that airborne mold spores are present in most indoor environments at low levels.
- B. There are no regulations for acceptable levels of mold in the indoor environment; therefore, exact numerical limits are not supported at this time. Specific fungal species of concern are recommended to have much lower levels in the indoor environment. Existing conditions and symptoms are unique to each season, building and occupant. Airborne mold spore levels may vary greatly by location, time of day and weather conditions. However, health impacts cannot be predicted based on this information; individuals experience varying levels of allergic and non-allergic response to mold. Controlling moisture, in each form, is critical to the prevention of indoor mold growth issues.

9.0 Summary Conclusions

- A. The November 12, 2024 laboratory analysis results indicate that the airborne mold spore levels in the locations sampled were low and acceptable.
- B. Relative humidity levels were slightly lower than recommended during the sampling period. However, this is typical for the season with dry conditions.
- C. PMEC did not identify any accessible, visible conditions within the areas sampled that would directly impact IAQ.

10.0 Limitations

- A. The assessment provided herein is based on the professional judgment of PMEC using approved industry standards and guidelines. Not all areas of the school building were accessed for inspection during the representative general spore trap air sampling. Assessment findings are based on the investigator's careful consideration of field observations and interpretation of analysis results in accordance with industry standards, including, but not limited to, IICRC S520 guidelines for Condition 1 – normal fungal ecology, 2008 AIHA (Green Book) publication "Recognition, Evaluation, and Control of Indoor Mold", and the ACGIH 1999 book "Bioaerosols – Assessment and Control".
- B. The analysis results are only representative of the conditions of the date and time of sample collection and are considered a "snapshot in time". PMEC's results and discussion listed herein represent the conditions present at the time of inspection and sampling.

ATTACHMENTS

Attachment A – Sampling Floor Plan (1 page)

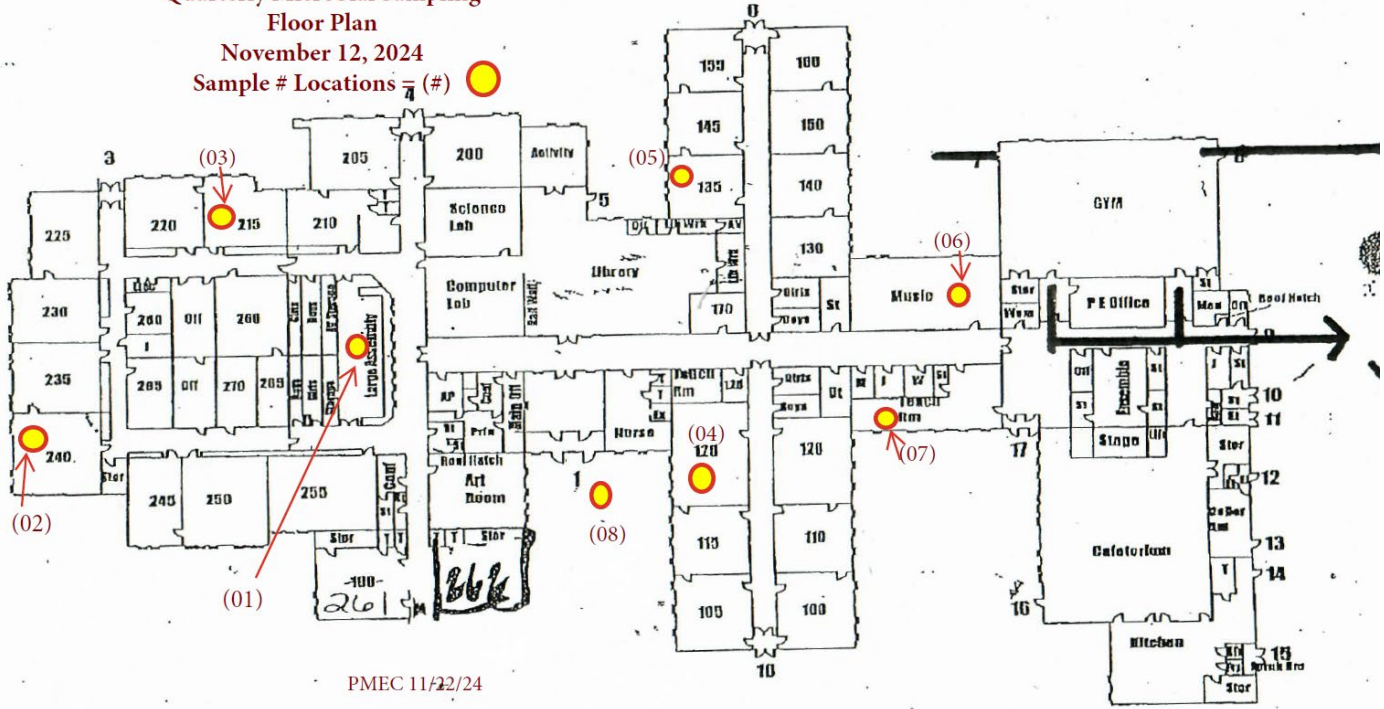
Attachment B – Hayes Microbial – November 12, 2024 Sample Analysis Results (pages 1-6)

Attachment A

Sample Location Floor Plan

Attachment A – Sampling Floor Plan

Hobomock Elementary School
Quarterly Microbial Sampling
Floor Plan
November 12, 2024
Sample # Locations = (#)



PMEC 11/22/24

Attachment B

Laboratory Analysis Results

**Hayes Microbial –
November 12, 2024 Sampling
(pages 1-6)**



#24051576

Analysis Report prepared for

Paul Matuszko Environmental Consulting

79 Cedar Street
Walpole, MA 02081

Phone: (617) 893-4476

241-163
Hobomock Elem. School
81 Learning Ln.
Pembroke, MA 02359

Collected: **November 12, 2024**
Received: **November 14, 2024**
Reported: **November 14, 2024**



EPA Laboratory ID: VA01419



Lab ID: #188863



DPH License: #PH-0198

We would like to thank you for trusting Hayes Microbial for your analytical needs!
We received 8 samples by FedEx in good condition for this project on November 14th, 2024.

The results in this analysis pertain only to this job, collected on the stated date, and should not be used in the interpretation of any other job. Information supplied by the customer can affect the validity of results. These results apply only to the samples as received. This report may not be duplicated, except in full, without the written consent of Hayes Microbial Consulting, LLC.

All information provided to Hayes Microbial is confidential information relating to our customers and their clients. We will not disclose, copy, or distribute any information verbally or written, except to those designated by the customer(s). We take confidentiality very seriously. No changes to the distribution list will be made without the express consent of the customer.

This laboratory bears no responsibility for sample collection activities, analytical method limitations, or your use of the test results. Interpretation and use of test results are your responsibility. Any reference to health effects or interpretation of mold levels is strictly the opinion of Hayes Microbial. In no event, shall Hayes Microbial or any of its employees be liable for lost profits or any special, incidental or consequential damages arising out of the use of these test results.

A handwritten signature in black ink that reads 'Stephen N. Hayes'.

Steve Hayes, BSMT (ASCP)
Laboratory Director
Hayes Microbial Consulting, LLC.

Sample Number*	1			2			3			4		
Sample Name*	LIR - Assembly Room			Classroom 240			Classroom 215			Classroom 125		
Sample Volume*	75 L			75 L			75 L			75 L		
Reporting Limit	13 spores/m ³			13 spores/m ³			13 spores/m ³			13 spores/m ³		
Background	2			2			2			2		
Fragments	ND			13/m ³			ND			ND		
Organism	Raw Count	Count / m ³	% of Total	Raw Count	Count / m ³	% of Total	Raw Count	Count / m ³	% of Total	Raw Count	Count / m ³	% of Total
Alternaria												
Ascospores	1	13	33.3%	3	40	50.0%				2	27	66.7%
Aspergillus Penicillium												
Basidiospores	1	13	33.3%	1	13	16.7%						
Bipolaris Drechslera												
Chaetomium												
Cladosporium							1	13	100.0%			
Curvularia										1	13	33.3%
Epicoccum												
Fusarium												
Memnoniella												
Myxomycetes	1	13	33.3%	2	27	33.3%						
Pithomyces												
Rusts/Smuts												
Stachybotrys												
Stemphylium												
Torula												
Ulocladium												
Total	3	39	100%	6	80	100%	1	13	100%	3	40	100%

Water Damage Indicator	Common Allergen	Slightly Higher than Baseline	Significantly Higher than Baseline	Ratio Abnormality
------------------------	-----------------	-------------------------------	------------------------------------	-------------------

* indicates data provided by the customer



Collected: **Nov 12, 2024**

Received: **Nov 14, 2024**

Reported: **Nov 14, 2024**

Project Analyst:
 Joseph Lape, *Joseph Lape*

Date:
11 - 14 - 2024

Reviewed By:
 Steve Hayes, BSMT *Stephen N. Hayes*

Date:
11 - 14 - 2024

Sample Number*	5			6			7			8		
Sample Name*	Classroom 135			Music Room			Faculty Dining Room			Outside Ambient Air		
Sample Volume*	75 L			75 L			75 L			75 L		
Reporting Limit	13 spores/m ³			13 spores/m ³			13 spores/m ³			13 spores/m ³		
Background	2			2			3			2		
Fragments	ND			13/m ³			ND			27/m ³		
Organism	Raw Count	Count / m ³	% of Total	Raw Count	Count / m ³	% of Total	Raw Count	Count / m ³	% of Total	Raw Count	Count / m ³	% of Total
Alternaria												
Ascospores	1	13	25.0%	1	13	50.0%	4	53	57.1%	4	53	23.5%
Aspergillus Penicillium												
Basidiospores	2	27	50.0%							3	40	17.6%
Bipolaris Drechslera												
Chaetomium												
Cladosporium							2	27	28.6%	7	93	41.2%
Curvularia	1	13	25.0%									
Epicoccum												
Fusarium												
Memnoniella												
Myxomycetes							1	13	14.3%	1	13	5.9%
Pithomyces												
Rusts/Smuts				1	13	50.0%				2	27	11.8%
Stachybotrys												
Stemphylium												
Torula												
Ulocladium												
Total	4	53	100%	2	26	100%	7	93	100%	17	226	100%

Water Damage Indicator	Common Allergen	Slightly Higher than Baseline	Significantly Higher than Baseline	Ratio Abnormality
------------------------	-----------------	-------------------------------	------------------------------------	-------------------

* indicates data provided by the customer



Collected: **Nov 12, 2024**

Received: **Nov 14, 2024**

Reported: **Nov 14, 2024**

Project Analyst:
 Joseph Lape, *Joseph Lape*

Date:
11 - 14 - 2024

Reviewed By:
 Steve Hayes, BSMT *Stephen N. Hayes*

Date:
11 - 14 - 2024

Spore Trap Information

Reporting Limit	The Reporting Limit is the lowest number of spores that can be detected based on the total volume of the sample collected and the percentage of the slide that is counted. At Hayes Microbial, 100% of the slide is read so the LOD is based solely on the total volume. Raw spore counts that exceed 500 spores will be estimated.					
Blanks	Results have not been corrected for field or laboratory blanks.					
Background	<p>The Background is the amount of debris that is present in the sample. This debris consists of skin cells, dirt, dust, pollen, drywall dust and other organic and non-organic matter. As the background density increases, the likelihood of spores, especially small spores such as those of <i>Aspergillus</i> and <i>Penicillium</i> may be obscured. The background is rated on a scale of 1 to 5 and each level is determined as follows:</p> <p>NBD: No background detected due to possible pump or cassette malfunction. Recollect sample. (Field Blanks will display NBD)</p> <p>1 : <5% of field occluded. No spores will be uncountable.</p> <p>2 : 5-25% of field occluded.</p> <p>3 : 25-75% of field occluded.</p> <p>4 : 75-90% of field occluded.</p> <p>5 : >90% of field occluded. Suggested recollection of sample.</p>					
Fragments	Fragments are small pieces of fungal mycelium or spores. They are not identifiable as to type and when present in very large numbers, may indicate the presence of mold amplification.					
Control Comparisons	There are no national standards for the numbers of fungal spores that may be present in the indoor environment. As a general rule and guideline that is widely accepted in the indoor air quality field, the numbers and types of spores that are present in the indoor environment should not exceed those that are present outdoors at any given time. There will always be some mold spores present in "normal" indoor environments. The purpose of sampling and counting spores is to help determine whether an abnormal condition exists within the indoor environment and if it does, to help pinpoint the area of contamination. Spore counts should not be used as the sole determining factor of mold contamination. There are many factors that can cause anomalies in the comparison of indoor and outdoor samples due to the dynamic nature of both of those environments.					
<table border="1"> <tr><td>Water Damage Indicator</td></tr> <tr><td>Common Allergen</td></tr> <tr><td>Slightly Higher than Baseline</td></tr> <tr><td>Significantly Higher than Baseline</td></tr> <tr><td>Ratio Abnormality</td></tr> </table>	Water Damage Indicator	Common Allergen	Slightly Higher than Baseline	Significantly Higher than Baseline	Ratio Abnormality	<p>Blue: These molds are commonly seen in conditions of prolonged water intrusion and usually indicate a problem.</p> <p>Green: Although all molds are potential allergens, these are the most common allergens that may be found indoors.</p> <p>Orange: The spore count is slightly higher than the outside count and may or may not indicate a source of contamination.</p> <p>Red: The spore count is significantly higher than the baseline count and probably indicates a source of contamination.</p> <p>Violet: The types of spores found indoors should be similar to the ones that were identified in the baseline sample. Significant increases (more than 25%) in the ratio of a particular spore type may indicate the presence of abnormal levels of mold, even if the total number of spores of that type is lower in the indoor environment than it was outdoors.</p>
Water Damage Indicator						
Common Allergen						
Slightly Higher than Baseline						
Significantly Higher than Baseline						
Ratio Abnormality						
Color Coding	Fungi that are present in indoor samples at levels lower than 200 per cubic meter are not color coded on the report, unless they are one of the water damage indicators.					
Significant Figures	Raw counts and column totals may reflect more than 2 significant figures, but results should only be considered significant to 2 figures.					

Ascospores	Habitat:	A large group consisting of more than 3000 species of fungi. Common plant pathogens and outdoor numbers become very high following rain. Most of the genera are indistinguishable by spore trap analysis and are combined on the report.
	Health Effects:	Health affects are poorly studied, but many are likely to be allergenic.

Basidiospores	Habitat:	A common group of Fungi that includes the mushrooms and bracket fungi. They are saprophytes and plant pathogens. In wet conditions they can cause structural damage to buildings.
	Health Effects:	Common allergens and are also associated with hypersensitivity pneumonitis.

Cladosporium	Habitat:	One of the most common genera worldwide. Found in soil and plant debris and on the leaf surfaces of living plants. The outdoor numbers are lower in the winter and often relatively high in the summer, especially in high humidity. The outdoor numbers often spike in the late afternoon and evening. Indoors, it can be found growing on textiles, wood, sheetrock, moist window sills and in HVAC supply ducts.
	Health Effects:	A common allergen, producing more than 10 allergenic antigens and a common cause of hypersensitivity pneumonitis.

Curvularia	Habitat:	They exist in soil and plant debris, and are plant pathogens.
	Health Effects:	They are allergenic and a common cause of allergic fungal sinusitis. An occasional cause of human infection, including keratitis, sinusitis, onychomycosis, mycetoma, pneumonia, endocarditis and disseminated infection, primarily in the immunocompromised.

Myxomycetes	Habitat:	Found on decaying plant material and as a plant pathogen.
	Health Effects:	Some allergenic properties reported, but generally pose no health concerns to humans.

Rusts/Smuts	Habitat:	Found on decaying plant material and as a plant pathogen.
	Health Effects:	Some allergenic properties reported.



Paul Matuszko Environmental Consulting
 79 Cedar Street
 Walpole, MA 02081

N

SHIP: FEDEX - PAK 50
 DATE: 11-14-2024



Job Number: 24-163	Job Name: <i>Hobomock Elem. School</i> <i>81 Learning Lane</i> Pembroke, MA 02359	Mobile: 6178934476	Email: pmatuszko@pmecsolution
Collector: Paul Matuszko		Note: <i>clear, cool NSF 30% AH</i>	
Date Collected: <i>11/12/24</i>			

Analysis Type	Analysis Description	Turnaround	Accepted Media Types
Spore Trap	S Identification & Enumeration of Fungal Spores	24 Hour	Air Cassettes, Impact Slides
	S+ Spore Trap Analysis with Dander, Fiber, and Pollen counts	24 Hour	Air Cassettes, Impact Slides
Direct ID	D ID & Semi-Quantative Enumeration of spores and mycelium	24 Hour	Bio-Tape, Tape, Swab, Bulk, Agar Plate
	D+ Direct Analysis with Fully Quantitative spore count	24 Hour	Bio-Tape, Tape, Swab, Bulk, Agar Plate
Culture	C1 Identification & Enumeration of Mold only	7 Day	Air Plate, Agar Plate, Swab, Bulk
	C2 Identification & Enumeration of Bacteria only	4 Day	Air Plate, Agar Plate, Swab, Bulk
	C3 Identification & Enumeration of Mold and Bacteria	7 Day	Air Plate, Agar Plate, Swab, Bulk
	C5 Coliform Screen for Sewage Bacteria	2 Day	Agar Plate, Swab, Bulk
Particle	TPA Total Particulate Analysis, ID & Count (Does Not Include Mold)	24 Hour	Air Cassettes, Impact Slides, Bio-Tape

#	Number	Sample	Analysis	Volume	Notes
1	<i>3782 1665</i>	<i>LRA- Assembly Room</i>	<i>S</i>	<i>75 L</i>	<i>5min @ 15:00 PM - Background sampling</i>
2	<i>3782 1612</i>	<i>classroom 240</i>	<i>S</i>		
3	<i>3782 1677</i>	<i>classroom 215</i>	<i>S</i>		
4	<i>3782 1589</i>	<i>classroom 125</i>	<i>S</i>		
5	<i>3782 1587</i>	<i>classroom 135</i>	<i>S</i>		
6	<i>3782 1637</i>	<i>Music Room</i>	<i>S</i>		
7	<i>3782 1601</i>	<i>Faculty Dining Room</i>	<i>S</i>		
8	<i>3782 1593</i>	<i>outside Ambient Air</i>	<i>S</i>		
9					
10					
11					
12					
13					
14					
15					
16					

Released by: <i>Paul Matuszko</i>	Date: <i>11/13/24</i>	Received By: <i>Ann</i>	Date: <i>11/14</i>
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